

# Package ‘GCalignR’

July 3, 2024

**Title** Simple Peak Alignment for Gas-Chromatography Data

**Version** 1.0.7

**Date** 2024-07-03

**Encoding** UTF-8

**Description** Aligns peak based on peak retention times and matches homologous peaks across samples. The underlying alignment procedure comprises three sequential steps. (1) Full alignment of samples by linear transformation of retention times to maximise similarity among homologous peaks (2) Partial alignment of peaks within a user-defined retention time window to cluster homologous peaks (3) Merging rows that are likely representing homologous substances (i.e. no sample shows peaks in both rows and the rows have similar retention time means). The algorithm is described in detail in Ottensmann et al., 2018 <[doi:10.1371/journal.pone.0198311](https://doi.org/10.1371/journal.pone.0198311)>.

**Depends** R (>= 3.2.5)

**Imports** ggplot2 (>= 2.2.1), graphics, stats, readr, reshape2, stringr, utils, pbapply, methods, tibble

**License** GPL (>= 2) | file LICENSE

**Language** en-GB

**LazyData** true

**RoxygenNote** 7.3.2

**Suggests** knitr, pander, rmarkdown, testthat, vegan (>= 2.4.2)

**VignetteBuilder** knitr

**BugReports** <https://github.com/mottensmann/GCalignR/issues>

**URL** <https://github.com/mottensmann/GCalignR>

**NeedsCompilation** no

**Author** Meinolf Ottensmann [aut, cre] (<<https://orcid.org/0000-0003-3112-6928>>),  
Martin Stoffel [aut],  
Hazel J. Nichols [aut],  
Joseph I. Hoffman [aut]

**Maintainer** Meinolf Ottensmann <[meinolf.ottensmann@web.de](mailto:meinolf.ottensmann@web.de)>

**Repository** CRAN

**Date/Publication** 2024-07-03 18:00:01 UTC

## Contents

aligned_peak_data	2
align_chromatograms	3
as.data.frame.GCalign	6
blank_substraction	7
check_input	8
choose_optimal_reference	9
draw_chromatogram	10
find_peaks	12
GCalignR	13
gc_heatmap	14
linear_transformation	16
merge_redundant_rows	18
norm_peaks	19
peak_data	20
peak_factors	20
peak_interspace	21
plot.GCalign	22
print.GCalign	23
read_empower2	24
read_peak_list	25
remove_blanks	25
remove_singletons	26
simple_chroma	27
<b>Index</b>	<b>29</b>

---

aligned_peak_data	<i>Aligned Gas-Chromatography data</i>
-------------------	--

---

### Description

This is an example of an aligned gas-chromatography dataset processed with [align\\_chromatograms](#). The raw data is accessible within this package as **peak\_data.RData** and is comprised of 41 Mother-Pup pairs of Antarctic Fur Seals (*Arctocephalus gazella*) sampled from two different colonies at Bird Island, South Georgia. In addition two blanks are included.

### Format

Object of class "GCalign" including three lists. List "**aligned**" includes data.frames for all variables present in the raw data ("time" and "area"). The list "**heatmap\_input**" holds data frames with retention times of the input data, linearly adjusted retention times as well as the final output, where peaks are aligned among samples. This file is primarily used in [gc\\_heatmap](#). The list "**Logfile**" summarises the alignment process and the data structure before, during and after running [align\\_chromatograms](#). For a convenient overview use [print.GCalign](#).

## Source

<http://www.pnas.org/content/suppl/2015/08/05/1506076112.DCSupplemental/pnas.1506076112.sd02.xlsx>

## References

Stoffel, M.A.; Caspers, B.A.; Forcada, J.; Giannakara, A.; Baier, M.; Eberhart-Phillips, L.; Mueller, C.; Hoffman, J.I. (2015): Chemical fingerprints encode mother-offspring similarity, colony membership, relatedness, and genetic quality in fur seals. In: Proceedings of the National Academy of Sciences of the United States of America 112 (36), S. E5005-12. DOI: 10.1073/pnas.1506076112.

---

align\_chromatograms     *Aligning peaks based on retention times*

---

## Description

This is the core function of [GCalignR](#) to align peak data. The input data is a peak list. Read through the documentation below and take a look at the vignettes for a thorough introduction. Three parameters `max_linear_shift`, `max_diff_peak2mean` and `min_diff_peak2peak` are required as well as the column name of the peak retention time variable `rt_col_name`. Arguments are described among optional parameters below.

## Usage

```
align_chromatograms(  
  data,  
  sep = "\t",  
  rt_col_name = NULL,  
  write_output = NULL,  
  rt_cutoff_low = NULL,  
  rt_cutoff_high = NULL,  
  reference = NULL,  
  max_linear_shift = 0.02,  
  max_diff_peak2mean = 0.02,  
  min_diff_peak2peak = 0.08,  
  blanks = NULL,  
  delete_single_peak = FALSE,  
  remove_empty = FALSE,  
  permute = TRUE,  
  ...  
)
```

## Arguments

`data`             Dataset containing peaks that need to be aligned and matched. For every peak a arbitrary number of numerical variables can be included (e.g. peak height, peak area) in addition to the mandatory retention time. The standard format is a tab-delimited text file according to the following layout: (1) The first row

contains sample names, the (2) second row column names of the corresponding peak lists. Starting with the third row, peak lists are included for every sample that needs to be incorporated in the dataset. Here, a peak list contains data for individual peaks in rows, whereas columns specify variables in the order given in the second row of the text file. Peak lists of individual samples are concatenated horizontally and need to be of the same width (i.e. the same number of columns in consistent order). Alternatively, the input may be a list of data frames. Each data frame contains the peak data for a single individual. Variables (i.e. columns) are named consistently across data frames. The names of elements in the list are used as sample identifiers. Cells may be filled with numeric or integer values but no factors or characters are allowed. NA and 0 may be used to indicate empty rows.

sep	The field separator character. The default is tab separated (sep = '\t'). See the "sep" argument in <a href="#">read.table</a> for details.
rt_col_name	A character giving the name of the column containing the retention times. The decimal separator needs to be a point.
write_output	A character vector of variable names. For each variable a text file containing the aligned dataset is written to the working directory. Vector elements need to correspond to column names of data.
rt_cutoff_low	A numeric value giving a retention time threshold. Peaks with retention time below the threshold are removed in a preprocessing step.
rt_cutoff_high	A numeric value giving a retention time threshold. Peaks with retention time above the threshold are removed in a preprocessing step.
reference	A character giving the name of sample from the dataset. By default, a sample is automatically selected from the dataset using the function <a href="#">choose_optimal_reference</a> . The reference is used for the full alignment of peak lists by linear transformation.
max_linear_shift	Numeric value giving the window size considered in the full alignment. Usually, the amplitude of linear drift is small in typical GC-FID datasets. Therefore, the default value of 0.05 minutes is adequate for most datasets. Increase this value if the drift amplitude is larger.
max_diff_peak2mean	Numeric value defining the allowed deviation of the retention time of a given peak from the mean of the corresponding row (i.e. scored substance). This parameter reflects the retention time range in which peaks across samples are still matched as homologous peaks (i.e. substance). Peaks with retention times exceeding the threshold are sorted into a different row.
min_diff_peak2peak	Numeric value defining the expected minimum difference in retention times among homologous peaks (i.e. substance). Rows that differ less in the mean retention time, are therefore merged if every sample contains either one or none of the respective compounds. This parameter is a major determinant in the classification of distinct peaks. Therefore careful consideration is required to adjust this setting to your needs (e.g. the resolution of your gas-chromatography pipeline). Large values may cause to merge truly different substances with similar retention times, if those are not simultaneously occurring within at least one

	individual, which might occur by chance for small sample sizes. By default set to 0.2 minutes.
blanks	Character vector of names of negative controls. Substances found in any of the blanks will be removed from the aligned dataset, before the blanks are deleted from the aligned data as well. This is an optional filtering step.
delete_single_peak	Boolean, determining whether substances that occur in just one sample are removed or not.
remove_empty	Boolean, allows to remove samples which lack any peak after the alignment finished. By default FALSE
permute	Boolean, by default a random permutation of samples is conducted prior for each row-wise alignment step. Setting this parameter to FALSE causes alignment of the dataset as it is. order of samples is constantly randomised during the alignment. Allows to prevent this behaviour for maximal repeatability if needed.
...	optional arguments passed to methods, see <a href="#">barplot</a> .

## Details

This function aligns and matches homologous peaks across samples using a three-step algorithm based on user-defined parameters that are explained in the next section. In brief: **(1)** A full alignment of peak retention times is conducted to correct for systematic linear drift of retention times among homologous peaks from run to run. Thereby a coarse alignment is achieved that maximises the similarity of retention times across homologous peaks prior to a **(2)** partial alignment and matching of peaks. This and the next step in the alignment is based on a retention time matrix that contains all samples as columns and peak retention times in rows. The goal is to match homologous peaks within the same row that represents a chemical substance. Here, peaks are recognised as homologous when the retention time matches within a user-defined error span (see `max_diff_peak2mean`) that approximates the expected retention time uncertainty. Here, the position of every peak in the matrix is evaluated in comparison to similar peaks across the complete dataset and homologous peaks are gradually grouped together row by row. After all peaks were matched, a **(3)** adjacent rows of similar retention time are scanned to detect redundancies. A pair of rows is identified as redundant and merged if mean retention times are closer than specified by `min_diff_peak2peak` and single samples only contain peak in one but not both rows. Therefore, this step allows to match peaks that are associated with higher variance than allowed during the previous step. Several optional processing steps are available, ranging from the removal of peaks representing contaminations (requires to include blanks as a control) to the removal of uninformative peaks that are present in just one sample (so called singletons).

## Value

Returns an object of class "GCalign" that is a list containing several objects that are listed below. Note, that the objects "heatmap\_input" and "Logfile" are best inspected by calling the provided functions `gc_heatmap` and `print`.

aligned	Aligned Gas Chromatography peak data subdivided into individual data frames for every variable. Samples are represented by columns, rows specify homologous peaks. The first column of every data frame is comprised of the mean
---------	--

retention time of the respective peak (i.e. row). Retention times of samples resemble the values of the raw data. Internally, linear adjustments are considered where appropriate

heatmap_input	Used internally to create heatmaps of the aligned data
Logfile	A protocol of the alignment process.
input_list	Input data in form of a list of data frames.
aligned_list	Aligned data in form of a list of data frames.
input_matrix	List of matrices. Each matrix contains the input data for a variable

### Author(s)

Martin Stoffel (martin.adam.stoffel@gmail.com) & Meinolf Ottensmann (meinolf.ottensmann@web.de)

### Examples

```
## Load example dataset
data("peak_data")
## Subset for faster processing
peak_data <- peak_data[1:3]
peak_data <- lapply(peak_data, function(x) x[1:50,])
## align data with default settings
out <- align_chromatograms(peak_data, rt_col_name = "time")
```

---

as.data.frame.GCalign *Output aligned data in form of a data frame for each variable*

---

### Description

Based on an object of class "GCalign" that was created using [align\\_chromatograms](#), a list of data frames for each variable in the dataset is returned. Within data frames rows represent substances and columns are variables (i.e. substances).

### Usage

```
## S3 method for class 'GCalign'
as.data.frame(x, row.names = NULL, optional = FALSE, ...)
```

### Arguments

x	An object of class "GCalign". See <a href="#">align_chromatograms</a> for details.
row.names	NULL or a character vector giving the row names for the data frame. Missing values are not allowed.

optional      logical. If TRUE, setting row names and converting column names (to syntactic names: see `make.names`) is optional. Note that all of R's **base** package `as.data.frame()` methods use `optional` only for column names treatment, basically with the meaning of `data.frame(*, check.names = !optional)`. See also the `make.names` argument of the `matrix` method.

...            additional arguments to be passed to or from methods.

### Author(s)

Meinolf Ottensmann (meinolf.ottensmann@web.de) & Martin Stoffel (martin.adam.stoffel@gmail.com)

### Examples

```
data("aligned_peak_data")
out <- as.data.frame(x = aligned_peak_data)
```

---

blank\_substraction      *Subtraction of blank readings from sample readings*

---

### Description

For each substance that is present in blanks, samples are corrected by subtraction of the respective quantity. If more than one sample is submitted, abundances are averaged. This procedure is sensitive to differences in the total concentration of samples and should be applied to samples where the preparation yields comparable concentrations for each sample.

### Usage

```
blank_substraction(input = NULL, blanks = NULL, conc_col_name = NULL)
```

### Arguments

`input`            A GCalign Object

`blanks`            Character vector of names of negative controls.

`conc_col_name`    If the input is a GCalign object the variable containing the abundance values needs to be specified.

### Details

Substances that are present in one or more blanks are identified in the aligned dataset, then the mean abundance is calculated for the blanks and the corresponding value is subtracted from each sample. If the control contains higher concentration (i.e. blank subtraction creates negative abundances) warnings will be shown and the respective value will be set to zero

**Examples**

```
## Not run
#out <- blank_substraction(aligned_peak_data, blanks = "M2", conc_col_name = "area")
```

---

check\_input

*Check input prior to processing in GCalignR*


---

**Description**

Checks input files for common formatting problems.

**Usage**

```
check_input(data, plot = FALSE, sep = "\t", message = TRUE, ...)
```

**Arguments**

data	Dataset containing peaks that need to be aligned and matched. For every peak a arbitrary number of numerical variables can be included (e.g. peak height, peak area) in addition to the mandatory retention time. The standard format is a tab-delimited text file according to the following layout: (1) The first row contains sample names, the (2) second row column names of the corresponding peak lists. Starting with the third row, peak lists are included for every sample that needs to be incorporated in the dataset. Here, a peak list contains data for individual peaks in rows, whereas columns specify variables in the order given in the second row of the text file. Peak lists of individual samples are concatenated horizontally and need to be of the same width (i.e. the same number of columns in consistent order). Alternatively, the input may be a list of data frames. Each data frame contains the peak data for a single individual. Variables (i.e.columns) are named consistently across data frames. The names of elements in the list are used as sample identifiers. Cells may be filled with numeric or integer values but no factors or characters are allowed. NA and 0 may be used to indicate empty rows.
plot	Boolean specifying if the distribution of peak numbers is plotted.
sep	The field separator character. The default is tab separated (sep = '\t'). See the "sep" argument in <a href="#">read.table</a> for details.
message	Boolean determining if passing all checks is indicated by a message.
...	optional arguments passed to methods, see <a href="#">barplot</a> .

**Details**

Sample names should contain just letters, numbers and underscores and no whitespaces. Each sample has to contain the same number of columns, one of which is the retention time and the others are arbitrary variables in consistent order across samples. Retention times are expected to be numeric, i.e. they are only allowed to contain numbers from 0-9 and "." as the only decimal character. Have a look at the vignettes for examples.



**Author(s)**

Martin Stoffel (martin.adam.stoffel@gmail.com) & Meinolf Ottensmann (meinolf.ottensmann@web.de)

**Examples**

```
## gc-data
data("peak_data")
## Checks format
check_input(peak_data)
## Includes a barplot of peak numbers in the raw data
check_input(peak_data, plot = TRUE)
```

---

choose\_optimal\_reference

*Select the optimal reference for full alignments of peak lists*

---

**Description**

Full alignments of peak lists require the specification of a fixed reference to which all other samples are aligned to. This function provides an simple algorithm to find the most suitable sample among a dataset. The so defined reference can be used for full alignments using [linear\\_transformation](#). The functions is evoked internally by [align\\_chromatograms](#) if no reference was specified by the user.

**Usage**

```
choose_optimal_reference(data = NULL, rt_col_name = NULL, sep = "\t")
```

**Arguments**

**data** Dataset containing peaks that need to be aligned and matched. For every peak a arbitrary number of numerical variables can be included (e.g. peak height, peak area) in addition to the mandatory retention time. The standard format is a tab-delimited text file according to the following layout: (1) The first row contains sample names, the (2) second row column names of the corresponding peak lists. Starting with the third row, peak lists are included for every sample that needs to be incorporated in the dataset. Here, a peak list contains data for individual peaks in rows, whereas columns specify variables in the order given in the second row of the text file. Peak lists of individual samples are concatenated horizontally and need to be of the same width (i.e. the same number of columns in consistent order). Alternatively, the input may be a list of data frames. Each data frame contains the peak data for a single individual. Variables (i.e.columns) are named consistently across data frames. The names of elements in the list are used as sample identifiers. Cells may be filled with numeric or integer values but no factors or characters are allowed. NA and 0 may be used to indicate empty rows.

rt_col_name	A character giving the name of the column containing the retention times. The decimal separator needs to be a point.
sep	The field separator character. The default is tab separated (sep = '\t'). See the "sep" argument in <a href="#">read.table</a> for details.

### Details

Every sample is considered in determining the optimal reference in comparison to all other samples by estimating the similarity to all other samples. For a reference-sample pair, the deviation in retention times between all reference peaks and the always nearest peak in the sample is summed up and divided by the number of reference peaks. The calculated value is a similarity score that converges to zero the more similar reference and sample are. For every potential reference, the median score of all pair-wise comparisons is used as a similarity proxy. The optimal sample is then defined by the minimum value among these scores. This functions is used internally in [align\\_chromatograms](#) to select a reference if non was specified by the user.

### Value

A list with following elements

sample	Name of the sample with the highest average similarity to all other samples
score	Median number of shared peaks with other samples

### Author(s)

Martin Stoffel (martin.adam.stoffel@gmail.com) & Meinolf Ottensmann (meinolf.ottensmann@web.de)

### Examples

```
## 1.) input is a list
## using a list of samples
data("peak_data")
## subset for faster processing
peak_data <- peak_data[1:3]
choose_optimal_reference(peak_data, rt_col_name = "time")
```

---

draw_chromatogram	<i>Visualise peak lists as a pseudo-chromatogram</i>
-------------------	--

---

### Description

Creates a graphical representation of one or multiple peak lists in the form of a pseudo- chromatogram. Peaks are represented by Gaussian distributions centred at the peak retention time. The peak height is arbitrary and does not reflect any measured peak intensity.

**Usage**

```
draw_chromatogram(
  data = NULL,
  rt_col_name = NULL,
  conc_col_name = NULL,
  width = 0.1,
  step = NULL,
  sep = "\t",
  breaks = NULL,
  rt_limits = NULL,
  samples = NULL,
  show_num = FALSE,
  show_rt = FALSE,
  plot = TRUE,
  shape = c("gaussian", "stick"),
  legend.position = "bottom"
)
```

**Arguments**

data	The input data can be either a GCalgnR input file or an GCalgn object. See <a href="#">align_chromatograms</a> for details on both.
rt_col_name	A character giving the name of the column containing the retention times. The decimal separator needs to be a point.
conc_col_name	Character, denoting a variable used to scale the peak height (e.g., peak area or peak height.)
width	Numeric value giving the standard deviation of Gaussian peaks. Decrease this value to separate overlapping peaks within samples. Default is 0.01.
step	character allowing to visualise different steps of the alignment when a GCalgn object is used. By default the aligned data is shown.
sep	The field separator character. The default is tab separated (sep = '\t'). See the "sep" argument in <a href="#">read.table</a> for details.
breaks	A numeric vector giving the breakpoints between ticks on the x axis.
rt_limits	A numeric vector of length two giving min and max values or retention times to plot.
samples	A character vector of sample names to draw chromatograms of a subset.
show_num	Boolean indicating whether sample numbers are drawn on top of each peak.
show_rt	Boolean indicating whether peak retention times are drawn on top of each peak.
plot	Boolean indicating if the plot is printed.
shape	A character determining the shape of peaks. Peaks are approximated as "gaussian" by default. Alternatively, peaks can be visualised as "sticks".
legend.position	See <a href="#">theme</a> for options of legend positions.

## Details

Peaks from the are depicted as Gaussian distributions. If the data is an "GCalign" object that was processed with `align_chromatograms`, chromatograms can be drawn for the dataset prior to alignment ("**input**"), after correcting linear drift ("**shifted**") or after the complete alignment was conducted ("**aligned**"). In the latter case, retention times refer to the mean retention time of a homologous peaks scored among samples and do not reflect any between-sample variation anymore. Depending on the range of retention times and the distance among substances the peak width can be adjusted to enable a better visual separation of peaks by changing the value of parameter `width`. Note, homologous peaks (= exactly matching retention time) will overlap completely and only the last sample plotted will be visible. Hence, the number of samples can be printed on top of each peak. The function returns a list containing the ggplot object along with the internally used data frame to allow for maximum control in adapting the plot (see examples section in this document).

## Value

A list containing the data frame created for plotting and the ggplot object. See `ggplot`.

## Author(s)

Meinolf Ottensmann (meinolf.ottensmann@web.de) & Martin Stoffel (martin.adam.stoffel@gmail.com)

## Examples

```
## load data
path <- (system.file("extdata", "simulated_peak_data.txt", package = "GCalignR"))
## run with defaults
x <- draw_chromatogram(data = path, rt_col_name = "rt")
## Customise and split samples in panels
x <- draw_chromatogram(data = path, rt_col_name = "rt", samples = c("A2", "A4"),
  plot = FALSE, show_num = FALSE)
x[["ggplot"]] + ggplot2::facet_wrap(~ sample, nrow = 2)
## plot without numbers
x <- draw_chromatogram(data = path, show_num = FALSE, rt_col_name = "rt")
```

---

find\_peaks

*Detect local maxima in time series*

---

## Description

Detects peaks in a vector and calculates the peak height. This function is only appropriate for symmetric gaussian peaks and does not take into account any baseline correction as it required in 'real word' data. Therefore, it does not substitute sophisticated peak detection and integration tools and is only used for illustration purposes in our vignettes.

## Usage

```
find_peaks(df)
```

**Arguments**

df                    A data frame containing x and y coordinates.

**Value**

A data frame containing x and y coordinates of peaks.

**Author(s)**

Meinolf Ottensmann (meinolf.ottensmann@web.de) & Martin Stoffel (martin.adam.stoffel@gmail.com)

**Examples**

```
## create df
df <- data.frame(x = 1:1000, y = dnorm(1:1000,300,20))
## plot
with(df, plot(x,y))
## detect peak
find_peaks(df)
```

---

GCalignR

*GCalignR: A Package to Align Gas Chromatography Peaks Based on Retention Times*

---

**Description**

GCalignR contains the functions listed below. Follow the links to access the documentation of each function.

[align\\_chromatograms](#) executes all alignment steps.

[as.data.frame.GCalign](#) exports aligned data to data frames.

[check\\_input](#) tests the input data for formatting issues.

[draw\\_chromatogram](#) visualises peak lists in form of a chromatogram.

[find\\_peaks](#) detects and calculates peak heights in chromatograms. Not intended to be used for peak integration in empirical data. Used for illustration purposes only.

[gc\\_heatmap](#) visualises aligned datasets using heatmaps that can be customised.

[norm\\_peaks](#) allows to compute the relative abundance of peaks with samples.

[peak\\_interspace](#) gives a histogram of the distance between peaks within samples over the whole dataset.

[read\\_peak\\_list](#) reads the content of a text file and converts it to a list.

[remove\\_blanks](#) removes peaks resembling contaminations from aligned datasets.

[remove\\_singletons](#) removes peaks that are unique for one individual sample.

[simple\\_chroma](#) creates simple chromatograms for testing and illustration purposes.

## Details

More details on the package are found in the vignettes that can be accessed via `browseVignettes("GCalignR")`.

## Author(s)

**Maintainer:** Meinolf Ottensmann <meinolf.ottensmann@web.de> ([ORCID](#))

Authors:

- Martin Stoffel <martin.adam.stoffel@gmail.com>
- Hazel J. Nichols
- Joseph I. Hoffman

## See Also

Useful links:

- <https://github.com/mottensmann/GCalignR>
- Report bugs at <https://github.com/mottensmann/GCalignR/issues>

---

gc\_heatmap

*Visualises peak alignments in form of a heatmap*

---

## Description

The goal of aligning peaks is to match homologous peaks that are thought to represent homologous substances in the same row across samples, although peaks have slightly different retention times across samples. This function makes it possible to evaluate the alignment quickly by inspecting the (i) distribution of peaks across samples, (ii) the variation for each homologous peak (column) as well as (iii) patterns that might hint at splitting peaks across rows. The mean retention time per homologous peak is here defined as the "true" retention time and deviations of individual peaks can be seen by a large deviation in the retention time to the mean. Subsetting of the retention time range (i.e. selecting peaks by the mean retention time) and samples (by name or by position) allow to quickly inspect regions of interest. Two types of heatmaps are available, a binary heatmap allows to determine if the retention time of single samples deviates by more than the user defined threshold from the mean. Optionally, a discrete heatmap allows to check deviations quantitatively. Large deviation can have multiple reasons. The most likely explanation is given by the fact that adjacent rows were merged as specified by the value `min_diff_peak2peak` in `align_chromatograms`. Here clear cases, in which peaks of multiple samples have been grouped in either of two or more rows can be merged and cause relatively high variation in peak retention times.

**Usage**

```
gc_heatmap(
  object = NULL,
  algorithm_step = c("aligned", "shifted", "input"),
  substance_subset = NULL,
  legend_type = c("legend", "colourbar"),
  samples_subset = NULL,
  type = c("binary", "discrete"),
  threshold = NULL,
  label_size = NULL,
  show_legend = TRUE,
  main_title = NULL,
  label = c("y", "xy", "x", "none")
)
```

**Arguments**

object	Object of class "GCalign", the output of a call to <a href="#">align_chromatograms</a> .
algorithm_step	Character indicating which step of the algorithm is plotted. Either "input", "shifted" or "aligned" specifying the raw, linearly shifted or aligned data respectively. Default is the heatmap for the aligned dataset.
substance_subset	A vector of integers containing indices of substances in ascending order of retention times to plot.
legend_type	A character specifying how to present deviations of retention times from the mean. Either in form of discrete steps or on a gradient scale using 'legend' or 'colourbar' respectively. Changes are only possible when type = "discrete"
samples_subset	A vector indicating which samples are plotted on the heatmap by giving either indices or names of samples.
type	A character specifying whether a deviations of retention times are shown 'binary' (i.e. in comparison to the threshold value) or on a 'discrete' scale with respect to the mean retention time.
threshold	A numeric value denoting the threshold above which the deviation of individual peak retention times from the mean retention time of the respective substance are highlighted in heatmaps. By default, the value of parameter <code>max_diff_peak2mean</code> (see <a href="#">align_chromatograms</a> ) that was used in aligning the data is used.
label_size	An integer determining the size of labels on y and x axis. By default a fitting label_size is calculate (beta!) to compromise between readability and messiness due to a potentially large number of substances and samples.
show_legend	Boolean determining whether a legend is included or not.
main_title	Character giving the title of the heatmap. If not specified, titles are generated automatically.
label	Character determining if labels are shown on axes. Depending on the number of peaks and/or samples, labels are difficult to read. Use subsets instead. Possible option are "xy", "x", "y" or "none"

**Value**

object of class "ggplot"

**Author(s)**

Martin Stoffel (martin.adam.stoffel@gmail.com) & Meinolf Ottensmann (meinolf.ottensmann@web.de)

**Examples**

```
## aligned gc-dataset
data("aligned_peak_data")
## Default settings: The final output is plotted
gc_heatmap(aligned_peak_data, algorithm_step = "aligned")

## Plot the input data
gc_heatmap(aligned_peak_data, algorithm_step = "input")

## Plot a subset of the first 50 scored substances
gc_heatmap(aligned_peak_data, algorithm_step="aligned", substance_subset = 1:50)

## Plot specific samples, apply a stricter threshold
gc_heatmap(aligned_peak_data, samples_subset = c("M2", "P7", "M13", "P13"), threshold = 0.02)
```

---

linear\_transformation *Full Alignment of Peak Lists by linear retention time correction.*

---

**Description**

Shifts all peaks within samples to maximise the similarity to a reference sample. For optimal results, a sufficient number of shared peaks are required to find an optimal solution. A reference needs to be specified, for instance using [choose\\_optimal\\_reference](#). Linear shifts are evaluated within a user-defined window in discrete steps. The highest similarity score defines the shift that will be applied. If more than a single shift step yields to the same similarity score, the smallest absolute value wins in order to avoid overcompensation. The function is invoked internally by [align\\_chromatograms](#).

**Usage**

```
linear_transformation(
  gc_peak_list,
  reference,
  max_linear_shift = 0.05,
  step_size = 0.01,
  rt_col_name,
  Logbook = NULL
)
```



**Arguments**

<code>gc_peak_list</code>	List of data.frames. Each data.frame contains GC-data (e.g. retention time, peak area, peak height) of one sample. Variables are stored in columns. Rows represent distinct peaks. Retention time is a required variable.
<code>reference</code>	A character giving the name of a sample included in the dataset. All samples are aligned to the reference.
<code>max_linear_shift</code>	Numeric value giving the window size considered in the full alignment. Usually, the amplitude of linear drift is small in typical GC-FID datasets. Therefore, the default value of 0.05 minutes is adequate for most datasets. Increase this value if the drift amplitude is larger.
<code>step_size</code>	Integer giving the step size in which linear shifts are evaluated between <code>max_linear_shift</code> and <code>-max_linear_shift</code> .
<code>rt_col_name</code>	A character giving the name of the column containing the retention times. The decimal separator needs to be a point.
<code>Logbook</code>	A list. If present, a summary of the applied linear shifts in full alignments of peak lists is appended to the list. If not specified, a list will be created automatically.

**Details**

A similarity score is calculated as the sum of deviations in retention times between all reference peaks and the closest peak in the sample. The principle idea is that the appropriate linear transformation will reduce the deviation in retention time between homologous peaks, whereas all other peaks should deviate randomly. Among all considered shifts, the minimum deviation score is selected for subsequent full alignment by shifting all peaks of the sample by the same value.

**Value**

A list containing two items.

<code>chroma_aligned</code>	List containing the transformed data
<code>Logbook</code>	Logbook, record of the applied shifts

**Author(s)**

Martin Stoffel (martin.adam.stoffel@gmail.com) & Meinolf Ottensmann (meinolf.ottensmann@web.de)

**Examples**

```
dat <- peak_data[1:10]
dat <- lapply(dat, function(x) x[1:50,])
x <- linear_transformation(gc_peak_list = dat, reference = "C2", rt_col_name = "time")
```

---

merge\_redundant\_rows *Merge redundant rows*

---

## Description

Sometimes, redundant rows (i.e. groups of resembling a homologous peak) remain in an aligned dataset. This is the case when two or more adjacent rows exhibit a difference in the mean retention time that is greater than `min_diff_peak2peak`, the parameter that determines a threshold below that redundancy is checked within [align\\_chromatograms](#). Therefore, this function allows to raise the threshold for a post processing step that groups the homologous peaks together without the need of repeating a potentially time-consuming alignment with adjusted parameters.

## Usage

```
merge_redundant_rows(data, min_diff_peak2peak = NULL)
```

## Arguments

`data` An object of class "GCalign". See [align\\_chromatograms](#) for details.  
`min_diff_peak2peak` A numerical giving a threshold in minutes below which rows of similar retention time are checked for redundancy.

## Details

Based on the value of parameter `threshold`, possibly redundant rows are identified by comparing mean retention times. Next, rows are checked for redundancy. When one or more samples contain peaks in a pair of compared rows, no redundancy is existent and the pair is skipped.

## Value

a list of two items

`GCalign` input data with updated input to [gc\\_heatmap](#)  
`peak_list` a list of data frames containing the updated dataset

## Author(s)

Meinolf Ottensmann (meinolf.ottensmann@web.de) & Martin Stoffel (martin.adam.stoffel@gmail.com)

## Examples

```
## Load example dataset
data("peak_data")
## Subset for faster processing
peak_data <- peak_data[1:3]
peak_data <- lapply(peak_data, function(x) x[1:50,])
## align data with strict parameters
```

```
out <- align_chromatograms(peak_data, rt_col_name = "time",
max_diff_peak2mean = 0.01, min_diff_peak2peak = 0.02)
## relax threshold to merge redundant rows
out2 <- merge_redundant_rows(data = out, min_diff_peak2peak = 0.05)
```

---

norm\_peaks

*Normalisation of peak abundancies*

---

## Description

Calculates the relative abundance of a peak by normalising an intensity measure with regard to the cumulative abundance of all peaks that are present within an individual sample. The desired measure of peak abundance needs to be included in a column of the input dataset aligned with [align\\_chromatograms](#).

## Usage

```
norm_peaks(
  data,
  conc_col_name = NULL,
  rt_col_name = NULL,
  out = c("data.frame", "list")
)
```

## Arguments

data	Object of class GCalign created with <a href="#">align_chromatograms</a> or a list of data frames that contain peak list of individual samples.
conc_col_name	Character giving the name of a column in data containing a variable describing the abundance of peaks (e.g. peak area or peak height).
rt_col_name	A character giving the name of the column containing the retention times. The decimal separator needs to be a point.
out	character defining the format of the returned data. Either "List" or "data.frame".

## Value

Depending on out either a list of data frame or a single data frame were rows represent samples and columns relative peak abundances. Abundances are given as percentages.

@author Martin Stoffel (martin.adam.stoffel@gmail.com) & Meinolf Ottensmann (meinolf.ottensmann@web.de)

## Examples

```
## aligned gc-dataset
data("aligned_peak_data")
## returns normalised peak area
norm_peaks(data = aligned_peak_data, conc_col_name = "area", rt_col_name = "time")
```

---

peak_data	<i>Gas-chromatography data for Antarctic Fur Seals (Arctocephalus gazella)</i>
-----------	--

---

### Description

This is an example of a typical gas-chromatography output file, listing a number of peaks with their respective retention times and abundance measures. Peaks were detected using Xcalibur 2.0.5 (Thermo Fisher Scientific). The data consists of 41 mother-pup pairs of two different colonies from Bird Island, South Georgia. In addition two blanks (i.e. negative controls) are included.

### Format

A list of `data.frame`'s. Each `data.frame` contains gas-chromatography peak data of a single sample. The variables within each `data.frame` are: "time" (peak retention time) and "area" (integral of the peak curve). Each list element i.e. each `data.frame` is named with the unique sample ID.

### Source

<http://www.pnas.org/content/suppl/2015/08/05/1506076112.DCSupplemental/pnas.1506076112.sd02.xlsx>

### References

Stoffel, M.A.; Caspers, B.A.; Forcada, J.; Giannakara, A.; Baier, M.; Eberhart-Phillips, L.; Mueller, C.; Hoffman, J.I. (2015): Chemical fingerprints encode mother-offspring similarity, colony membership, relatedness, and genetic quality in fur seals. In: *Proceedings of the National Academy of Sciences of the United States of America* 112 (36), S. E5005-12. DOI: 10.1073/pnas.1506076112.

---

peak_factors	<i>Grouping factors corresponding to gas-chromatography data of Antarctic Fur Seals (Arctocephalus gazella)</i>
--------------	---

---

### Description

List of factors corresponding to samples in [peak\\_data](#)

### Format

A data frame where columns represent factors, rows are samples.

### Source

<http://www.pnas.org/content/suppl/2015/08/05/1506076112.DCSupplemental/pnas.1506076112.sd02.xlsx>

## References

Stoffel, M.A.; Caspers, B.A.; Forcada, J.; Giannakara, A.; Baier, M.; Eberhart-Phillips, L.; Mueller, C.; Hoffman, J.I. (2015): Chemical fingerprints encode mother-offspring similarity, colony membership, relatedness, and genetic quality in fur seals. In: Proceedings of the National Academy of Sciences of the United States of America 112 (36), S. E5005-12. DOI: 10.1073/pnas.1506076112.

---

peak_interspace	<i>Estimate the observed space between peaks within chromatograms</i>
-----------------	---

---

## Description

The parameter `min_diff_peak2peak` is a major determinant in the alignment of a dataset with [align\\_chromatograms](#). This function helps to infer a suitable value based on the input data. The underlying assumption here is that distinct peaks within a separated by a larger gap than homologous peaks across samples. Tightly spaced peaks within a sample will appear on the left side of the plotted distribution and can indicate the presence of split peaks in the data.

## Usage

```
peak_interspace(  
  data,  
  rt_col_name = NULL,  
  sep = "\t",  
  quantiles = NULL,  
  quantile_range = c(0, 1),  
  by_sample = FALSE  
)
```

## Arguments

<code>data</code>	Dataset containing peaks that need to be aligned and matched. For every peak a arbitrary number of numerical variables can be included (e.g. peak height, peak area) in addition to the mandatory retention time. The standard format is a tab-delimited text file according to the following layout: (1) The first row contains sample names, the (2) second row column names of the corresponding peak lists. Starting with the third row, peak lists are included for every sample that needs to be incorporated in the dataset. Here, a peak list contains data for individual peaks in rows, whereas columns specify variables in the order given in the second row of the text file. Peak lists of individual samples are concatenated horizontally and need to be of the same width (i.e. the same number of columns in consistent order). Alternatively, the input may be a list of data frames. Each data frame contains the peak data for a single individual. Variables (i.e. columns) are named consistently across data frames. The names of elements in the list are used as sample identifiers. Cells may be filled with numeric or integer values but no factors or characters are allowed. NA and 0 may be used to indicate empty rows.
-------------------	--

rt_col_name	A character giving the name of the column containing the retention times. The decimal separator needs to be a point.
sep	The field separator character. The default is tab separated (sep = '\t'). See the "sep" argument in <a href="#">read.table</a> for details.
quantiles	A numeric vector. Specified quantiles are calculated from the distribution.
quantile_range	A numeric vector of length two that allows to subset an arbitrary interquartile range.
by_sample	A logical that allows to calculate peak interspaces individually for each sample. By default all samples are combined to give the global distribution of next-peak differences in retention times. When by_sample = TRUE, a series of plots (one for each sample) is created and a keystroke is required to proceed.

**Value**

List containing summary statistics of the peak interspace distribution

**Author(s)**

Martin Stoffel (martin.adam.stoffel@gmail.com) & Meinolf Ottensmann (meinolf.ottensmann@web.de)

**Examples**

```
## plotting with defaults
peak_interspace(data = peak_data, rt_col_name = "time")
## plotting up to the 0.95 quantile
peak_interspace(data = peak_data, rt_col_name = "time", quantile_range = c(0,0.95))
## return the 0.1 quantile
peak_interspace(data = peak_data, rt_col_name = "time", quantiles = 0.1)
```

---

plot.GCalign

*Plot diagnostics for an GCalign Object*

---

**Description**

Visualises the aligned data based on four diagnostic plots. One plot shows the distribution of peak numbers per sample in the raw data and after alignment. A second plot gives the distribution of linear shifts that were applied in order to conduct a full alignment of samples with respect to reference. A third sample gives a distribution of the variation in retention times of homologous peaks. The fourth plot shows a frequency distribution of peaks shared among samples.

**Usage**

```
## S3 method for class 'GCalign'
plot(
  x,
  which_plot = c("all", "shifts", "variation", "peak_numbers", "peaks_shared"),
  ...
)
```

**Arguments**

x                    Object of class GCalign, created with [align\\_chromatograms](#)  
 which\_plot        A character defining which plot is created. Options are "shifts", "variation",  
                     "peak\_numbers" and "peaks\_shared". By default all four are created.  
 ...                Optional arguments passed on to methods. See [plot](#), [hist](#) and [barplot](#). Note  
                     that optional arguments are not passed on when plotting all figures.

**Value**

Depending on the selected plot a data frame containing the data source of the respective plot is returned. If all plots are created, no output will be returned.

**Author(s)**

Martin Stoffel (martin.adam.stoffel@gmail.com) & Meinolf Ottensmann (meinolf.ottensmann@web.de)

**Examples**

```
## GCalign object
data("aligned_peak_data")

## All plots are shown by default
plot(aligned_peak_data)

## Distribution of peak numbers
plot(aligned_peak_data, which_plot = "peak_numbers")

## variation of retention times
plot(aligned_peak_data, which_plot = "variation")
```

---

 print.GCalign

*Summarising Peak Alignments with GCalignR*


---

**Description**

print method for class "GCalign"

**Usage**

```
## S3 method for class 'GCalign'
print(x, write_text_file = FALSE, ...)
```

**Arguments**

x                    Object of class GCalign, created with [align\\_chromatograms](#)  
 write\_text\_file    A boolean allowing to write a text file.  
 ...                Optional arguments passed on to methods are currently not supported.

**Author(s)**

Martin Stoffel (martin.adam.stoffel@gmail.com) & Meinolf Ottensmann (meinolf.ottensmann@web.de)

**Examples**

```
## GCalign object
data("aligned_peak_data")
## print summary
print(aligned_peak_data)
```

---

read\_empower2

*Import data from single EMPOWER2 HPLC files*

---

**Description**

reads output files of the EMPOWER 2 SOFTWARE (Waters). Input files must contain data of single samples deposited within the same directory.

**Usage**

```
read_empower2(
  path = NULL,
  pattern = ".txt",
  sep = "\t",
  skip = 2,
  id = "SampleName"
)
```

**Arguments**

path	path to a folder containing input files
pattern	pattern used to select files. By default ".txt"
sep	The field separator character. The default is tab separated (sep = '\t'). See the "sep" argument in <a href="#">read.table</a> for details.
skip	rows to skip before reading data
id	column containing sample name

**Value**

a list of data frames (each corresponding to a sample)



---

read_peak_list	<i>Read content of a text file and convert it to a list</i>
----------------	---

---

### Description

Reads the content of text file that is formatted as described in [align\\_chromatograms](#) and converts it to a list.

### Usage

```
read_peak_list(data, sep = "\t", rt_col_name, check = T)
```

### Arguments

data	A text file containing a peak list. See <a href="#">align_chromatograms</a> for details.
sep	The field separator character. The default is tab separated (sep = '\t'). See the "sep" argument in <a href="#">read.table</a> for details.
rt_col_name	A character giving the name of the column containing the retention times. The decimal separator needs to be a point.
check	logical

### Value

A list of data frames containing peak data for every sample of data.

### Author(s)

Meinolf Ottensmann (meinolf.ottensmann@web.de) & Martin Stoffel (martin.adam.stoffel@gmail.com)

### Examples

```
path <- system.file("extdata", "simulated_peak_data.txt", package = "GCalignR")
x <- read_peak_list(data = path, rt_col_name = "rt")
```

---

remove_blanks	<i>Remove peaks present in negative control samples</i>
---------------	---

---

### Description

Removes peaks that are present in blanks (i.e. negative control samples) to eliminate contaminations in the aligned data. Afterwards, blanks are deleted itself. This function is only applicable when blanks were not discarded during a previous alignment using [align\\_chromatograms](#).

**Usage**

```
remove_blanks(data, blanks)
```

**Arguments**

**data** An object of class "GCalign". See [align\\_chromatograms](#) for details. Alternatively, a list of data frames. Whereby each data frame contains the peak list for an individual sample.

**blanks** Character vector of names of negative controls. Substances found in any of the blanks will be removed from the aligned dataset, before the blanks are deleted from the aligned data as well. This is an optional filtering step.

**Value**

a list of data frames for each individual.

**Author(s)**

Meinolf Ottensmann (meinolf.ottensmann@web.de) & Martin Stoffel (martin.adam.stoffel@gmail.com)

**Examples**

```
data("peak_data")
## subset for faster processing
data <- lapply(peak_data[1:5], function(x) x[20:35,])
x <- align_chromatograms(data, rt_col_name = "time")
out <- remove_blanks(data = x, blanks = c("C2", "C3"))
## number of deleted peaks
nrow(x[["aligned_list"]][["M2"]]) - nrow(out[["M2"]])
```

---

remove\_singletons      *Remove singletons*

---

**Description**

Identifies and removes singletons (i.e. peaks that are unique for one sample) from the aligned dataset.

**Usage**

```
remove_singletons(data)
```

**Arguments**

**data** An object of class "GCalign". See [align\\_chromatograms](#) for details. Alternatively, a list of data frames. Whereby each data frame contains the peak list for an individual sample.

**Value**

a list of data frames for each individual.

**Author(s)**

Meinolf Ottensmann (meinolf.ottensmann@web.de) & Martin Stoffel (martin.adam.stoffel@gmail.com)

**Examples**

```
data("peak_data")
## subset for faster processing
data <- lapply(peak_data[1:5], function(x) x[20:35,])
x <- align_chromatograms(data, rt_col_name = "time")
out <- remove_singletons(data = x)
```

---

simple\_chroma

*Simulate simple chromatograms*

---

**Description**

Creates chromatograms with user defined peaks for illustrative purposes. Linear drift is applied in sample order if more than one sample is created. See parameters of the function.

**Usage**

```
simple_chroma(  
  peaks = c(10, 13, 25, 37, 50),  
  N = 1,  
  min = 0,  
  max = 30,  
  Names = NULL,  
  sd = NULL  
)
```

**Arguments**

peaks	A numeric vector giving the retention times on which gaussian distribution, defining peaks, are centered. If more than one sample is generated $N > 1$ , peaks defines a population of peaks, from which samples are generated.
N	An integer giving the number of chromatograms to create. By default $N = 1$ .
min	A numeric giving the minimum retention time.
max	A numeric giving the maximum retention time.
Names	A character vector giving sample names. If not specified, names are generated automatically.
sd	A numeric vector of the same length as peaks giving the standard deviation of each peak. Only supported if $N = 1$ .

**Value**

A data frame containing x and y coordinates and corresponding sample names.

**Author(s)**

Meinolf Ottensmann (meinolf.ottensmann@web.de) & Martin Stoffel (martin.adam.stoffel@gmail.com)

**Examples**

```
## create a chromatogram
x <- simple_chroma(peaks = c(5,10,15), N = 1, min = 0, max = 30, Names = "MyChroma")
## plot chromatogram
with(x, plot(x,y, xlab = "time", ylab = "intensity"))
```

# Index

- \* **beta**
  - blank\_substraction, 7
  - read\_empower2, 24
- \* **datasets**
  - aligned\_peak\_data, 2
  - peak\_data, 20
  - peak\_factors, 20
- align\_chromatograms, 2, 3, 6, 9–16, 18, 19, 21, 23, 25, 26
- aligned\_peak\_data, 2
- as.data.frame.GCalign, 6, 13
  
- barplot, 5, 8, 23
- blank\_substraction, 7
  
- check\_input, 8, 13
- choose\_optimal\_reference, 4, 9, 16
  
- data.frame, 7
- draw\_chromatogram, 10, 13
  
- find\_peaks, 12, 13
  
- gc\_heatmap, 2, 13, 14, 18
- GCalignR, 3, 13
- GCalignR-package (GCalignR), 13
- ggplot, 12
  
- hist, 23
  
- linear\_transformation, 9, 16
  
- make.names, 7
- merge\_redundant\_rows, 18
  
- norm\_peaks, 13, 19
  
- peak\_data, 20, 20
- peak\_factors, 20
- peak\_interspace, 13, 21
- plot, 23
  
- plot.GCalign, 22
- print.GCalign, 2, 23
  
- read.table, 4, 8, 10, 11, 22, 24, 25
- read\_empower2, 24
- read\_peak\_list, 13, 25
- remove\_blanks, 13, 25
- remove\_singletons, 13, 26
  
- simple\_chroma, 13, 27
  
- theme, 11