Package 'HelpersMG'

January 4, 2019

Type Package

Title Tools for Earth Meteorological Analysis

Version 3.5

Date 2019-01-04

Author Marc Girondot

Maintainer Marc Girondot <marc.girondot@u-psud.fr>

Depends lme4, coda, R (>= 2.14.0)

Suggests RNetCDF, ncdf4, maps, XML, fields, shiny, Matrix, ppcor, pbmcapply, parallel, visNetwork, igraph

Description Contains many functions useful for managing 'NetCDF' files (see <http://en.wikipedia.org/wiki/NetCDF>), get tide levels on any point of the globe, get moon phase and time for sun rise and fall, analyse and reconstruct periodic time series of temperature with irregular sinusoidal pattern, show scales and wind rose in plot with change of color of text, Metropolis-Hastings algorithm for Bayesian MCMC analysis, plot graphs or boxplot with error bars, search files in disk by there names or their content, read the contents of all files from a folder at one time.

License GPL-2

LazyData yes

LazyLoad yes

Encoding UTF-8

RoxygenNote 6.1.1

NeedsCompilation no

Repository CRAN

Date/Publication 2019-01-04 16:00:18 UTC

R topics documented:

`HelpersMG-package` *Tools for Earth Meteorological Analysis*

Description

Contains functions useful for managing
'NetCDF' files (see http://en.wikipedia.org/wiki/NetCDF),
get tide levels on any point of the globe,
get moon phase and time for sun rise and fall,
analyse and reconstruct daily time series of temperature
with irregular sinusoidal pattern,
show scales and wind rose in plot with change of color of text,
Metropolis-Hastings algorithm for Bayesian MCMC analysis,
plot graphs or boxplot with error bars,
search files in disk by there names or their content,
read the contents of all files from a folder at one time,
calculate IC50 for ecotoxicological studies,
calculate the probability mass function of the sum of negative binomial
distributions.
The lastest version of this package can always been installed using:
install.packages("http://www.ese.u-psud.fr/epc/conservation/CRAN/HelpersMG.tar.gz", repos=NULL,
type="source")

Details

Helpers functions for several packages

Package: HelpersMG

Type:	Package
Version:	3.5 build 215
Date:	2019-01-04
License:	GPL (>= 2)
LazyLoad:	yes

Author(s)

Marc Girondot <marc.girondot@u-psud.fr>

Examples

```
## Not run:
library(HelpersMG)
print('-----------------------------------')
print('Examples for mcmcComposite objects')
print('-----------------------------------')
require(coda)
x <- rnorm(30, 10, 2)
dnormx <- function(x, par) return(-sum(dnorm(x, mean=par['mean'], sd=par['sd'], log=TRUE)))
parameters_mcmc <- data.frame(Density=c('dnorm', 'dlnorm'),
Prior1=c(10, 0.5), Prior2=c(2, 0.5), SDProp=c(0.35, 0.2),
Min=c(-3, 0), Max=c(100, 10), Init=c(10, 2), stringsAsFactors = FALSE,
row.names=c('mean', 'sd'))
mcmc_run <- MHalgoGen(n.iter=100000, parameters=parameters_mcmc, data=x,
likelihood=dnormx, n.chains=1, n.adapt=100, thin=1, trace=1)
plot(mcmc_run, xlim=c(0, 20))
plot(mcmc_run, xlim=c(0, 10), parameters="sd")
mcmcforcoda <- as.mcmc(mcmc_run)
# Optimal rejection rate should be 0.234
rejectionRate(mcmcforcoda)
heidel.diag(mcmcforcoda)
raftery.diag(mcmcforcoda)
autocorr.diag(mcmcforcoda)
acf(mcmcforcoda[[1]][,"mean"], lag.max=20, bty="n", las=1)
acf(mcmcforcoda[[1]][,"sd"], lag.max=20, bty="n", las=1)
batchSE(mcmcforcoda, batchSize=100)
# The batch standard error procedure is usually thought to
# be not as accurate as the time series methods used in summary
summary(mcmcforcoda)$statistics[,"Time-series SE"]
summary(mcmc_run)
as.parameters(mcmc_run)
lastp <- as.parameters(mcmc_run, index="last")
parameters_mcmc[,"Init"] <- lastp
# The n.adapt set to 1 is used to not record the first set of parameters
# then it is not duplicated (as it is also the last one for
# the object mcmc_run)
mcmc_run2 <- MHalgoGen(n.iter=10000, parameters=parameters_mcmc, data=x,
likelihood=dnormx, n.chains=1, n.adapt=1, thin=1, trace=1)
mcmc_run3 <- merge(mcmc_run, mcmc_run2)
####### no adaptation, n.adapt must be 0
```

```
parameters_mcmc[,"Init"] <- c(mean(x), sd(x))
mcmc_run3 <- MHalgoGen(n.iter=10000, parameters=parameters_mcmc, data=x,
likelihood=dnormx, n.chains=1, n.adapt=0, thin=1, trace=1)
print('-----------------------------------------')
print('Examples for Daily patterns of temperature')
print('-----------------------------------------')
# Generate a timeserie of time
time.obs <- NULL
for (i in 0:9) time.obs <- c(time.obs, c(0, 6, 12, 18)+i*24)
# For these time, generate a timeseries of temperatures
temp.obs <- rep(NA, length(time.obs))
temp.obs[3+(0:9)*4] <- rnorm(10, 25, 3)
temp.obs[1+(0:9)*4] <- rnorm(10, 10, 3)
for (i in 1:(length(time.obs)-1))
  if (is.na(temp.obs[i]))
  temp.obs[i] <- mean(c(temp.obs[i-1], temp.obs[i+1]))
  if (is.na(temp.obs[length(time.obs)]))
  temp.obs[length(time.obs)] <- temp.obs[length(time.obs)-1]/2
observed <- data.frame(time=time.obs, temperature=temp.obs)
# Search for the minimum and maximum values
r <- minmax.periodic(time.minmax.daily=c(Min=2, Max=15),
observed=observed, period=24)

# Estimate all the temperatures for these values
t <- temperature.periodic(minmax=r)

plot_errbar(x=t[,"time"], y=t[,"temperature"],
errbar.y=ifelse(is.na(t[,"sd"]), 0, 2*t[,"sd"]),
type="l", las=1, bty="n", errbar.y.polygon = TRUE,
xlab="hours", ylab="Temperatures", ylim=c(0, 35),
errbar.y.polygon.list = list(col="grey"))

plot_add(x=t[,"time"], y=t[,"temperature"], type="l")

## End(Not run)
```

`as.mcmc.mcmcComposite` *Extract mcmc object from a mcmcComposite object*

Description

Take a mcmcComposite object and create a mcmc.list object to be used with coda package.

Usage

```
## S3 method for class 'mcmcComposite'
as.mcmc(x, ...)
```

Arguments

x	A mcmcComposite obtained as a result of `MHalgoGen()` function
...	Not used

Details

as.mcmc Extract mcmc object from the result of phenology_MHmcmc to be used with coda package

Value

A mcmc.list object

Author(s)

Marc Girondot

See Also

Other mcmcComposite functions: `MHalgoGen`, `as.parameters`, `merge.mcmcComposite`, `plot.mcmcComposite`, `summary.mcmcComposite`

Examples

```
## Not run:
library(HelpersMG)
require(coda)
x <- rnorm(30, 10, 2)
dnormx <- function(data, x) {
 data <- unlist(data)
 return(-sum(dnorm(data, mean=x['mean'], sd=x['sd'], log=TRUE)))
}
parameters_mcmc <- data.frame(Density=c('dnorm', 'dlnorm'),
Prior1=c(10, 0.5), Prior2=c(2, 0.5), SDProp=c(1, 1),
Min=c(-3, 0), Max=c(100, 10), Init=c(10, 2), stringsAsFactors = FALSE,
row.names=c('mean', 'sd'))
mcmc_run <- MHalgoGen(n.iter=1000, parameters=parameters_mcmc, data=x,
likelihood=dnormx, n.chains=1, n.adapt=100, thin=1, trace=1)
plot(mcmc_run, xlim=c(0, 20))
plot(mcmc_run, xlim=c(0, 10), parameters="sd")
mcmcforcoda <- as.mcmc(mcmc_run)
#' heidel.diag(mcmcforcoda)
raftery.diag(mcmcforcoda)
autocorr.diag(mcmcforcoda)
acf(mcmcforcoda[[1]][,"mean"], lag.max=20, bty="n", las=1)
acf(mcmcforcoda[[1]][,"sd"], lag.max=20, bty="n", las=1)
batchSE(mcmcforcoda, batchSize=100)
# The batch standard error procedure is usually thought to
# be not as accurate as the time series methods used in summary
summary(mcmcforcoda)$statistics[,"Time-series SE"]
summary(mcmc_run)
as.parameters(mcmc_run)
```

```
lastp <- as.parameters(mcmc_run, index="last")
parameters_mcmc[,"Init"] <- lastp
# The n.adapt set to 1 is used to not record the first set of parameters
# then it is not duplicated (as it is also the last one for
# the object mcmc_run)
mcmc_run2 <- MHalgoGen(n.iter=1000, parameters=parameters_mcmc, data=x,
likelihood=dnormx, n.chains=1, n.adapt=1, thin=1, trace=1)
mcmc_run3 <- merge(mcmc_run, mcmc_run2)
####### no adaptation, n.adapt must be 0
parameters_mcmc[,"Init"] <- c(mean(x), sd(x))
mcmc_run3 <- MHalgoGen(n.iter=1000, parameters=parameters_mcmc, data=x,
likelihood=dnormx, n.chains=1, n.adapt=0, thin=1, trace=1)

## End(Not run)
```

`as.parameters` *Extract parameters from mcmcComposite object*

Description

Take a mcmcComposite object and create a vector object with parameter value at specified iteration.
If `index="best"`, the function will return the parameters for the highest likelihood. It also indicates at which iteration the maximum lihelihood has been observed.
If `index="last"`, the fuction will return the parameters for the last likelihood.
`index` can also be a numeric value.

Usage

```
as.parameters(x, index = "best", chain = 1)
```

Arguments

`x`	A mcmcComposite obtained as a result of `MHalgoGen()` function
`index`	At which iteration the parameters must be taken
`chain`	The number of the chain inwhich to get parameters

Value

A vector with parameters at maximum likelihood or index position

Author(s)

Marc Girondot

See Also

Other mcmcComposite functions: `MHalgoGen`, `as.mcmc.mcmcComposite`, `merge.mcmcComposite`, `plot.mcmcComposite`, `summary.mcmcComposite`

Examples

```
## Not run:
library(HelpersMG)
require(coda)
x <- rnorm(30, 10, 2)
dnormx <- function(data, x) {
 data <- unlist(data)
 return(-sum(dnorm(data, mean=x['mean'], sd=x['sd'], log=TRUE)))
}
parameters_mcmc <- data.frame(Density=c('dnorm', 'dlnorm'),
Prior1=c(10, 0.5), Prior2=c(2, 0.5), SDProp=c(1, 1),
Min=c(-3, 0), Max=c(100, 10), Init=c(10, 2), stringsAsFactors = FALSE,
row.names=c('mean', 'sd'))
mcmc_run <- MHalgoGen(n.iter=1000, parameters=parameters_mcmc, data=x,
likelihood=dnormx, n.chains=1, n.adapt=100, thin=1, trace=1)
plot(mcmc_run, xlim=c(0, 20))
plot(mcmc_run, xlim=c(0, 10), parameters="sd")
mcmcforcoda <- as.mcmc(mcmc_run)
#' heidel.diag(mcmcforcoda)
raftery.diag(mcmcforcoda)
autocorr.diag(mcmcforcoda)
acf(mcmcforcoda[[1]][,"mean"], lag.max=20, bty="n", las=1)
acf(mcmcforcoda[[1]][,"sd"], lag.max=20, bty="n", las=1)
batchSE(mcmcforcoda, batchSize=100)
# The batch standard error procedure is usually thought to
# be not as accurate as the time series methods used in summary
summary(mcmcforcoda)$statistics[,"Time-series SE"]
summary(mcmc_run)
as.parameters(mcmc_run)
lastp <- as.parameters(mcmc_run, index="last")
parameters_mcmc[,"Init"] <- lastp
# The n.adapt set to 1 is used to not record the first set of parameters
# then it is not duplicated (as it is also the last one for
# the object mcmc_run)
mcmc_run2 <- MHalgoGen(n.iter=1000, parameters=parameters_mcmc, data=x,
likelihood=dnormx, n.chains=1, n.adapt=1, thin=1, trace=1)
mcmc_run3 <- merge(mcmc_run, mcmc_run2)
####### no adaptation, n.adapt must be 0
parameters_mcmc[,"Init"] <- c(mean(x), sd(x))
mcmc_run3 <- MHalgoGen(n.iter=1000, parameters=parameters_mcmc, data=x,
likelihood=dnormx, n.chains=1, n.adapt=0, thin=1, trace=1)

## End(Not run)
```

---

asc *Return the codes (in UTF-8) of a string*

Description

Return the codes (in UTF-8) of a string.

Usage

```
asc(x)
```

Arguments

x	The string to be analyzed

Details

asc returns the codes (in UTF-8) of a string

Value

A vector with ITF-8 codes of a string

Author(s)

Based on this blog: http://datadebrief.blogspot.com/2011/03/ascii-code-table-in-r.html

See Also

Other Characters: chr, d, tnirp

Examples

```
asc("abcd")
asc("ABCD")
```

---

barplot_errbar *Plot a barplot graph with error bar on y*

Description

To plot data, just use it as a normal barplot but add the errbar.y values or errbar.y.minus, errbar.y.plus if bars for y axis are asymetric. Use y.plus and y.minus to set absolut limits for error bars. Note that y.plus and y.minus have priority over errbar.y, errbar.y.minus and errbar.y.plus.

Usage

```
barplot_errbar(..., errbar.y = NULL, errbar.y.plus = NULL,
  errbar.y.minus = NULL, y.plus = NULL, y.minus = NULL,
  errbar.tick = 1/50, errbar.lwd = par("lwd"),
  errbar.lty = par("lty"), errbar.col = par("fg"), add = FALSE)
```

Arguments

...	Parameters for barplot() such as main= or ylim=
errbar.y	The length of error bars for y. Recycled if necessary.
errbar.y.plus	The length of positive error bars for y. Recycled if necessary.
errbar.y.minus	The length of negative error bars for y. Recycled if necessary.
y.plus	The absolut position of the positive error bar for y. Recycled if necessary.
y.minus	The absolut position of the nagative error bar for y. Recycled if necessary.
errbar.tick	Size of small ticks at the end of error bars defined as a proportion of total width or height graph size.
errbar.lwd	Error bar line width, see par("lwd")
errbar.lty	Error bar line type, see par("lwd")
errbar.col	Error bar line color, see par("col")
add	If true, add the graph to the previous one.

Details

barplot_errbar plot a barplot with error bar on y

Value

A numeric vector (or matrix, when beside = TRUE), say mp, giving the coordinates of all the bar midpoints drawn, useful for adding to the graph.
If beside is true, use colMeans(mp) for the midpoints of each group of bars, see example.

Author(s)

Marc Girondot

See Also

plot_errorbar

Other plot and barplot functions: ScalePreviousPlot, plot_add, plot_errbar

Examples

```
## Not run:
barplot_errbar(rnorm(10, 10, 3),
xlab="axe x", ylab="axe y", bty="n",
errbar.y.plus=rnorm(10, 1, 0.1), col=rainbow(10),
names.arg=paste("Group",1:10), cex.names=0.6)
y <- rnorm(10, 10, 3)
barplot_errbar(y,
                xlab="axe x", ylab="axe y", bty="n",
            y.plus=y+2)

## End(Not run)
```

cArrows *Draw curved lines with arrowhead*

Description

Draw a curved line with arrowhead.

Usage

```
cArrows(x1, y1, x2, y2, code = 2, size = 1, width = 1.2/4/cin,
  open = TRUE, sh.adj = 0.1, sh.lwd = 1, sh.col = if (is.R())
  par("fg") else 1, sh.lty = 1, h.col = sh.col, h.col.bo = sh.col,
  h.lwd = sh.lwd, h.lty = sh.lty, curved = FALSE,
  beautiful.arrow = 2/3)
```

Arguments

x1	coordinates of points from which to draw.
y1	coordinates of points from which to draw.
x2	coordinates of points to which to draw.
y2	coordinates of points to which to draw.
code	integer code (1, 2, or 3), determining kind of arrows to be drawn.
size	size of the arrowhead.
width	width of the arrowhead.
open	shape of the arrowhead.
sh.adj	Shift the beginning of the line.
sh.lwd	width of the line.
sh.col	color of the line.
sh.lty	type of line.
h.col	color of the arrowhead.
h.col.bo	color of the arrowhead border.
h.lwd	width of the arrowhead.
h.lty	type of line for the arrowhead.
curved	0 is a straigth line, positive of negative value make the line curved.
beautiful.arrow	if open is false, make the arrowhead more beautiful.

Details

cArrows draws curved lines with arrowhead

Value

A list wit lab.x and lab.y being the position where to draw label

Author(s)

Modified from iGraph

Examples

```
plot(c(1, 10), c(1, 10), type="n", bty="n")
cArrows(x1=2, y1=2, x2=6, y2=6, curved=1)
cArrows(x1=2, y1=2, x2=6, y2=6, curved=0)
cArrows(x1=2, y1=2, x2=6, y2=6, curved=1, sh.adj=1)
cArrows(x1=2, y1=2, x2=6, y2=6, curved=-1, open=FALSE)
cArrows(x1=9, y1=2, x2=6, y2=6, curved=-1, open=FALSE, sh.col="red")
cArrows(x1=9, y1=9, x2=6, y2=6, curved=-1, open=FALSE, h.col="red")
cArrows(x1=2, y1=9, x2=6, y2=6, curved=1, open=FALSE, h.col="red", h.col.bo="red")
```

ChangeCoordinate *Return a value in a changed coordinate*

Description

Return a value in a changed coordinate system.

Usage

```
ChangeCoordinate(x = stop("At least one value to convert must be provided"),
initial = stop("Set of two values must be provided as references"),
transformed = stop("Set of two transformed values must be provided"))
```

Arguments

x	value to convert
initial	Set of two values in the original system
transformed	Set of the two values in the converted system

Details

ChangeCoordinate returns a value in a changed coordinate

Value

A value in the new system

Author(s)

Marc Girondot

Examples

```
ChangeCoordinate(x=c(10, 20), initial=c(1, 100), transformed=c(0, 1))
```

---

chr *Return the characters defined by the codes*

Description

Return a string with characters defined by the codes.

Usage

```
chr(n)
```

Arguments

n	The code to be used to return a character

Details

chr returns the characters defined by the codes

Value

A string with characters defined by the codes

Author(s)

Based on this blog: http://datadebrief.blogspot.com/2011/03/ascii-code-table-in-r.html

See Also

Other Characters: asc, d, tnirp

Examples

```
chr(65:75)
chr(unlist(tapply(144:175, 144:175, function(x) {c(208, x)})))
```

clean.knitr *Delete temporary files created during knitr compile*

Description

Delete temporary files created during knitr compile in working directory.
This function works only in UNIX system (LINUX or MacOSX).

Usage

```
clean.knitr()
```

Details

clean.knitr deletes temporary files created during knitr compile

Value

Nothing

Author(s)

Marc Girondot

Examples

```
## Not run:
clean.knitr()

## End(Not run)
```

compare *Run a shiny application for basic functions of comparison*

Description

Run a shiny application for basic functions of comparison.

Usage

```
compare()
```

Details

compare runs a shiny application for basic functions of comparison

Value

Nothing

Author(s)

Marc Girondot

References

Girondot, M., Guillon, J.-M., 2018. The w-value: An alternative to t- and X2 tests. Journal of Biostatistics & Biometrics 1, 1-3.

See Also

Other w-value functions: `contingencyTable.compare`, `series.compare`

Examples

```
## Not run:
library(HelpersMG)
compare()

## End(Not run)
```

`compare_AIC` *Compares the AIC of several outputs*

Description

This function is used to compare the AIC of several outputs obtained with the same data but with different set of parameters.
The parameters must be lists with $aic or $AIC or $value and $par elements or if AIC(element) is defined.
if `$value` and `$par` are present in the object, the AIC is calculated as `2*factor.value*value+2*length(par)`.
If `$value` is -log(likeihood), then factor.value must be 1 and if `$value` is log(likeihood), then factor.value must be -1.
If several objects are within the same list, their AIC are summed.
For example, compare_AIC(g1=list(group), g2=list(separe1, separe2)) can be used to compare a single model onto two different sets of data against each set of data fited with its own set of parameters.
Take a look at `ICtab` in package `bbmle` which is similar.

Usage

```
compare_AIC(..., factor.value = 1, silent = FALSE)
```

Arguments

...	Successive results to be compared as lists.
factor.value	The $value of the list object is multiplied by factor.value to calculate AIC.
silent	If TRUE, nothing is displayed.

Details

compare_AIC compares the AIC of several outputs obtained with the same data.

Value

A list with DeltaAIC and Akaike weight for the models.

Author(s)

Marc Girondot

See Also

Other AIC: ExtractAIC.glm, compare_AICc, compare_BIC

Examples

```
## Not run:
library("HelpersMG")
# Here two different models are fitted
x <- 1:30
y <- rnorm(30, 10, 2)+log(x)
plot(x, y)
d <- data.frame(x=x, y=y)
m1 <- lm(y ~ x, data=d)
m2 <- lm(y ~ log(x), data=d)
compare_AIC(linear=m1, log=m2)
# Here test if two datasets can be modeled with a single model
x2 <- 1:30
y2 <- rnorm(30, 15, 2)+log(x2)
plot(x, y, ylim=c(5, 25))
plot_add(x2, y2, col="red")
d2 <- data.frame(x=x2, y=y2)
m1_2 <- lm(y ~ x, data=d2)
x_grouped <- c(x, x2)
y_grouped <- c(y, y2)
d_grouped <- data.frame(x=x_grouped, y=y_grouped)
m1_grouped <- lm(y ~ x, data=d_grouped)
compare_AIC(separate=list(m1, m1_2), grouped=m1_grouped)

## End(Not run)
```

compare_AICc *Compares the AICc of several outputs*

Description

This function is used to compare the AICc of several outputs obtained with the same data but with different set of parameters.
Each object must have associated `logLik()` method with df and nobs attributes.
AICc for object x will be calculated as `2*factor.value*logLik(x)+(2*attributes(logLik(x))$df*(attributes(logL`

Usage

```
compare_AICc(..., factor.value = -1, silent = FALSE)
```

Arguments

`...`	Successive results to be compared as lists.
`factor.value`	The $value of the list object is multiplied by factor.value to calculate BIC.
`silent`	If TRUE, nothing is displayed.

Details

compare_AICc compares the AICc of several outputs obtained with the same data.

Value

A list with DeltaAICc and Akaike weight for the models.

Author(s)

Marc Girondot

See Also

Other AIC: `ExtractAIC.glm`, `compare_AIC`, `compare_BIC`

Examples

```
## Not run:
library("HelpersMG")
# Here two different models are fitted
x <- 1:30
y <- rnorm(30, 10, 2)+log(x)
plot(x, y)
d <- data.frame(x=x, y=y)
m1 <- lm(y ~ x, data=d)
m2 <- lm(y ~ log(x), data=d)
```

```
compare_BIC(linear=m1, log=m2, factor.value=-1)
# Here test if two datasets can be modeled with a single model
x2 <- 1:30
y2 <- rnorm(30, 15, 2)+log(x2)
plot(x, y, ylim=c(5, 25))
plot_add(x2, y2, col="red")
d2 <- data.frame(x=x2, y=y2)
m1_2 <- lm(y ~ x, data=d2)
x_grouped <- c(x, x2)
y_grouped <- c(y, y2)
d_grouped <- data.frame(x=x_grouped, y=y_grouped)
m1_grouped <- lm(y ~ x, data=d_grouped)
compare_AICc(separate=list(m1, m1_2), grouped=m1_grouped, factor.value=-1)
# Or simply
compare_AICc(m1=list(AICc=100), m2=list(AICc=102))

## End(Not run)
```

`compare_BIC` *Compares the BIC of several outputs*

Description

This function is used to compare the BIC of several outputs obtained with the same data but with different set of parameters.
Each object must have associated `logLik()` method with df and nobs attributes.
BIC for object x will be calculated as `2*factor.value*sum(logLik(x))+sum(attributes(logLik(x))$df)*log(attrib`
When several data (i..n) are included, the global BIC is calculated as:
`2*factor.value*sum(logLik(x)) for i..n+sum(attributes(logLik(x))$df) for i..n*log(attributes(logLik(`

Usage

```
compare_BIC(..., factor.value = -1, silent = FALSE)
```

Arguments

`...`	Successive results to be compared as lists.
`factor.value`	The $value of the list object is multiplied by factor.value to calculate BIC.
`silent`	If TRUE, nothing is displayed.

Details

compare_BIC compares the BIC of several outputs obtained with the same data.

Value

A list with DeltaBIC and Akaike weight for the models.

Author(s)

Marc Girondot

See Also

Other AIC: ExtractAIC.glm, compare_AICc, compare_AIC

Examples

```
## Not run:
library("HelpersMG")
# Here two different models are fitted
x <- 1:30
y <- rnorm(30, 10, 2)+log(x)
plot(x, y)
d <- data.frame(x=x, y=y)
m1 <- lm(y ~ x, data=d)
m2 <- lm(y ~ log(x), data=d)
compare_BIC(linear=m1, log=m2, factor.value=-1)
# Here test if two datasets can be modeled with a single model
x2 <- 1:30
y2 <- rnorm(30, 15, 2)+log(x2)
plot(x, y, ylim=c(5, 25))
plot_add(x2, y2, col="red")
d2 <- data.frame(x=x2, y=y2)
m1_2 <- lm(y ~ x, data=d2)
x_grouped <- c(x, x2)
y_grouped <- c(y, y2)
d_grouped <- data.frame(x=x_grouped, y=y_grouped)
m1_grouped <- lm(y ~ x, data=d_grouped)
compare_BIC(separate=list(m1, m1_2), grouped=m1_grouped, factor.value=-1)

## End(Not run)
```

---

contingencyTable.compare

Contingency table comparison using Akaike weight

Description

This function is used as a replacement of chisq.test() to not use p-value.

Usage

```
contingencyTable.compare(table, criterion = c("AIC", "AICc", "BIC"),
  probs = NULL)
```

Arguments

table	A matrix or a data.frame with series in rows and number of each category in column
criterion	Which criterion is used for model selection
probs	Series of probabilities used for conformity comparison

Details

contingencyTable.compare compares contingency table using Akaike weight.

Value

The probability that a single proportion model is sufficient to explain the data

Author(s)

Marc Girondot

References

Girondot, M., Guillon, J.-M., 2018. The w-value: An alternative to t- and X2 tests. Journal of Biostatistics & Biometrics 1, 1-4.

See Also

Other w-value functions: compare, series.compare

Examples

```
## Not run:
library("HelpersMG")

# Symmetry of Lepidochelys olivacea scutes
table <- t(data.frame(SriLanka=c(200, 157), AfricaAtl=c(19, 12),
                      Guyana=c(8, 6), Suriname=c(162, 88),
                      MexicoPac1984=c(42, 34), MexicoPac2014Dead=c(8, 9),
                      MexicoPac2014Alive=c(13, 12),
                      row.names =c("Symmetric", "Asymmetric")))
table
contingencyTable.compare(table)

table <- t(data.frame(SriLanka=c(200, 157), AfricaAtl=c(19, 12), Guyana=c(8, 6),
                      Suriname=c(162, 88), MexicoPac1984=c(42, 34),
                      MexicoPac2014Dead=c(8, 9),
                      MexicoPac2014Alive=c(13, 12), Lepidochelys.kempii=c(99, 1),
                      row.names =c("Symmetric", "Asymmetric")))
table
contingencyTable.compare(table)

# Conformity to a model
```

```
table <- matrix(c(33, 12, 25, 75), ncol = 2, byrow = TRUE)
probs <- c(0.5, 0.5)
contingencyTable.compare(table, probs=probs)

# Conformity to a model
table <- matrix(c(33, 12), ncol = 2, byrow = TRUE)
probs <- c(0.5, 0.5)
contingencyTable.compare(table, probs=probs)

# Conformity to a model
table <- matrix(c(33, 12, 8, 25, 75, 9), ncol = 3, byrow = TRUE)
probs <- c(0.8, 0.1, 0.1)
contingencyTable.compare(table, probs=probs)

# Comparison of chisq.test() and this function
table <- matrix(c(NA, NA, 25, 75), ncol = 2, byrow = TRUE)

pv <- NULL
aw <- NULL
par(new=FALSE)
n <- 100

for (GroupA in 0:n) {
  table[1, 1] <- GroupA
  table[1, 2] <- n-GroupA
  pv <- c(pv, chisq.test(table)$p.value)
  aw <- c(aw, contingencyTable.compare(table, criterion="BIC")[1])
}

x <- 0:n
y <- pv
y2 <- aw
plot(x=x, y=y, type="l", bty="n", las=1, xlab="Number of type P in Group B", ylab="Probability",
     main="", lwd=2)
lines(x=x, y=y2, type="l", col="red", lwd=2)

# w-value
(l1 <- x[which(aw>0.05)[1]])
(l2 <- rev(x)[which(rev(aw)>0.05)[1]])

aw[l1]
pv[l1]

aw[l2+2]
pv[l2+2]

# p-value
l1 <- which(pv>0.05)[1]
l2 <- max(which(pv>0.05))

aw[l1]
pv[l1]
```

```
aw[l2]
pv[l2]

y[which(y2>0.05)[1]]
y[which(rev(y2)>0.05)[1]]

par(xpd=TRUE)
text(x=25, y=1.15, labels="Group A: 25 type P / 100", pos=1)

segments(x0=25, y0=0, x1=25, y1=1, lty=3)

# plot(1, 1)

v1 <- c(expression(italic("p")*"-value"), expression("after "*chi^2*"-test"))
v2 <- c(expression(italic("w")*"-value for A"), expression("and B identical models"))
legend("topright", legend=c(v1, v2),
       y.intersp = 1,
       col=c("black", "black", "red", "red"), bty="n", lty=c(1, 0, 1, 0))

segments(x0=0, x1=n, y0=0.05, y1=0.05, lty=2)
text(x=101, y=0.05, labels = "0.05", pos=4)

## End(Not run)
```

`convert.tz` *Convert one Date-Time from one timezone to another*

Description

Convert one Date-Time from one timezone to another.
Available timezones can be shown using OlsonNames().

Usage

```
convert.tz(x, tz = Sys.timezone())
```

Arguments

x	The date-time in POSIXlt or POSIXct format
tz	The timezone

Details

convert.tz Convert one Date-Time from one timezone to another

Value

A POSIXlt or POSIXct date converted

Author(s)

Marc Girondot

See Also

Function `with_tz()` from `lubridate` package does the same. I keep it here only for compatibility with old scripts.

Examples

```
d <- as.POSIXlt("2010-01-01 17:34:20", tz="UTC")
convert.tz(d, tz="America/Guatemala")
```

---

d *Write an ASCII Representation of an Object*

Description

Writes an ASCII text representation of an R object.
It can be used as a replacement of dput() for named vectors.
The controls "keepNA", "keepInteger" and "showAttributes" are utilized for named vectors.

Usage

```
d(x, file = "", control = c("keepNA", "keepInteger", "showAttributes"),
  collapse = ", \n  ")
```

Arguments

`x`	A named vector object
`file`	either a character string naming a file or a connection. "" indicates output to the console.
`control`	character vector indicating deparsing options. See .deparseOpts for their description.
`collapse`	Characters used to separate values.

Details

d Write an ASCII Representation of an Object

Value

A string

Author(s)

Marc Girondot

See Also

Other Characters: asc, chr, tnirp

Examples

```
d(c(A=10, B=20))
dput(c(A=10, B=20))
```

DIx *Return an index of quantitative asymmetry and complexity named Developmental Instability Index (DIx)*

Description

Return an index of quantitative asymmetry and complexity.
Higher is the value, higher is the complexity (number of objects) and diversity (difference between them).
The indice is based on the product of the average angular distance of Edwards (1971) for all permutations of measures for both sides with the geometric mean of the inverse of Shannon entropy H for both sides. Let p1 and p2 two vectors of relative measures of objects with sum(p1) = 1 and sum(p2)=1 and n1 being the number of objects in p1 and n2 being the number of objects in p2.
Edwards distance for all permutations of p1 and p2 objects are computed and the average value E is calculated.
The maximun possible Shannon index for identical n1 is max1 = sum((1/n1) * log(1/n1)).
Shannon index is v1 = sum(p1 * log(p1)).
If version == 2, the complementary of Shannon index for these n1 objects is used: c1 = 2 * max1 - v1
If version == 1, the Shannon index is used directly.
The geometry mean between both sides defined the measure of diversity within each side: S=sqrt(c1 * c2)
The Developmental Instability Index is then S * E

Usage

```
DIx(l1, l2, details = FALSE, version = 1)
```

Arguments

l1	Set of measures at one side of an organism
l2	Set of measures at the other side of an organism
details	If TRUE, will show the details of computing
version	Can be 1 or 2; see description

Details

DIx returns an index of quantitative asymmetry and complexity

Value

A numeric value

Author(s)

Marc Girondot

References

Edwards, A.W.F., 1971. Distances between populations on the basis of gene frequencies. Biometrics 27, 873–881.
Shannon C.E. 1948 A mathematical theory of communication. Bell System Technical Journal 27(3), 379-423.

Examples

```
## Not run:
l1 <- c(0.1, 0.1, 0.05, 0.2, 0.3, 0.25)
l2 <- c(0.2, 0.3, 0.5)
DIx(l1, l2)

l1 <- c(0.1, 0.1, 0.05, 0.2, 0.3, 0.25)
l2 <- c(0.1, 0.1, 0.05, 0.2, 0.3, 0.25)
DIx(l1, l2)

l1 <- c(0.2, 0.3, 0.5)
l2 <- c(0.2, 0.3, 0.5)
DIx(l1, l2)

l1 <- c(0.2, 0.2, 0.2, 0.2, 0.2)
l2 <- c(0.2, 0.3, 0.5)
DIx(l1, l2)

l1 <- c(0.2, 0.2, 0.2, 0.2, 0.2)
l2 <- c(0.3333, 0.3333, 0.3333)
DIx(l1, l2)

l1 <- c(0.2, 0.2, 0.2, 0.2, 0.2)
l2 <- c(0.2, 0.2, 0.2, 0.2, 0.2)
DIx(l1, l2)

l1 <- c(0.3333, 0.3333, 0.3333)
l2 <- c(0.3333, 0.3333, 0.3333)
DIx(l1, l2)

## End(Not run)
```

dSnbinom *Density for the sum of random variable with negative binomial distributions.*

Description

Density for the sum of random variable with negative binomial distributions.
If all prob values are the same, infinite is automatically set to 0.

Usage

```
dSnbinom(x = stop("You must provide a x value"), size = NULL,
  prob = NULL, mu = NULL, log = FALSE, infinite = 100)
```

Arguments

x	vector of (non-negative integer) quantiles.
size	target for number of successful trials, or dispersion parameter (the shape parameter of the gamma mixing distribution). Must be strictly positive, need not be integer.
prob	probability of success in each trial. 0 < prob <= 1.
mu	alternative parametrization via mean.
log	logical; if TRUE, probabilities p are given as log(p).
infinite	Number of maximal iterations; check different values to determine the error in estimation.

Details

dSnbinom returns the density for the sum of random variable with negative binomial distributions

Value

dSnbinom gives the density

Author(s)

Marc Girondot

References

Furman, E., 2007. On the convolution of the negative binomial random variables. Statistics & Probability Letters 77, 169-172.

See Also

Other Distribution of sum of random variable with negative binomial distributions: pSnbinom, qSnbinom, rSnbinom

Examples

```
## Not run:
alpha <- c(1, 2, 5, 1, 2)
p <- c(0.1, 0.12, 0.13, 0.14, 0.14)
# Test with lower iterations: 2 or 50 rather than 10 [default]; precision is very good still with 10
dSnbinom(20, size=alpha, prob=p, infinite=50)
dSnbinom(20, size=alpha, prob=p, infinite=10)
dSnbinom(20, size=alpha, prob=p, infinite=2)
# However it is not always the case; It depends on the parametrization (see Furman 2007)
dSnbinom(20, size=2, mu=c(0.01, 0.02, 0.03), infinite=1000)
dSnbinom(20, size=2, mu=c(0.01, 0.02, 0.03), infinite=100)
dSnbinom(20, size=2, mu=c(0.01, 0.02, 0.03), infinite=50)
dSnbinom(20, size=2, mu=c(0.01, 0.02, 0.03), infinite=10)
dSnbinom(20, size=2, mu=c(0.01, 0.02, 0.03), infinite=2)
# Test with a single distribution
dSnbinom(20, size=1, mu=20)
# when only one distribution is available, it is the same as dnbinom()
dnbinom(20, size=1, mu=20)
# If a parameter is supplied as only one value, it is supposed to be constant
dSnbinom(20, size=1, mu=c(14, 15, 10))
# The function is vectorized:
plot(0:200, dSnbinom(0:200, size=alpha, prob=p), bty="n", type="h", xlab="x", ylab="Density")
# Comparison with simulated distribution using rep replicates
alpha <- c(2.1, 2.05, 2)
mu <- c(10, 30, 20)
rep <- 100000
distEmpirique <- rSnbinom(rep, size=alpha, mu=mu)
tabledistEmpirique <- rep(0, 301)
names(tabledistEmpirique) <- as.character(0:300)
tabledistEmpirique[names(table(distEmpirique))] <- table(distEmpirique)/rep

plot(0:300, dSnbinom(0:300, size=alpha, mu=mu, infinite=1000), type="h", bty="n",
   xlab="x", ylab="Density", ylim=c(0,0.02))
plot_add(0:300, tabledistEmpirique, type="l", col="red")
legend(x=200, y=0.02, legend=c("Empirical", "Theoretical"),
   text.col=c("red", "black"), bty="n")

# Example with the approximation mu=mean(mu)
plot(0:300, dSnbinom(0:300, size=alpha, mu=mu, infinite=0), type="h", bty="n",
   xlab="x", ylab="Density", ylim=c(0,0.02))
plot_add(0:300, tabledistEmpirique, type="l", col="red")
legend(x=200, y=0.02, legend=c("Empirical", "Theoretical"),
   text.col=c("red", "black"), bty="n")

# example to fit the distribution
data <- rnbinom(1000, size=1, mu=10)
hist(data)
ag <- rep(1:100, 10)
r <- aggregate(data, by=list(ag), FUN=sum)
hist(r[,2])

parx <- c(size=1, mu=10)
```

```
dSnbinomx <- function(x, par) {
  -sum(dSnbinom(x=x[,2], mu=rep(par["mu"], 10), size=par["size"], log=TRUE,
                infinite = 1000))
}

fit_mu_size <- optim(par = parx, fn=dSnbinomx, x=r, method="BFGS", control=c(trace=TRUE))
fit_mu_size$par

## End(Not run)
```

`duplicate.packages` *List the duplicated packages with their locations*

Description

A data.frame with the duplicated packages and their locations and version.
The columns Lib1 and Version1 should have the oldest version of the packages. Then you can try:
li <- duplicate.packages() if (nrow(li) != 0) for (i in 1:nrow(li)) remove.packages(rownames(li)[i], lib=li[i, "Lib1"])

Usage

```
duplicate.packages()
```

Details

duplicate.packages lists the duplicated packages with their locations

Value

A data.frame with 4 elements for each duplicated packages:
- versions: the version of the packages
- libraries: the locations

Author(s)

Marc Girondot

Examples

```
## Not run:
library(HelpersMG)
duplicate.packages()

## End(Not run)
```

`ellipse` *Plot an ellipse*

Description

Plot a ellipse dined by the center and the radius. The options for binomial confidence are:
- alpha is 1 - confidence interval
- method must be one of these "wilson", "exact", "asymptotic"
col parameter can be a list of colors. See examples

Usage

```
ellipse(center.x = 0, center.y = 0, radius.x = 1, radius.y = 1,
  radius.x.lower = NULL, radius.x.upper = NULL,
  radius.y.lower = NULL, radius.y.upper = NULL, alpha = 0,
  binconf.x = NULL, binconf.y = NULL, control.binconf = list(alpha =
  0.05, method = "wilson"), length = 100, ...)
```

Arguments

`center.x`	Center of the ellipse on x axis
`center.y`	Center of the ellipse on y axis
`radius.x`	Radius along the x axis
`radius.y`	Radius along the y axis
`radius.x.lower`	Radius along the x axis, at left of center
`radius.x.upper`	Radius along the x axis, at right of center
`radius.y.lower`	Radius along the y axis, at bottom of center
`radius.y.upper`	Radius along the y axis, at top of center
`alpha`	Rotation in radians
`binconf.x`	A data.frame or a matrix with two columns, x and n or with three columns, PointEst, Lower, and Upper
`binconf.y`	A data.frame or a matrix with two columns, x and n or with three columns, PointEst, Lower, and Upper
`control.binconf`	A list with options for binomial confidence
`length`	Number of points to draw the ellipse
`...`	Graphical parameters

Details

ellipse plots an ellipse

Value

Nothing

Author(s)

marc.girondot@u-psud.fr

Examples

```
plot(0:1, 0:1, xlim=c(0, 1), ylim=c(0,1), lty=2, type="l", las=1, bty="n",
     xlab="Variable x", ylab="variable y")

ellipse(center.x = c(0.2, 0.3, 0.25), center.y = c(0.7, 0.6, 0.55),
        radius.x = c(0.1, 0.1, 0.1), radius.y = c(0.15, 0.2, 0.4),
        border=NA, col=rgb(red = 0.1, green = 0.1, blue = 0.1, alpha = 0.1))

ellipse(center.x = 0.5, center.y = 0.5,
        radius.x.lower = 0.1, radius.x.upper = 0.3,
        radius.y = 0.2,
        border=NA, col=rgb(red = 0.1, green = 0.1, blue = 0.1, alpha = 0.1))

ellipse(center.x = 0.6, center.y = 0.3,
        radius.x.lower = 0.3, radius.x.upper = 0.3,
        radius.y.lower = 0.2, radius.y.upper = 0.4,
        border=NA, col=rgb(red = 0.1, green = 0.1, blue = 0.1, alpha = 0.1))

plot(0:1, 0:1, xlim=c(0, 1), ylim=c(0,1), lty=2, type="l", bty="n", asp=1,
     xlab="Variable x", ylab="variable y", axes=FALSE)
axis(1, at=c(0, 0.25, 0.5, 0.75, 1))
axis(2, at=c(0, 0.25, 0.5, 0.75, 1), las=1)

ellipse(center.x = 0.5, center.y = 0.5, radius.x = 0.2, radius.y = 0.4,
       border=NA, col=rgb(red = 0.1, green = 0.1, blue = 0.1, alpha = 0.1))
ellipse(center.x = 0.5, center.y = 0.5, radius.x = 0.2, radius.y = 0.4,
       border=NA, col=rgb(red = 0.1, green = 0.1, blue = 0.1, alpha = 0.1), alpha = pi/4)

plot(0:1, 0:1, xlim=c(0, 1), ylim=c(0,1), lty=2, type="l", las=1, bty="n",
     xlab="Variable x", ylab="variable y")

for (k in 0:8)
  ellipse(center.x=0.5, center.y=0.5, radius.x=0.1, radius.y=0.4,
          alpha=seq(from=0, to=pi/4, length=9)[k],
          border=rainbow(9)[k])

# Exemple with confidence of proportions
males <- c(10, 25, 3, 4)
N <- c(12, 52, 17, 10)

males2 <- c(12, 20, 3, 6)
N2 <- c(15, 50, 20, 12)

plot(0:1, 0:1, xlim=c(0, 1), ylim=c(0,1), lty=2, type="l", las=1, bty="n",
```

```
     xlab="Variable x", ylab="variable y")

ellipse(binconf.x = data.frame(x=males, n=N), binconf.y = data.frame(x=males2, n=N2),
        border=NA, col=rgb(red = 0.1, green = 0.5, blue = 0.1, alpha = 0.1))

plot(0:1, 0:1, xlim=c(0, 1), ylim=c(0,1), lty=2, type="l", las=1, bty="n",
     xlab="Variable x", ylab="variable y")

ellipse(binconf.x = data.frame(x=males, n=N),
        binconf.y = data.frame(PointEst=c(0.1, 0.2, 0.3, 0.5),
                               Lower=c(0.02, 0.12, 0.25, 0.30),
                               Upper=c(0.18, 0.29, 0.35, 0.67)),
        border=NA, col=rgb(red = 0.1, green = 0.5, blue = 0.1, alpha = 0.1))

# Examples with a gradient
plot(0:1, 0:1, xlim=c(0, 1), ylim=c(0,1), lty=2, type="l", las=1, bty="n",
     xlab="Variable x", ylab="variable y")
ellipse(center.x = 0.6, center.y = 0.3,
        radius.x.lower = 0.3, radius.x.upper = 0.3,
        radius.y.lower = 0.2, radius.y.upper = 0.4,
        border=NA, col=grey.colors(100, alpha = 0.1))

plot(0:1, 0:1, xlim=c(0, 1), ylim=c(0,1), lty=2, type="l", las=1, bty="n",
     xlab="Variable x", ylab="variable y")
ellipse(binconf.x = data.frame(x=males, n=N), binconf.y = data.frame(x=males2, n=N2),
        border=NA, col=grey.colors(100, alpha = 0.1))
```

`ExtractAIC.glm` *Return AIC, AICc or BIC from a glm object*

Description

For glm fits the family's aic() function is used to compute the AIC.
The choice between different criteria is done by setting a global option AIC. It can be checked using show.option=TRUE. Indeed, it is not possible to use the ... parameter due to a bug in some functions of MASS package. If you want to use this function as a replacement for setpAIC(), do extractAIC.glm <- ExtractAIC.glm before.

Usage

```
ExtractAIC.glm(fit, scale = 0, k = 2, ...)
```

Arguments

`fit`	fitted model, the result of a fitter glm.
`scale`	unused for glm.
`k`	numeric specifying the 'weight' of the equivalent degrees of freedom (=: edf) part in the AIC formula.

... further arguments (currently unused because addterm.glm and dropterm.glm using this function do not transmit them).

Details

ExtractAIC.glm returns AIC, AICc or BIC from a glm object

Value

A numeric named vector of length 2, with first and second elements giving
edf the 'equivalent degrees of freedom' for the fitted model fit.
x the Information Criterion for fit.

Author(s)

Modified from stats:::extract.AIC.glm

See Also

Other AIC: compare_AICc, compare_AIC, compare_BIC

Examples

```
extractAIC.glm <- ExtractAIC.glm
n <- 100
x <- rnorm(n, 20, 2)
A <- rnorm(n, 20, 5)
g <- glm(x ~ A)
extractAIC(g, show.option=TRUE)
options(AIC="AIC")
extractAIC(g)
options(AIC="BIC")
extractAIC(g)
options(AIC="AICc")
extractAIC(g)
```

---

growlnotify *Send growl notification for MacOs X system.*

Description

This function is used to send a notification to MacOS user.

Usage

```
growlnotify(textinfo = "")
```

Arguments

textinfo	Text to display in the growlnotify window

Details

growlnotify send growl notification for MacOs X systems.

Value

None

Author(s)

Marc Girondot

Examples

```
## Not run:
# If growlnotify is used on a non-mac system, it just quits.
growlnotify("It works if you are on a Mac with GrowlNotify installed!")

## End(Not run)
```

IC_clean_data *Clean the dataframe before to be used with IC_threshold_matrix*

Description

This function must be used if missing values are present in the dataset.
It ensures that all correlations and partial correlations can be calculated. The columns of the dataframe are removed one per one until all can be calculated without error. It is possible to say that one or more columns must be retained because they are of particular importance in the analysis. The use and method parameters are used by cor() function. The function uses by default a parallel computing in Unix or MacOSX systems. If progress is TRUE and the package pbmcapply is present, a progress bar is displayed. If debug is TRUE, some informations are shown during the process. https://fr.wikipedia.org/wiki/Iconographie_des_corrélations

Usage

```
IC_clean_data(data = stop("A dataframe object is required"),
  use = c("pairwise.complete.obs", "everything", "all.obs",
  "complete.obs", "na.or.complete"), method = c("pearson", "kendall",
  "spearman"), variable.retain = NULL, test.partial.correlation = TRUE,
  progress = TRUE, debug = FALSE)
```

Arguments

data	The data.frame to be cleaned
use	an optional character string giving a method for computing covariances in the presence of missing values. This must be (an abbreviation of) one of the strings "everything", "all.obs", "complete.obs", "na.or.complete", or "pairwise.complete.obs".

method	a character string indicating which correlation coefficient (or covariance) is to be computed. One of "pearson" (default), "kendall", or "spearman": can be abbreviated.
variable.retain	a vector with the name of columns to keep
test.partial.correlation	should the partial correlations be tested ?
progress	show a progress bar
debug	if TRUE, information about progression of cleaning are shown

Details

IC_clean_data checks and corrects the dataframe to be used with IC_threshold_matrix

Value

A dataframe

Author(s)

Marc Girondot

References

Lesty, M., 1999. Une nouvelle approche dans le choix des régresseurs de la régression multiple en présence d'interactions et de colinéarités. Revue de Modulad 22, 41-77.

See Also

Other Iconography of correlations: IC_correlation_simplify, IC_threshold_matrix, plot.IconoCorel

Examples

```
## Not run:
library("HelpersMG")
es <- matrix(c("e1", "52", "12", "12", "5",
"e2", "59", "12.5", "9", "5",
"e3", "55", "13", "15", "9",
"e4", "58", "14.5", "5", "5",
"e5", "66", "15.5", "11", "13.5",
"e6", "62", "16", "15", "18",
"e7", "63", "17", "12", "18",
"e8", "69", "18", "9", "18"), ncol=5, byrow = TRUE)
colnames(es) <- c("Élève", "Poids", "Âge", "Assiduité", "Note")
es <- as.data.frame(es, stringsasFactor=FALSE)
es[, 2] <- as.numeric(as.character(es[, 2]))
es[, 3] <- as.numeric(as.character(es[, 3]))
es[, 4] <- as.numeric(as.character(es[, 4]))
es[, 5] <- as.numeric(as.character(es[, 5]))
```

```
es

df <- IC_clean_data(es, debug = TRUE)
cor_matrix <- IC_threshold_matrix(data=df, threshold = NULL, progress=FALSE)
cor_threshold <- IC_threshold_matrix(data=df, threshold = 0.3)
par(mar=c(1,1,1,1))
set.seed(4)
plot(cor_threshold)
cor_threshold_Note <- IC_correlation_simplify(matrix=cor_threshold, variable="Note")
plot(cor_threshold_Note)

## End(Not run)
```

`IC_correlation_simplify` *Simplify the correlation matrix*

Description

This function can be used to simplify the network of correlations.
If no vector of variables is given, the variables not linked to any other variable are removed. If a vector of variables is given, only link to these variables are retained. https://fr.wikipedia.org/wiki/Iconographie_des_c

Usage

```
IC_correlation_simplify(matrix, variable = NULL)
```

Arguments

`matrix`	The correlation matrix to simplify
`variable`	a vector with the name of columns to keep

Details

IC_correlation_simplify simplifies the correlation matrix

Value

A list

Author(s)

Marc Girondot

References

Lesty, M., 1999. Une nouvelle approche dans le choix des régresseurs de la régression multiple en présence d'interactions et de colinéarités. Revue de Modulad 22, 41-77.

See Also

Other Iconography of correlations: IC_clean_data, IC_threshold_matrix, plot.IconoCorel

Examples

```
## Not run:
library("HelpersMG")
es <- matrix(c("e1", "52", "12", "12", "5",
"e2", "59", "12.5", "9", "5",
"e3", "55", "13", "15", "9",
"e4", "58", "14.5", "5", "5",
"e5", "66", "15.5", "11", "13.5",
"e6", "62", "16", "15", "18",
"e7", "63", "17", "12", "18",
"e8", "69", "18", "9", "18"), ncol=5, byrow = TRUE)
colnames(es) <- c("Élève", "Poids", "Âge", "Assiduité", "Note")
es <- as.data.frame(es, stringsasFactor=FALSE)
es[, 2] <- as.numeric(as.character(es[, 2]))
es[, 3] <- as.numeric(as.character(es[, 3]))
es[, 4] <- as.numeric(as.character(es[, 4]))
es[, 5] <- as.numeric(as.character(es[, 5]))

es

df <- IC_clean_data(es, debug = TRUE)
cor_matrix <- IC_threshold_matrix(data=df, threshold = NULL, progress=FALSE)
cor_threshold <- IC_threshold_matrix(data=df, threshold = 0.3)
par(mar=c(1,1,1,1))
set.seed(4)
plot(cor_threshold)
cor_threshold_Note <- IC_correlation_simplify(matrix=cor_threshold, variable="Note")
plot(cor_threshold_Note)

## End(Not run)
```

IC_threshold_matrix *Calculate correlation matrix*

Description

This function calculates the matrix of correlations thresholded using partial correlation.
If the threshold is not given, the object that is produced can be used later for thresholding.
For model OAT: a correlation is retained if it is higher that the threshold and if all partial correlationw of the two variables and any third one are all lower than the threshold.
For model AAT: a correlation is retained if it is higher than the threshold and the partial correlation is lower than the threshold. In this case, no missing value is accepted.
The use and method parameters are used by cor() function. The function uses by default a parallel computing in Unix or MacOSX systems. If progress is TRUE and the package pbmcapply is present, a progress bar is displayed. If debug is TRUE, some informations are shown during the

process but parallel computing is not used.
https://fr.wikipedia.org/wiki/Iconographie_des_corrélations

Usage

```
IC_threshold_matrix(data = stop("A dataframe or an IconoCorel object is required"),
threshold = NULL, use = c("pairwise.complete.obs", "everything",
"all.obs", "complete.obs", "na.or.complete"), method = c("pearson",
"kendall", "spearman"), model = c("OAT", "ATT"), progress = TRUE,
debug = FALSE)
```

Arguments

data	A dataframe or an IconoCorel object from a previous run of IC_threshold_matrix
threshold	threshold for partial and full correlations
use	an optional character string giving a method for computing covariances in the presence of missing values. This must be (an abbreviation of) one of the strings "everything", "all.obs", "complete.obs", "na.or.complete", or "pairwise.complete.obs".
method	a character string indicating which correlation coefficient (or covariance) is to be computed. One of "pearson" (default), "kendall", or "spearman": can be abbreviated.
model	a character string indicating if linear model uses all variables at a time (AAT) or one at a time (OAT).
progress	show a progress bar
debug	display information about progression of computing

Details

IC_threshold_matrix calculates correlation matrix thresholed by partial correlation

Value

A list

Author(s)

Marc Girondot

References

Lesty, M., 1999. Une nouvelle approche dans le choix des régresseurs de la régression multiple en présence d'interactions et de colinéarités. Revue de Modulad 22, 41-77.

See Also

Other Iconography of correlations: IC_clean_data, IC_correlation_simplify, plot.IconoCorel

Examples

```
## Not run:
library("HelpersMG")
es <- matrix(c("e1", "52", "12", "12", "5",
"e2", "59", "12.5", "9", "5",
"e3", "55", "13", "15", "9",
"e4", "58", "14.5", "5", "5",
"e5", "66", "15.5", "11", "13.5",
"e6", "62", "16", "15", "18",
"e7", "63", "17", "12", "18",
"e8", "69", "18", "9", "18"), ncol=5, byrow = TRUE)
colnames(es) <- c("Élève", "Poids", "Âge", "Assiduité", "Note")
es <- as.data.frame(es, stringsasFactor=FALSE)
es[, 2] <- as.numeric(as.character(es[, 2]))
es[, 3] <- as.numeric(as.character(es[, 3]))
es[, 4] <- as.numeric(as.character(es[, 4]))
es[, 5] <- as.numeric(as.character(es[, 5]))

es

df <- IC_clean_data(es, debug = TRUE)
cor_matrix <- IC_threshold_matrix(data=df, threshold = NULL, progress=FALSE)
cor_threshold <- IC_threshold_matrix(data=cor_matrix, threshold = 0.3)
par(mar=c(1,1,1,1))
set.seed(4)
plot(cor_threshold)
cor_threshold_Note <- IC_correlation_simplify(matrix=cor_threshold, variable="Note")
plot(cor_threshold_Note)

# Using the model All at a time

cor_threshold_AAT <- IC_threshold_matrix(data=df, threshold = 0.3, model="AAT")
par(mar=c(1,1,1,1))
set.seed(4)
plot(cor_threshold_AAT, show.legend.strength="bottomleft")



############
dta <- structure(list(Élève = structure(1:8, .Label = c("e1", "e2",
"e3", "e4", "e5", "e6", "e7", "e8"), class = "factor"), Poids = c(52L,
59L, 55L, 58L, 66L, 62L, 63L, 69L), Âge = c(12, 12.5, 13, 14.5,
15.5, 16, 17, 18), Assiduité = c(12L, 9L, 15L, 5L, 11L, 15L,
12L, 9L), Note = c(5, 5, 9, 5, 13.5, 18, 18, 18), e1 = c(1L,
0L, 0L, 0L, 0L, 0L, 0L, 0L), e2 = c(0L, 1L, 0L, 0L, 0L, 0L, 0L,
0L), e3 = c(0L, 0L, 1L, 0L, 0L, 0L, 0L, 0L), e4 = c(0L, 0L, 0L,
1L, 0L, 0L, 0L, 0L), e5 = c(0L, 0L, 0L, 0L, 1L, 0L, 0L, 0L),
    e6 = c(0L, 0L, 0L, 0L, 0L, 1L, 0L, 0L), e7 = c(0L, 0L, 0L,
    0L, 0L, 0L, 1L, 0L), e8 = c(0L, 0L, 0L, 0L, 0L, 0L, 0L, 1L
    )), .Names = c("Élève", "Poids", "Âge", "Assiduité",
"Note", "e1", "e2", "e3", "e4", "e5", "e6", "e7", "e8"), class = "data.frame", row.names = c(NA,
-8L))
```

```
dta0 <- dta[, 2:ncol(dta)]
ic0 <- IC_threshold_matrix(data = dta0)
cor_threshold <- IC_threshold_matrix(data=ic0, threshold = 0.3)
par(mar=c(1,1,1,1))
set.seed(4)
library("igraph")

plot(cor_threshold, vertex.color="red", show.legend.strength = FALSE)
plot(IC_correlation_simplify(matrix=cor_threshold),
     show.legend.strength = FALSE, show.legend.direction = FALSE)


## End(Not run)
```

`index.periodic` *Estimate indices in periodic timeseries based on anchored minimum and maximum*

Description

Estimate indices in periodic timeseries based on anchored minimum and maximum.
The data.frame minmax can be generated manually. It should have three columns (time, index, SD), with all the successive minimum and maximum indices.
It can be used with sun.info() to get the time of minimum and maximum air temperature or with getTide() to reconstruct the sea level.

Usage

```
index.periodic(minmax, time = NULL, replicates = 100,
  progressbar = FALSE)
```

Arguments

`minmax`	A data.frame returned by minmax.periodic
`time`	The time at which produced the estimate
`replicates`	Number of replicates to estimate SD
`progressbar`	Does a progression bar must be shown

Details

index.periodic estimate indices in periodic timeseries based on anchored minimum and maximum

Value

A data.frame with a column time and a column index

Author(s)

Marc Girondot <marc.girondot@u-psud.fr>

See Also

Other Periodic patterns of indices: minmax.periodic, moon.info, sun.info, tide.info

Examples

```
## Not run:
# Generate a timeserie of time
time.obs <- NULL
for (i in 0:9) time.obs <- c(time.obs, c(0, 6, 12, 18)+i*24)
# For these time, generate a timeseries of temperatures
temp.obs <- rep(NA, length(time.obs))
temp.obs[3+(0:9)*4] <- rnorm(10, 25, 3)
temp.obs[1+(0:9)*4] <- rnorm(10, 10, 3)
for (i in 1:(length(time.obs)-1))
  if (is.na(temp.obs[i]))
  temp.obs[i] <- mean(c(temp.obs[i-1], temp.obs[i+1]))
  if (is.na(temp.obs[length(time.obs)]))
  temp.obs[length(time.obs)] <- temp.obs[length(time.obs)-1]/2
observed <- data.frame(time=time.obs, temperature=temp.obs)
# Search for the minimum and maximum values
r <- minmax.periodic(time.minmax.daily=c(Min=2, Max=15),
observed=observed, period=24, colname.index="temperature")

# Estimate all the temperatures for these values
t <- index.periodic(minmax=r)

plot_errbar(x=t[,"time"], y=t[,"index"],
errbar.y=ifelse(is.na(t[,"sd"]), 0, 2*t[,"sd"]),
type="l", las=1, bty="n", errbar.y.polygon = TRUE,
xlab="hours", ylab="Temperatures", ylim=c(0, 35),
errbar.y.polygon.list = list(col="grey"))

plot_add(x=t[,"time"], y=t[,"index"], type="l")

plot_add(observed$time, observed$temperature, pch=19, cex=0.5)

## End(Not run)
```

---

ind_long_lat *Return or the index in ncdf object from lat/longitude or inverse*

Description

Return or the index in ncdf object from lat/longitude or reverse.

Usage

```
ind_long_lat(ncdf = stop("The ncdf data must be supplied"),
  long = NULL, lat = NULL, indice.long = NULL, indice.lat = NULL,
  name.lon = "lon", name.lat = "lat")
```

Arguments

ncdf	An object read from package ncdf4, ncdf or RNetCDF
long	Longitude in decimal format
lat	Latitude in decimal format
indice.long	Index of longitude
indice.lat	Index of latitude
name.lon	Name of argument for longitude, default is lon
name.lat	Name of argument for latitude, default is lat

Details

ind_long_lat is used to manage ncdf information

Value

Or the index in ncdf object from lat/longitude or inverse

Author(s)

Marc Girondot

Examples

```
## Not run:
url <- "ftp://ftp.cdc.noaa.gov/Datasets/noaa.oisst.v2.highres/"
url <- paste0(url, "sst.day.mean.2012.v2.nc")
dest <- paste(Sys.getenv("HOME"), "/sst.day.mean.2012.v2.nc", sep="")
download.file(url, dest)
library("ncdf4")
dta2012 <- nc_open(dest)
indices <- ind_long_lat(ncdf=dta2012, lat=5.89, long=-20.56)
coordinates <- ind_long_lat(ncdf=dta2012, indice.lat=20, indice.long=30)
# library("RNetCDF")
# dta2012 <- open.nc(dest)
# indices <- ind_long_lat(ncdf=dta2012, lat=5.89, long=-20.56)
# coordinates <- ind_long_lat(ncdf=dta2012, indice.lat=20, indice.long=30)
# ncdf library is depreciated in CRAN
# library("ncdf")
# dta2012 <- open.ncdf(dest)
# indices <- ind_long_lat(ncdf=dta2012, lat=5.89, long=-20.56)
# coordinates <- ind_long_lat(ncdf=dta2012, indice.lat=20, indice.long=30)

## End(Not run)
```

`inside.search` *Search a string within files of a folder*

Description

Search for a string inside the files of a folder and return where the string is found.
The pattern for files that must be included uses regex for filtering.

Usage

```
inside.search(path = ".", pattern = "*\\.R$",
  showallfilenames = FALSE, ..., fixed = TRUE, ignore.case = FALSE,
  text = stop("A text to be searched for is necessary"))
```

Arguments

`path`	Path of the folder to search in
`pattern`	Pattern for file names to search in
`showallfilenames`	logical. Show all the filenames search for in
`...`	Options for readLines(), example warn = FALSE
`fixed`	logical. If TRUE, pattern is a string to be matched as is. Overrides all conflicting arguments (see gsub)
`ignore.case`	logical. if FALSE, the pattern matching for text is case sensitive and if TRUE, case is ignored during matching.
`text`	Text to search in files

Details

inside.search Search a string within files of a folder

Value

Return an invisible vector with filenames in which the pattern occurs

Author(s)

Marc Girondot

Examples

```
## Not run:
library(HelpersMG)
# Search for files in path with names based on pattern that have the string search inside.
inside.search(path=".", pattern="*\\.R$", search="embryogrowth")

## End(Not run)
```

`invlogit` *Return the inverse logit*

Description

Return the inverse logit.

Usage

```
invlogit(n)
```

Arguments

`n`	The value to inverse to get the probability

Details

invlogit returns the inverse logit

Value

A value

Author(s)

Marc Girondot

See Also

Other logit: `logit`

Examples

```
n <- logit(0.5)
invlogit(n)
```

LD50 *Estimate the parameters that best describe LD50*

Description

Estimate the parameters that best describe LD50
Logistic and logit models are the same but wit different parametrization:
logistic=1/(1+exp((1/S)*(P-d))) logit=1/(1+exp(P+d*S))

Usage

```
LD50(df = NULL, alive = NULL, dead = NULL, N = NULL,
  doses = NULL, l = 0.05, parameters.initial = NULL,
  fixed.parameters = NULL, SE = NULL, equation = "logistic",
  replicates = 1000, range.CI = 0.95, limit.low.TRD.minimum = 5,
  limit.high.TRD.maximum = 1000, print = TRUE, doses.plot = seq(from
  = 0, to = 1000, by = 0.1))
```

Arguments

df	A dataframe with at least two columns named alive, dead or N and doses columns
alive	A vector with alive individuals at the end of experiment
dead	A vector with dead individuals at the end of experiment
N	A vector with total numbers of tested individuals
doses	The constant incubation doses used to fit sex ratio
l	The limit to define TRD (see Girondot, 1999)
parameters.initial	Initial values for P, S or K search as a vector, ex. c(P=29, S=-0.3)
fixed.parameters	Parameters that will not be changed during fit
SE	Standard errors for parameters
equation	Could be "logistic", "logit", "probit", Hill", "Richards", "Hulin" or "Double-Richards"
replicates	Number of replicates to estimate confidence intervals
range.CI	The range of confidence interval for estimation, default=0.95
limit.low.TRD.minimum	Minimum lower limit for TRD
limit.high.TRD.maximum	Maximum higher limit for TRD
print	Do the results must be printed at screen? TRUE (default) or FALSE
doses.plot	Sequences of doses that will be used for plotting. If NULL, does not estimate them

Details

LD50 estimates the parameters that best describe LD50

Value

A list with the LD50, Transitional Range of Doses and their SE

Author(s)

Marc Girondot <marc.girondot@u-psud.fr>

See Also

Other LD50 functions: logLik.LD50, plot.LD50, predict.LD50

Examples

```
## Not run:
library("HelpersMG")
data <- data.frame(Doses=c(80, 120, 150, 150, 180, 200),
Alive=c(10, 12, 8, 6, 2, 1),
Dead=c(0, 1, 5, 6, 9, 15))
LD50_logistic <- LD50(data, equation="logistic")
predict(LD50_logistic, doses=c(140, 170))
plot(LD50_logistic)
LD50_probit <- LD50(data, equation="probit")
predict(LD50_probit, doses=c(140, 170))
plot(LD50_probit)
LD50_logit <- LD50(data, equation="logit")
predict(LD50_logit, doses=c(140, 170))
plot(LD50_logit)
LD50_hill <- LD50(data, equation="hill")
predict(LD50_hill, doses=c(140, 170))
plot(LD50_hill)
LD50_Richards <- LD50(data, equation="Richards")
predict(LD50_Richards, doses=c(140, 170))
plot(LD50_Richards)
LD50_Hulin <- LD50(data, equation="Hulin")
predict(LD50_Hulin, doses=c(140, 170))
plot(LD50_Hulin)
LD50_DoubleRichards <- LD50(data, equation="Double-Richards")
predict(LD50_DoubleRichards, doses=c(140, 170))
plot(LD50_DoubleRichards)

## End(Not run)
```

list.packages *List the installed packages with their locations*

Description

List the installed packages with their locations and version.

Usage

```
list.packages()
```

Details

list.packages lists the installed packages with their locations

Value

A list with the installed packages and their version.

Author(s)

Marc Girondot

Examples

```
## Not run:
library(HelpersMG)
list.packages()

## End(Not run)
```

local.search *Return path of file searched for in local disk based on its file name*

Description

Return path of file searched for in local disk based on its file name.
It has been tested only with Windows XP and MacOSX. In MacOSX, you must have created the locate database first. Use OnyX utilities for this purpose.

Usage

```
local.search(pattern, directory = "", folder = "$HOME",
  intern = TRUE, ignore.stdout = FALSE, ignore.stderr = TRUE)
```

Arguments

pattern	The name of file to be searched for. Can use wildcards *
directory	The path of directory to be explored in for Windows
folder	The path of folder to be explored in for Unix based systems
intern	A logical (not NA) which indicates whether to capture the output of the command as an R character vector (see system()).
ignore.stdout	a logical (not NA) indicating whether messages written to 'stdout' should be ignored (see system()).
ignore.stderr	a logical (not NA) indicating whether messages written to 'stderr' should be ignored (see system()).

Details

local.search() returns path of file serached in local disk based on its file name

Value

A vector with paths

Author(s)

Marc Girondot

Examples

```
## Not run:
RnwFiles <- local.search("*.Rnw")
nc.files <- local.search("*.nc", folder=paste0("'",getwd(),"'"))

## End(Not run)
```

---

logit *Return the logit*

Description

Return the logit.

Usage

```
logit(p)
```

Arguments

p	The probability

Details

logit returns the logit

Value

A value

Author(s)

Marc Girondot

See Also

Other logit: `invlogit`

Examples

```
n <- logit(0.5)
invlogit(n)
```

`logLik.LD50` *Return Log Likelihood of a fit generated by LD50*

Description

Return Log Likelihood of a fit generated by LD50

Usage

```
## S3 method for class 'LD50'
logLik(object, ...)
```

Arguments

`object`	A result file generated by fitRMU
`...`	Not used

Details

logLik.LD50 Return Log Likelihood of a fit for LD50

Value

The Log Likelihood value for the fitted model with data

Author(s)

Marc Girondot

See Also

Other LD50 functions: LD50, plot.LD50, predict.LD50

Examples

```
## Not run:
data <- data.frame(Doses=c(80, 120, 150, 150, 180, 200),
Alive=c(10, 12, 8, 6, 2, 1),
Dead=c(0, 1, 5, 6, 9, 15))
LD50_logistic <- LD50(data, equation="logistic")
logLik(LD50_logistic)
AIC(LD50_logistic)

## End(Not run)
```

---

merge.mcmcComposite *Merge two mcmcComposite results*

Description

Merge two mcmcComposite results and produced a new one mcmcComposite object.
Note that the initial value for the second run must use the last value of the first one as shown in example.

Usage

```
## S3 method for class 'mcmcComposite'
merge(x, y, ...)
```

Arguments

x	A mcmcComposite obtained as a result of MHalgoGen() function
y	A mcmcComposite obtained as a result of MHalgoGen() function
...	not used

Details

merge.mcmcComposite Merge two mcmcComposite results

Value

A mcmcComposite result

Author(s)

Marc Girondot

See Also

Other mcmcComposite functions: MHalgoGen, as.mcmc.mcmcComposite, as.parameters, plot.mcmcComposite, summary.mcmcComposite

Examples

```
## Not run:
library(HelpersMG)
require(coda)
x <- rnorm(30, 10, 2)
dnormx <- function(data, x) {
 data <- unlist(data)
 return(-sum(dnorm(data, mean=x['mean'], sd=x['sd'], log=TRUE)))
}
parameters_mcmc <- data.frame(Density=c('dnorm', 'dlnorm'),
Prior1=c(10, 0.5), Prior2=c(2, 0.5), SDProp=c(1, 1),
Min=c(-3, 0), Max=c(100, 10), Init=c(10, 2), stringsAsFactors = FALSE,
row.names=c('mean', 'sd'))
mcmc_run <- MHalgoGen(n.iter=1000, parameters=parameters_mcmc, data=x,
likelihood=dnormx, n.chains=1, n.adapt=100, thin=1, trace=1)
plot(mcmc_run, xlim=c(0, 20))
plot(mcmc_run, xlim=c(0, 10), parameters="sd")
mcmcforcoda <- as.mcmc(mcmc_run)
#' heidel.diag(mcmcforcoda)
raftery.diag(mcmcforcoda)
autocorr.diag(mcmcforcoda)
acf(mcmcforcoda[[1]][,"mean"], lag.max=20, bty="n", las=1)
acf(mcmcforcoda[[1]][,"sd"], lag.max=20, bty="n", las=1)
batchSE(mcmcforcoda, batchSize=100)
# The batch standard error procedure is usually thought to
# be not as accurate as the time series methods used in summary
summary(mcmcforcoda)$statistics[,"Time-series SE"]
summary(mcmc_run)
as.parameters(mcmc_run)
lastp <- as.parameters(mcmc_run, index="last")
parameters_mcmc[,"Init"] <- lastp
# The n.adapt set to 1 is used to not record the first set of parameters
# then it is not duplicated (as it is also the last one for
# the object mcmc_run)
mcmc_run2 <- MHalgoGen(n.iter=1000, parameters=parameters_mcmc, data=x,
likelihood=dnormx, n.chains=1, n.adapt=1, thin=1, trace=1)
mcmc_run3 <- merge(mcmc_run, mcmc_run2)
####### no adaptation, n.adapt must be 0
parameters_mcmc[,"Init"] <- c(mean(x), sd(x))
mcmc_run3 <- MHalgoGen(n.iter=1000, parameters=parameters_mcmc, data=x,
likelihood=dnormx, n.chains=1, n.adapt=0, thin=1, trace=1)

## End(Not run)
```

MHalgoGen *Monte-Carlo Markov-chain with Metropolis-Hastings algorithm*

Description

The parameters must be stored in a data.frame with named rows for each parameter with the following columns:

- Density. The density function name, example `dnorm`, `dlnorm`, `dunif`
- Prior1. The first parameter to send to the `Density` function
- Prior2. The second parameter to send to the `Density` function
- SDProp. The standard error from new proposition value of this parameter
- Min. The minimum value for this parameter
- Max. The maximum value for this parameter
- Init. The initial value for this parameter

This script has been deeply modified from a MCMC script provided by Olivier Martin (INRA, Paris-Grignon).
The likelihood function must take a parameter named x.
For adaptive mcmc, see:
Rosenthal, J. S. 2011. Optimal Proposal Distributions and Adaptive MCMC. Pages 93-112 in S. Brooks, A. Gelman, G. Jones, and X.-L. Meng, editors. MCMC Handbook. Chapman and Hall/CRC.

Usage

```
MHalgoGen(likelihood = stop("A likelihood function must be supplied"),
  parameters = stop("Priors  must be supplied"), ..., n.iter = 10000,
  n.chains = 1, n.adapt = 100, thin = 30, trace = FALSE,
  adaptive = FALSE, adaptive.lag = 500, adaptive.fun = function(x) {
     ifelse(x > 0.234, 1.3, 0.7) }, intermediate = NULL,
  filename = "intermediate.Rdata", previous = NULL)
```

Arguments

`likelihood`	The function that returns -ln likelihood using data and parameters
`parameters`	A data.frame with priors; see description and examples
`...`	Parameters to be transmitted to likelihood function
`n.iter`	Number of iterations for each chain
`n.chains`	Number of chains
`n.adapt`	Number of iteration to stabilize likelihood
`thin`	Interval for thinning likelihoods
`trace`	Or FALSE or period to show progress

adaptive	Should an adaptive process for SDProp be used
adaptive.lag	Lag to analyze the SDProp value in an adaptive context
adaptive.fun	Function used to change the SDProp
intermediate	Or NULL of period to save intermediate result
filename	Name of file in which intermediate results are saved
previous	The content of the file in which intermediate results are saved

Details

MHalgoGen is a function to use mcmc with Metropolis-Hastings algorithm

Value

A mcmcComposite object with all characteristics of the model and mcmc run

Author(s)

Marc Girondot

See Also

Other mcmcComposite functions: as.mcmc.mcmcComposite, as.parameters, merge.mcmcComposite, plot.mcmcComposite, summary.mcmcComposite

Examples

```
## Not run:
library(HelpersMG)
require(coda)
val <- rnorm(30, 10, 2)
dnormx <- function(data, x) {
 data <- unlist(data)
 return(-sum(dnorm(data, mean=x['mean'], sd=x['sd'], log=TRUE)))
}
parameters_mcmc <- data.frame(Density=c('dnorm', 'dlnorm'),
Prior1=c(10, 0.5), Prior2=c(2, 0.5), SDProp=c(0.35, 0.2),
Min=c(-3, 0), Max=c(100, 10), Init=c(10, 2), stringsAsFactors = FALSE,
row.names=c('mean', 'sd'))
mcmc_run <- MHalgoGen(n.iter=50000, parameters=parameters_mcmc, data=val,
likelihood=dnormx, n.chains=1, n.adapt=100, thin=1, trace=1)
plot(mcmc_run, xlim=c(0, 20))
plot(mcmc_run, xlim=c(0, 10), parameters="sd")
library(graphics)
library(fields)
# show a scatter plot of the result
x <- mcmc_run$resultMCMC[[1]][, 1]
y <- mcmc_run$resultMCMC[[1]][, 2]
marpre <- par(mar=c(4, 4, 2, 6)+0.4)
smoothScatter(x, y)
# show a scale
```

```
n <- matrix(0, ncol=128, nrow=128)
xrange <- range(x)
yrange <- range(y)
for (i in 1:length(x)) {
  posx <- 1+floor(127*(x[i]-xrange[1])/(xrange[2]-xrange[1]))
  posy <- 1+floor(127*(y[i]-yrange[1])/(yrange[2]-yrange[1]))
  n[posx, posy] <- n[posx, posy]+1
}
image.plot(legend.only=TRUE, zlim= c(0, max(n)), nlevel=128,
 col=colorRampPalette(c("white", blues9))(128))
# Compare with a heatmap
x <- seq(from=8, to=12, by=0.2)
y <- seq(from=1, to=4, by=0.2)
df <- expand.grid(mean=x, sd=y)
df <- cbind(df, L=rep(0, length(nrow(df))))
for (i in 1:nrow(df)) df[i, "L"] <- -sum(dnorm(val, df[i, 1], df[i, 2], log = TRUE))
hm <- matrix(df[, "L"], nrow=length(x))
par(mar = marpre)
image.plot(x=x, y=y, z=hm, las=1)
# Diagnostic function from coda library
mcmcforcoda <- as.mcmc(mcmc_run)
#' heidel.diag(mcmcforcoda)
raftery.diag(mcmcforcoda)
autocorr.diag(mcmcforcoda)
acf(mcmcforcoda[[1]][,"mean"], lag.max=20, bty="n", las=1)
acf(mcmcforcoda[[1]][,"sd"], lag.max=20, bty="n", las=1)
batchSE(mcmcforcoda, batchSize=100)
# The batch standard error procedure is usually thought to
# be not as accurate as the time series methods used in summary
summary(mcmcforcoda)$statistics[,"Time-series SE"]
summary(mcmc_run)
as.parameters(mcmc_run)
lastp <- as.parameters(mcmc_run, index="last")
parameters_mcmc[,"Init"] <- lastp
# The n.adapt set to 1 is used to not record the first set of parameters
# then it is not duplicated (as it is also the last one for
# the object mcmc_run)
mcmc_run2 <- MHalgoGen(n.iter=1000, parameters=parameters_mcmc, x=x,
likelihood=dnormx, n.chains=1, n.adapt=1, thin=1, trace=1)
mcmc_run3 <- merge(mcmc_run, mcmc_run2)
####### no adaptation, n.adapt must be 0
parameters_mcmc[,"Init"] <- c(mean(x), sd(x))
mcmc_run3 <- MHalgoGen(n.iter=1000, parameters=parameters_mcmc, x=x,
likelihood=dnormx, n.chains=1, n.adapt=0, thin=1, trace=1)
# Here is how to use adaptive mcmc
mcmc_run <- MHalgoGen(n.iter=50000, parameters=parameters_mcmc, data=val, adaptive = FALSE,
likelihood=dnormx, n.chains=1, n.adapt=100, thin=1, trace=1)
1-rejectionRate(as.mcmc(mcmc_run))
mcmc_run <- MHalgoGen(n.iter=50000, parameters=parameters_mcmc, data=val, adaptive = TRUE,
likelihood=dnormx, n.chains=1, n.adapt=100, thin=1, trace=1)
1-rejectionRate(as.mcmc(mcmc_run))
# To see the dynamics :
var <- "mean"
```

```
par(mar=c(4, 4, 1, 1)+0.4)
plot(1:nrow(mcmc_run$resultMCMC[[1]]), mcmc_run$resultMCMC[[1]][, var], type="l",
       xlab="Iterations", ylab=var, bty="n", las=1)

## End(Not run)
```

`minmax.periodic` *Search for minimum and maximum indices in periodic timeseries*

Description

Search for minimum and maximum for periodic timeseries when only intermediate values are known.
For each couple of value with an increasing or decreasing segment of the sinusoid function, it is possible to estimate a minimum and maximum values using analytical algebra.
Then the average and standard deviations of all minima and maxima are evaluated.
It should be noted that any extremum can be estimated at least twice, one by increasing segment and one by decreasing segment. Both are used here to produce SD.
`time.minmax.daily` should be used when the time at which maximum and minimum indices are regular and `time.minmax` permits to define this time day by day.

Usage

```
minmax.periodic(time.minmax.daily = NULL, time.minmax = NULL,
  progressbar = FALSE,
  observed = stop("data.frame with observed indices"), period = 24,
  colname.time = "time", colname.index = "index", colname.SD = "SD",
  plot = FALSE)
```

Arguments

`time.minmax.daily`	A named vector with Min and Max being the time in the day with minimum and maximum indices (temperature or level)
`time.minmax`	A named vector daily with time in the day at which minimum and maximum indices are observed
`progressbar`	Tell if a progression bar must be shown
`observed`	A dataframe with at least two columns: time and temperatures. A third column SD can indicate the know error in index
`period`	The unit of day period (24 for hours, 24*60 for minutes)
`colname.time`	The name of the column for time in observed
`colname.index`	The name of the column for indices in observed
`colname.SD`	The name of the column for SD in observed
`plot`	If TRUE, show a plot with the different estimates

Details

minmax.periodic search for minimum and maximum indices (temperatures or levels) in periodic timeseries

Value

A data.frame with a column time, a column index and a column SD

Author(s)

Marc Girondot

See Also

Other Periodic patterns of indices: index.periodic, moon.info, sun.info, tide.info

Examples

```
## Not run:
library("HelpersMG")
# Generate a timeserie of time
time.obs <- NULL
for (i in 0:9) time.obs <- c(time.obs, c(0, 6, 12, 18)+i*24)
# For these time, generate a timeseries of temperatures
temp.obs <- rep(NA, length(time.obs))
temp.obs[3+(0:9)*4] <- rnorm(10, 25, 3)
temp.obs[1+(0:9)*4] <- rnorm(10, 10, 3)
for (i in 1:(length(time.obs)-1))
  if (is.na(temp.obs[i]))
  temp.obs[i] <- mean(c(temp.obs[i-1], temp.obs[i+1]))
  if (is.na(temp.obs[length(time.obs)]))
  temp.obs[length(time.obs)] <- temp.obs[length(time.obs)-1]/2
observed <- data.frame(time=time.obs, temperature=temp.obs)
# Search for the minimum and maximum values
r <- minmax.periodic(time.minmax.daily=c(Min=2, Max=15),
observed=observed, period=24, colname.index="temperature")

# Estimate all the temperatures for these values
t <- index.periodic(minmax=r)

plot_errbar(x=t[,"time"], y=t[,"index"],
errbar.y=ifelse(is.na(t[,"sd"]), 0, 2*t[,"sd"]),
type="l", las=1, bty="n", errbar.y.polygon = TRUE,
xlab="hours", ylab="Temperatures", ylim=c(0, 35),
errbar.y.polygon.list = list(col="grey"))

plot_add(x=t[,"time"], y=t[,"index"], type="l")

plot_add(observed$time, observed$temperature, pch=19, cex=0.5)

## End(Not run)
```

`modeled.hist` *Return the theoretical value for the histogram bar*

Description

Return the theoretical value for the histogram bar based on a model of distribution.

Usage

```
modeled.hist(breaks, FUN, ..., sum = 1)
```

Arguments

`breaks`	Vector with the breaks; it can be obtained directly from hist()
`FUN`	Function to be used to integrate the density, ex. pnorm
`...`	Parameters to be used by FUN
`sum`	Total numbers in the histogram; 1 for emperical frequencies

Details

modeled.hist returns the theoretical value for the histogram bar based on a model of distribution.

Value

A list with x (the center of the bar) and y components

Author(s)

Marc Girondot

Examples

```
## Not run:
n <- rnorm(100, mean=10, sd=2)
breaks <- 0:20
hist(n, breaks=breaks)

s <- modeled.hist(breaks=breaks, FUN=pnorm, mean=10, sd=2, sum=100)

points(s$x, s$y, pch=19)
lines(s$x, s$y)

n <- rlnorm(100, meanlog=2, sdlog=0.4)
b <- hist(n, ylim=c(0, 70))

s <- modeled.hist(breaks=b$breaks, FUN=plnorm, meanlog=2, sdlog=0.4, sum=100)

points(s$x, s$y, pch=19)
```

```
lines(s$x, s$y)

## End(Not run)
```

`modifyVector` *Modifies Elements of a Vector*

Description

Modifies a vector by changing a subset of elements to match a second vector.

Usage

```
modifyVector(x, val)
```

Arguments

`x`	A named vector.
`val`	A named vector with components to replace corresponding components in x.

Details

modifyVector modifies elements of a vector

Value

A modified version of x, with the elements of val replacing the elements of x

Author(s)

Marc Girondot

Examples

```
library("HelpersMG")
e <- c(M=10, L=20, J=30)
modifyVector(e, c(U=10, M=30))
```

`moon.info` *Moon phase based on a date*

Description

The script gives an index (base 100) that represents moon phase.
If the return value (from 0 to 100) is between:
0 and 1.6931595 or 98.3068405 and 100, it is full moon,
23.3068405 and 26.6931595, last quarter,
48.3068405 and 51.6931595, new moon,
73.3068405 and 76.6931595, first quarter
When phase is set to TRUE, a character representing the moon phase is returned.

Usage

```
moon.info(date = Sys.Date(), phase = FALSE)
```

Arguments

`date`	A date in class Date. By default, it will use today date
`phase`	If TRUE, a vector of characters with NM, FQ, FL LQ will be returned

Details

moon.info calculates the moon phase based on a date.

Value

Return a value describing the moon phase:
0 and 100 are full moon, 50 is new moon, 25 last quarter and 75 first quater

Author(s)

Marc Girondot <marc.girondot@u-psud.fr>

See Also

Other Periodic patterns of indices: `index.periodic`, `minmax.periodic`, `sun.info`, `tide.info`

Examples

```
## Not run:
library("HelpersMG")
moon.info(as.Date("2001-12-31"))
moon.info(as.Date("14/04/2010", "%d/%m/%Y"))
moon.info(as.Date("22/06/07", "%d/%m/%y"))
moon.info(seq(from=as.Date("2012-03-01"),
to=as.Date("2012-04-15"), by="days"))
```

```
moon.info(seq(from=as.Date("2012-03-01"),
to=as.Date("2012-04-15"), by="days"), phase=TRUE)

## End(Not run)
```

`newcompassRose` *Display a compass rose*

Description

Displays a basic compass rose, usually to orient a map.
newcompassRose displays a conventional compass rose at the position requested.
The size of the compass rose is determined by the character expansion, as the central "rose" is calculated relative to the character size.
Rotation is in degrees counterclockwise.

Usage

```
newcompassRose(x, y, rot = 0, cex = 1, col = "black",
  col.arrows.light = "white", col.arrows.dark = "black")
```

Arguments

`x`	The position of the center of the compass rose in user units.
`y`	The position of the center of the compass rose in user units.
`rot`	Rotation for the compass rose in degrees. See Details.
`cex`	The character expansion to use in the display.
`col`	The color of text
`col.arrows.light`	The color of lighter lines
`col.arrows.dark`	The color of darker lines

Details

newcompassRose Display a compass rose

Value

none

Author(s)

modified from Jim Lemon; See compassRose sp

Examples

```
## Not run:
library(HelpersMG)
require("maps")
map("world", "China")
newcompassRose(x=110, y=35, col.arrows.light="grey")

## End(Not run)
```

---

newdbeta *Density for the Beta distributions.*

Description

Density for the Beta distribution with parameters mu and v or shape1 and shape2 (and optional non-centrality parameter ncp).

Usage

```
newdbeta(x, mu = NULL, v = NULL, shape1, shape2, ncp = 0,
  log = FALSE)
```

Arguments

x	vector of quantiles.
mu	mean of the Beta distribution.
v	variance of the Beta distribution.
shape1	non-negative parameters of the Beta distribution.
shape2	non-negative parameters of the Beta distribution.
ncp	non-centrality parameter.
log	logical; if TRUE, probabilities p are given as log(p).

Details

newdbeta returns the density for the Beta distributions

The Beta distribution with parameters shape1 = a and shape2 = b has density
gamma(a+b)/(gamma(a)gamma(b))x^(a-1)(1-x)^(b-1)
for a > 0, b > 0 and 0 <= x <= 1 where the boundary values at x=0 or x=1 are defined as by continuity (as limits).
The mean is a/(a+b) and the variance is ab/((a+b)^2 (a+b+1)). These moments and all distributional properties can be defined as limits.

Value

newdbeta gives the density for the Beta distributions

Author(s)

Marc Girondot

Examples

```
pi <- rbeta(100, shape1=0.48, shape2=0.12)
hist(pi, freq=FALSE, breaks=seq(from=0, to=1, by=0.1), ylim=c(0, 8), las=1)
library("HelpersMG")
mx <- ScalePreviousPlot()$ylim["end"]/
      max(newdbeta(seq(from=0.01, to=0.99, by=0.01), mu = 0.8, v=0.1))
curve(newdbeta(x, mu = 0.8, v=0.1)*mx, add=TRUE, col="red")
```

---

newmap.scale *Add Scale to Existing Unprojected Map*

Description

Adds a scale to an existing map, both as a ratio and a distance gauge. If x or y are not specified, this will be taken to be near the lower left corner of the map.

Usage

```
newmap.scale(x, y, relwidth = 0.15, metric = TRUE, ratio = TRUE,
  col.line = "black", ...)
```

Arguments

x	Location of left end of distance gauge.
y	Location of left end of distance gauge.
relwidth	Proportion of width of display to be used for the scale. The default is 0.15.
metric	If TRUE, the distance gauge will be in km, otherwise miles.
ratio	If FALSE, the scale ratio of the map is not displayed.
col.line	The color of lines for the gauge.
...	Further plotting parameters may be specified as for the command text().

Details

newmap.scale Add Scale to Existing Unprojected Map

Value

The exact calculated scale is returned.

Author(s)

See map.scale maps

Examples

```
## Not run:
library("maps")
library("HelpersMG")
map("world", "China")
newmap.scale(col.line = "red", col="blue")

## End(Not run)
```

`plot.IconoCorel` *Clean the dataframe before to be used with IC_threshold_matrix*

Description

This function plots the data as a network. It returns an invisible object that can be used with visIgraph from package visNetwork. `https://fr.wikipedia.org/wiki/Iconographie_des_corrélations`

Usage

```
## S3 method for class 'IconoCorel'
plot(x, ..., show.legend.direction = "bottomright",
  show.legend.strength = "topleft", title = "Correlation iconography",
  vertex.label.color = "black", vertex.label = NULL,
  vertex.color = "white", plot = TRUE)
```

Arguments

`x`	The correlation matrix to show
`...`	other options of plot.igraph()
`show.legend.direction`	the position of the legend of direction; FALSE to not show it
`show.legend.strength`	the position of the legend with intensity of correlation; FALSE to not show it
`title`	the title of the plot
`vertex.label.color`	a vector with the colors of labels
`vertex.label`	a vector with the labels
`vertex.color`	a vector of colors
`plot`	if TRUE, the plot is shown

Details

plot.IconoCorel checks and corrects the dataframe to be used with IC_threshold_matrix

Value

A igraph object

Author(s)

Marc Girondot

References

Lesty, M., 1999. Une nouvelle approche dans le choix des régresseurs de la régression multiple en présence d'interactions et de colinéarités. Revue de Modulad 22, 41-77.

See Also

Other Iconography of correlations: IC_clean_data, IC_correlation_simplify, IC_threshold_matrix

Examples

```
## Not run:
library("HelpersMG")
es <- matrix(c("e1", "52", "12", "12", "5",
"e2", "59", "12.5", "9", "5",
"e3", "55", "13", "15", "9",
"e4", "58", "14.5", "5", "5",
"e5", "66", "15.5", "11", "13.5",
"e6", "62", "16", "15", "18",
"e7", "63", "17", "12", "18",
"e8", "69", "18", "9", "18"), ncol=5, byrow = TRUE)
colnames(es) <- c("Élève", "Poids", "Âge", "Assiduité", "Note")
es <- as.data.frame(es, stringsasFactor=FALSE)
es[, 2] <- as.numeric(as.character(es[, 2]))
es[, 3] <- as.numeric(as.character(es[, 3]))
es[, 4] <- as.numeric(as.character(es[, 4]))
es[, 5] <- as.numeric(as.character(es[, 5]))

es

df <- IC_clean_data(es, debug = TRUE)
cor_matrix <- IC_threshold_matrix(data=df, threshold = NULL, progress=FALSE)
cor_threshold <- IC_threshold_matrix(data=df, threshold = 0.3)
par(mar=c(1,1,1,1))
set.seed(4)
library("igraph")
library("visNetwork")
kk <- plot(cor_threshold, vertex.color="red")
# it can be shown also with the visNetwork package
visIgraph(kk)
cor_threshold_Note <- IC_correlation_simplify(matrix=cor_threshold, variable="Note")
plot(cor_threshold_Note)

## End(Not run)
```

plot.LD50 *Plot results of LD50() that best describe LD50*

Description

Plot the estimates that best describe lethality of exposures.

Usage

```
## S3 method for class 'LD50'
plot(x, ..., las.x = 1, las.y = 1, lab.PT = "LD50",
  lab.TRD = paste0("Transitional range of doses l=", l * 100, "%"),
  col.TRD = "gray", col.TRD.CI = rgb(0.8, 0.8, 0.8, 0.5),
  col.PT.CI = rgb(0.8, 0.8, 0.8, 0.5), show.CI = TRUE)
```

Arguments

x	A result file generated by IC50()
...	Parameters for plot()
las.x	las parameter for x axis
las.y	las parameter for y axis
lab.PT	Label to describe pivotal temperature
lab.TRD	Label to describe transitional range of temperature
col.TRD	The color of TRD
col.TRD.CI	The color of CI of TRD based on range.CI
col.PT.CI	The color of CI of PT based on range.CI
show.CI	Do the CI for the curve should be shown

Details

plot.LD50 plot result of IC50() that best describe IC50

Value

Nothing

Author(s)

Marc Girondot

References

Girondot, M. 1999. Statistical description of temperature-dependent sex determination using maximum likelihood. Evolutionary Ecology Research, 1, 479-486.

Godfrey, M.H., Delmas, V., Girondot, M., 2003. Assessment of patterns of temperature-dependent sex determination using maximum likelihood model selection. Ecoscience 10, 265-272.

Hulin, V., Delmas, V., Girondot, M., Godfrey, M.H., Guillon, J.-M., 2009. Temperature-dependent sex determination and global change: are some species at greater risk? Oecologia 160, 493-506.

Girondot M., Submited. On the concept of embryological thermosensitive period for sex determination in reptiles.

See Also

Other LD50 functions: `LD50`, `logLik.LD50`, `predict.LD50`

Examples

```
## Not run:
#' data <- data.frame(Doses=c(80, 120, 150, 150, 180, 200),
Alive=c(10, 12, 8, 6, 2, 1),
Dead=c(0, 1, 5, 6, 9, 15))
LD50_logistic <- LD50(data, equation="logistic")
predict(LD50_logistic, doses=c(140, 170))
plot(LD50_logistic)

## End(Not run)
```

`plot.mcmcComposite` *Plot the result of a mcmcComposite object*

Description

Plot the results within a mcmcCompositve object.
The parameters to use can be called by:
`parameters="all"`
`parameters=1:4`
`parameters=c("PAR1", "PAR2", "PAR5")`
`parameters=c(TRUE, TRUE, FALSE, TRUE)`
If scale.prior is TRUE, another scale is shown at right.
legend can take these values: FALSE, TRUE, topleft, topright, bottomleft, bottomright.

Usage

```
## S3 method for class 'mcmcComposite'
plot(x, ..., chain = 1, parameters = 1,
  scale.prior = TRUE, legend = "topright",
  ylab = "Posterior density", las = 1, col.prior = "red",
  lty.prior = 1, lwd.prior = 1, col.posterior = "white",
  lty.posterior = 1, lwd.posterior = 1, ylab.prior = "Prior density")
```

Arguments

x	A mcmcComposite object
...	Graphical parameters to be send to hist()
chain	The chain to use
parameters	Name of parameters or their number (see description)
scale.prior	If TRUE, the prior is scaled at the same size as posterior
legend	If FALSE, the legend is not shown
ylab	y-label for posterior
las	las parameter (orientation of y-axis graduation)
col.prior	Color for prior curve
lty.prior	Type of line for prior curve
lwd.prior	Width of line for prior curve
col.posterior	Color for posterior histogram
lty.posterior	Type of line for posterior histogram
lwd.posterior	Width of line for posterior histogram
ylab.prior	y-label for prior

Details

plot.mcmcComposite plots the result of a MCMC search

Value

None

Author(s)

Marc Girondot

See Also

Other mcmcComposite functions: MHalgoGen, as.mcmc.mcmcComposite, as.parameters, merge.mcmcComposite, summary.mcmcComposite

Examples

```
## Not run:
library(HelpersMG)
require(coda)
x <- rnorm(30, 10, 2)
dnormx <- function(data, x) {
 data <- unlist(data)
 return(-sum(dnorm(data, mean=x['mean'], sd=x['sd'], log=TRUE)))
}
parameters_mcmc <- data.frame(Density=c('dnorm', 'dlnorm'),
Prior1=c(10, 0.5), Prior2=c(2, 0.5), SDProp=c(1, 1),
```

```
Min=c(-3, 0), Max=c(100, 10), Init=c(10, 2), stringsAsFactors = FALSE,
row.names=c('mean', 'sd'))
mcmc_run <- MHalgoGen(n.iter=50000, parameters=parameters_mcmc, data=x,
adaptive = TRUE,
likelihood=dnormx, n.chains=1, n.adapt=100, thin=1, trace=1)
plot(mcmc_run, xlim=c(0, 20))
plot(mcmc_run, xlim=c(0, 10), parameters="sd")
mcmcforcoda <- as.mcmc(mcmc_run)
#' heidel.diag(mcmcforcoda)
raftery.diag(mcmcforcoda)
autocorr.diag(mcmcforcoda)
acf(mcmcforcoda[[1]][,"mean"], lag.max=20, bty="n", las=1)
acf(mcmcforcoda[[1]][,"sd"], lag.max=20, bty="n", las=1)
batchSE(mcmcforcoda, batchSize=100)
# The batch standard error procedure is usually thought to
# be not as accurate as the time series methods used in summary
summary(mcmcforcoda)$statistics[,"Time-series SE"]
summary(mcmc_run)
as.parameters(mcmc_run)
lastp <- as.parameters(mcmc_run, index="last")
parameters_mcmc[,"Init"] <- lastp
# The n.adapt set to 1 is used to not record the first set of parameters
# then it is not duplicated (as it is also the last one for
# the object mcmc_run)
mcmc_run2 <- MHalgoGen(n.iter=50000, parameters=parameters_mcmc, data=x,
adaptive = TRUE,
likelihood=dnormx, n.chains=1, n.adapt=1, thin=1, trace=1)
mcmc_run3 <- merge(mcmc_run, mcmc_run2)
####### no adaptation, n.adapt must be 0
parameters_mcmc[,"Init"] <- c(mean(x), sd(x))
mcmc_run3 <- MHalgoGen(n.iter=1000, parameters=parameters_mcmc, data=x,
adaptive = TRUE,
likelihood=dnormx, n.chains=1, n.adapt=0, thin=1, trace=1)

## End(Not run)
```

`plot_add` *Add a plot to a previous one*

Description

To plot data, just add use it as a normal plot. It will plot the new data without axes, or labels for axes.
This function is complementary to matlines() and matpoints() from package graphics.

Usage

```
plot_add(...)
```

Arguments

...	Parameters for plot()

Details

plot_add adds a plot to a previous one

Value

Nothing

Author(s)

Marc Girondot

See Also

Other plot and barplot functions: ScalePreviousPlot, barplot_errbar, plot_errbar

Examples

```
## Not run:
plot(x=1:100, y=sin(1:100), type="l", bty="n", xlim=c(1,200), xlab="x", ylab="y")
plot_add(x=1:200, y=cos(1:200), type="l", bty="n", col="red")

## End(Not run)
```

---

plot_errbar *Plot a xy graph with error bar on x and/or y*

Description

To plot data, just use it as a normal plot but add the errbar.x and errbar.y values or errbar.x.minus, errbar.x.plus if bars for x axis are asymetric and errbar.y.minus, errbar.y.plus if bars for y axis are asymetric. Use x.plus, x.minus, y.plus and y.minus to set absolut limits for error bars. Note that x.plus and x.minus have priority over errbar.x, errbar.x.minus and errbar.x.plus and that y.plus and y.minus have priority over errbar.y, errbar.y.minus and errbar.y.plus.
The parameter errbar.y.polygon=TRUE permits to define error as an envolop for y axis.

Usage

```
plot_errbar(..., errbar.x = NULL, errbar.y = NULL,
  errbar.x.plus = NULL, errbar.x.minus = NULL, errbar.y.plus = NULL,
  errbar.y.minus = NULL, x.plus = NULL, x.minus = NULL,
  y.plus = NULL, y.minus = NULL, errbar.tick = 1/50,
  errbar.lwd = par("lwd"), errbar.lty = par("lty"),
  errbar.col = par("fg"), errbar.y.polygon = FALSE,
  errbar.y.polygon.list = list(NULL), add = FALSE)
```

Arguments

`...`	Parameters for plot() such as main= or ylim=
`errbar.x`	The length of error bars for x. Recycled if necessary.
`errbar.y`	The length of error bars for y. Recycled if necessary.
`errbar.x.plus`	The length of positive error bars for x. Recycled if necessary.
`errbar.x.minus`	The length of negative error bars for x. Recycled if necessary.
`errbar.y.plus`	The length of positive error bars for y. Recycled if necessary.
`errbar.y.minus`	The length of negative error bars for y. Recycled if necessary.
`x.plus`	The absolut position of the positive error bar for x. Recycled if necessary.
`x.minus`	The absolut position of the negative error bar for x. Recycled if necessary.
`y.plus`	The absolut position of the positive error bar for y. Recycled if necessary.
`y.minus`	The absolut position of the nagative error bar for y. Recycled if necessary.
`errbar.tick`	Size of small ticks at the end of error bars defined as a proportion of total width or height graph size.
`errbar.lwd`	Error bar line width, see par("lwd")
`errbar.lty`	Error bar line type, see par("lwd")
`errbar.col`	Error bar line color, see par("col")
`errbar.y.polygon`	If true, the errors are shown as a filed polygon.
`errbar.y.polygon.list`	List of parameters to be used for polygon.
`add`	If true, add the graph to the previous one.

Details

plot_errbar plot a xy graph with error bar on x and/or y

Value

Nothing

Author(s)

Marc Girondot

See Also

`barplot_errorbar`

Other plot and barplot functions: `ScalePreviousPlot`, `barplot_errbar`, `plot_add`

Examples

```
## Not run:
plot_errbar(1:100, rnorm(100, 1, 2),
xlab="axe x", ylab="axe y", bty="n", xlim=c(1,100),
errbar.x=2, errbar.y=rnorm(100, 1, 0.1))
x <- 1:100
plot_errbar(x=1:100, rnorm(100, 1, 2),
                xlab="axe x", ylab="axe y", bty="n", xlim=c(1,100),
             x.minus=x-2, x.plus=x+2)
x <- 1:100
plot_errbar(x=1:100, rnorm(100, 1, 2),
                xlab="axe x", ylab="axe y", bty="n",
                pch=21, bg="white",
             x.minus=x-10, x.plus=x+10)
x <- (1:200)/10
y <- sin(x)
plot_errbar(x=x, y=y, xlab="axe x", ylab="axe y", bty="n", xlim=c(1,20),
     y.minus=y-1, y.plus=y+1, ylim=c(-3, 3), type="l",
errbar.y.polygon=TRUE,
errbar.y.polygon.list=list(border=NA, col=rgb(0, 0, 0, 0.5)))

## End(Not run)
```

`predict.LD50` *Estimate survival according to doses*

Description

Estimate survival according to doses.
The returned data.frame has the following components:
doses, SE, survival, CI.minus.sexratio, CI.plus.sexratio, range.CI

Usage

```
## S3 method for class 'LD50'
predict(object, doses = NULL, SE = NULL,
  range.CI = 0.95, replicates = 1000, progressbar = FALSE, ...)
```

Arguments

`object`	A result file generated by LD50
`doses`	A vector of temperatures
`SE`	The standard error for doses, optional
`range.CI`	The range of confidence interval for estimation, default=0.95
`replicates`	Number of replicates to estimate CI
`progressbar`	Logical. Does a progression bar must be shown
`...`	Not used

Details

predict.LD50 Estimate survival according to doses

Value

A data.frame with informations about survival

Author(s)

Marc Girondot

See Also

Other LD50 functions: LD50, logLik.LD50, plot.LD50

Examples

```
## Not run:
#' data <- data.frame(Doses=c(80, 120, 150, 150, 180, 200),
Alive=c(10, 12, 8, 6, 2, 1),
Dead=c(0, 1, 5, 6, 9, 15))
LD50_logistic <- LD50(data, equation="logistic")
predict(LD50_logistic, doses=c(140, 170))
plot(LD50_logistic

## End(Not run)
```

---

pSnbinom *Distribution function for the sum of random variable with negative binomial distributions.*

Description

Distribution function for the sum of random variable with negative binomial distributions.

Usage

```
pSnbinom(q = stop("At least one quantile must be provided"),
  size = NULL, prob = NULL, mu = NULL, lower.tail = TRUE,
  log.p = FALSE, infinite = 100)
```

Arguments

q	vector of quantiles.
size	target for number of successful trials, or dispersion parameter (the shape parameter of the gamma mixing distribution). Must be strictly positive, need not be integer.
prob	probability of success in each trial. 0 < prob <= 1.

mu	alternative parametrization via mean.
lower.tail	logical; if TRUE (default), probabilities are P[X <= x], otherwise, P[X > x].
log.p	logical; if TRUE, probabilities p are given as log(p).
infinite	Number of maximal iterations; check different values to determine the error in estimation.

Details

pSnbinom returns the distribution function for the sum of random variable with negative binomial distributions

Value

pSnbinom returns distribution function

Author(s)

Marc Girondot

See Also

Other Distribution of sum of random variable with negative binomial distributions: dSnbinom, qSnbinom, rSnbinom

Examples

```
## Not run:
alpha <- c(2.1, 2.05, 2)
mu <- c(10, 30, 20)
p <- pSnbinom(q=10, size=alpha, mu=mu, lower.tail = TRUE)

## End(Not run)
```

---

qSnbinom *Quantile function for the sum of random variable with negative binomial distributions.*

Description

Quantile function for the sum of random variable with negative binomial distributions.

Usage

```
qSnbinom(p = stop("At least one probability must be provided"),
  size = stop("size parameter is mandatory"), prob = NULL, mu = NULL,
  lower.tail = TRUE, log.p = FALSE, infinite = 100)
```

Arguments

p	vector of probabilities.
size	target for number of successful trials, or dispersion parameter (the shape parameter of the gamma mixing distribution). Must be strictly positive, need not be integer.
prob	probability of success in each trial. 0 < prob <= 1.
mu	alternative parametrization via mean.
lower.tail	logical; if TRUE (default), probabilities are P[X <= x], otherwise, P[X > x].
log.p	logical; if TRUE, probabilities p are given as log(p).
infinite	Number of maximal iterations; check different values to determine the error in estimation.

Details

qSnbinom returns the quantile function for the sum of random variable with negative binomial distributions

Value

qSnbinom returns quantile function

Author(s)

Marc Girondot

See Also

Other Distribution of sum of random variable with negative binomial distributions: dSnbinom, pSnbinom, rSnbinom

Examples

```
## Not run:
alpha <- c(2.1, 2.05, 2)
mu <- c(10, 30, 20)
q <- qSnbinom(p=0.1, size=alpha, mu=mu, lower.tail = TRUE)

## End(Not run)
```

qvlmer *Quasi Variances for lmer Model Coefficients*

Description

Computes a set of quasi variances (and corresponding quasi standard errors) for estimated model coefficients relating to the levels of a categorical (i.e., factor) explanatory variable. For details of the method see Firth (2000), Firth (2003) or Firth and de Menezes (2004). Quasi variances generalize and improve the accuracy of "floating absolute risk" (Easton et al., 1991). This device for economical model summary was first suggested by Ridout (1989).
Modified from qvcalc.lm() of packages qvcalc by David Firth, d.firth@warwick.ac.uk

Usage

```
qvlmer(object, factorname = NULL, coef.indices = NULL,
  dispersion = NULL, ...)
```

Arguments

object	A object obtained using lmer from package lme4
factorname	Either NULL, or a character vector of length 1
coef.indices	Either NULL, or a numeric vector of length at least 3
dispersion	An optional scalar multiplier for the covariance matrix, to cope with overdispersion for example
...	Other arguments to pass to qvcalc.default

Details

qvlmer is Quasi Variances for lmer Model Coefficients

Value

A list of class qv.

Author(s)

marc.girondot@u-psud.fr

References

Easton, D. F, Peto, J. and Babiker, A. G. A. G. (1991) Floating absolute risk: an alternative to relative risk in survival and case-control analysis avoiding an arbitrary reference group. Statistics in Medicine 10, 1025–1035.

Firth, D. (2000) Quasi-variances in Xlisp-Stat and on the web. Journal of Statistical Software 5.4, 1–13. At http://www.jstatsoft.org

Firth, D. (2003) Overcoming the reference category problem in the presentation of statistical models. Sociological Methodology 33, 1–18.

Firth, D. and de Mezezes, R. X. (2004) Quasi-variances. Biometrika 91, 65–80.

McCullagh, P. and Nelder, J. A. (1989) Generalized Linear Models. London: Chapman and Hall.

Menezes, R. X. de (1999) More useful standard errors for group and factor effects in generalized linear models. D.Phil. Thesis, Department of Statistics, University of Oxford.

Ridout, M.S. (1989). Summarizing the results of fitting generalized linear models to data from designed experiments. In: Statistical Modelling: Proceedings of GLIM89 and the 4th International Workshop on Statistical Modelling held in Trento, Italy, July 17–21, 1989 (A. Decarli et al., eds.), pp 262–269. New York: Springer.

Examples

```
## Not run:
x <- rnorm(100)
y <- rnorm(100)
G <- as.factor(sample(c("A", "B", "C", "D"), 100, replace = TRUE))
R <- as.factor(rep(1:25, 4))
library(lme4)
m <- lmer(y ~ x + G + (1 | R))
qvlmer(m, factorname="G")

## End(Not run)
```

---

r2norm — *Random generation for Gaussian distributions different at left and right*

Description

Random generation for Gaussian distributions different at left and right

Usage

```
r2norm(n, mean = 0, sd_low = 1, sd_high = 1)
```

Arguments

n	number of observations.
mean	vector of means
sd_low	vector of standard deviations below the mean.
sd_high	vector of standard deviations above the mean.

Details

r2norm returns random numbers for Gaussian distributions different at left and right

Value

r2norm returns random numbers

Author(s)

Marc Girondot

Examples

```
## Not run:
n <- r2norm(1000, mean=25, sd_low=2, sd_high=10)

hist(n)

## End(Not run)
```

`read_folder` *Read files present in a folder and creates a list with the content of these files*

Description

To create a list, the syntax is:
datalist <- read_folder(folder=".", read=read.delim, header=FALSE)
It returns an error if the folder does not exist.
The names of the elements of the list are the filenames.
The parameter file can be used to predefine a list of file. If file is NULL, all the files of the folder/directory are used.

Usage

```
read_folder(folder = try(file.choose(), silent = TRUE), file = NULL,
  wildcard = "*.*", read = read.delim, ...)
```

Arguments

`folder`	Where to search for files; can be or a file path or a folder path
`file`	list of files
`wildcard`	Define which files are to be read (examples: "*.*", "*.xls", "essai*.txt"). It can be also a vector with all filenames.
`read`	Function used to read file. Ex: read.delim or read.xls from gdata package
`...`	Parameters send to the read function

Details

read_folder reads all files present in a folder

Value

Return a list with the data in the files of the folder (directory for windows users)

Author(s)

Marc Girondot

Examples

```
## Not run:
library(HelpersMG)
# Read all the .csv files from the current folder/directory
contentaslist <- read_folder(folder=".", wildcard="*.csv", read=read.csv2)
# Read all the files from the current folder/directory
contentaslist <- read_folder(folder=".", wildcard="*.*", read=read.csv2)
# Read two files from the current folder/directory
files <- c("filename1.csv", "filename2.csv")
contentaslist <- read_folder(folder=".", wildcard=files, read=read.csv2)

## End(Not run)
```

---

rSnbinom *Random generation for the sum of random variable with negative binomial distributions.*

Description

Random numbers for the sum of random variable with negative binomial distributions.

Usage

```
rSnbinom(n = 1, size = NULL, prob = NULL, mu = NULL)
```

Arguments

n	number of observations.
size	target for number of successful trials, or dispersion parameter (the shape parameter of the gamma mixing distribution). Must be strictly positive, need not be integer.
prob	probability of success in each trial. 0 < prob <= 1.
mu	alternative parametrization via mean.

Details

rSnbinom returns random numbers for the sum of random variable with negative binomial distributions

Value

rSnbinom returns random number

Author(s)

Marc Girondot

See Also

Other Distribution of sum of random variable with negative binomial distributions: dSnbinom, pSnbinom, qSnbinom

Examples

```
## Not run:
alpha <- c(2.1, 2.05, 2)
mu <- c(10, 30, 20)
rep <- 100000
distEmpirique <- rSnbinom(n=rep, size=alpha, mu=mu)
tabledistEmpirique <- rep(0, 301)
names(tabledistEmpirique) <- as.character(0:300)
tabledistEmpirique[names(table(distEmpirique))] <- table(distEmpirique)/rep

plot(0:300, dSnbinom(0:300, size=alpha, mu=mu, infinite=1000), type="h", bty="n",
   xlab="x", ylab="Density", ylim=c(0,0.02))
plot_add(0:300, tabledistEmpirique, type="l", col="red")
legend(x=200, y=0.02, legend=c("Empirical", "Theoretical"),
   text.col=c("red", "black"), bty="n")

## End(Not run)
```

ScalePreviousPlot *Return the scale of the previous plot*

Description

Return a list with the limits of the previous plot, the center, the range, and the position of label on this axe.

Usage

```
ScalePreviousPlot()
```

Details

ScalePreviousPlot returns the scale of the previous plot

Value

A list with xlim and ylim

Author(s)

Marc Girondot

See Also

Other plot and barplot functions: barplot_errbar, plot_add, plot_errbar

Examples

```
## Not run:
par(xaxs="i", yaxs="i")
plot(x=1:100, y=sin(1:100), type="l", bty="n", xlim=c(1,200), xlab="x", ylab="y")
xlim= ScalePreviousPlot()$xlim[1:2]
ylim= ScalePreviousPlot()$ylim[1:2]
par(xaxs="r", yaxs="i")
plot(x=1:100, y=sin(1:100), type="l", bty="n", xlim=c(1,200), xlab="x", ylab="y")
xlim= ScalePreviousPlot()$xlim[1:2]
ylim= ScalePreviousPlot()$ylim[1:2]
# Here is an example of the use of the label output
plot(x=1:100, y=sin(1:100), type="l", bty="n", xlim=c(1,200), xlab="", ylab="")
text(x=ScalePreviousPlot()$xlim["label"], y=ScalePreviousPlot()$ylim["center"],
  xpd=TRUE, "Legend for Y axes", pos=3, srt=90)
text(x=ScalePreviousPlot()$xlim["center"], y=ScalePreviousPlot()$ylim["label"],
  xpd=TRUE, "Legend for X axes", pos=1)

## End(Not run)
```

SEfromHessian *Standard error of parameters based on Hessian matrix*

Description

Standard error of parameters based on Hessian matrix.
The strategy is as follow:
First it tries to inverse the Hessian matrix. If it fails, it uses the near positive definite matrix of the Hessian.
So now the inverse of the Hessian matrix can be computed.
The diagonal of the inverse of the Hessian matrix is calculated. If all values are positive, the SEs are the square root of the inverse of the Hessian.
If not all values are positive, it will estimate the pseudo-variance matrix based on GILL & King (2004). It necessitates a Cholesky matrix.
If from some reason it fails (for example all SE are 0 in output), then the strategy of Rebonato and Jackel will be used to generate the Cholesky matrix.

Usage

```
SEfromHessian(a, hessian = FALSE)
```

Arguments

a	An Hessian matrix
hessian	If TRUE, return a list with the hessian and SE

Details

SEfromHessian returns standard error of parameters based on Hessian matrix

Value

SEfromHessian returns a vector with standard errors

Author(s)

Marc Girondot

References

GILL J. AND G. KING 2004. What to do when your Hessian is not invertible: Alternatives to model respecification in nonlinear estimation. Sociological Methods & Research 33: 54-87.

Rebonato and Jackel, "The most general methodology for creating a valid correlation matrix for risk management and option pricing purposes", Journal of Risk, Vol 2, No 2, 2000.

Examples

```
## Not run:
val=rnorm(100, mean=20, sd=5)
# Return -ln L of values in val in Gaussian distribution with mean and sd in par
fitnorm<-function(par, val) {
  -sum(dnorm(val, par["mean"], par["sd"], log = TRUE))
}
# Initial values for search
p<-c(mean=20, sd=5)
# fit the model
result <- optim(par=p, fn=fitnorm, val=val, method="BFGS", hessian=TRUE)
SE <- SEfromHessian(result$hessian)
library(MASS)
fitdistr(val, densfun = "normal")

## End(Not run)
```

`series.compare` *Data series comparison using Akaike weight*

Description

This function is used as a replacement of t.test() to not use p-value.

Usage

```
series.compare(..., criterion = c("BIC", "AIC", "AICc"),
  var.equal = TRUE)
```

Arguments

`...`	Series of data (at least two or data are in a table with series in different rows)
`criterion`	Which criterion is used for model selection. can be AIC, AICc or BIC
`var.equal`	Should the variances of all series being equal? Default TRUE

Details

series.compare compares series of data using Akaike weight.

Value

The probability that a single proportion model is sufficient to explain the data

Author(s)

Marc Girondot

References

Girondot, M., Guillon, J.-M., 2018. The w-value: An alternative to t- and X2 tests. Journal of Biostatistics & Biometrics 1, 1-4.

See Also

Other w-value functions: `compare`, `contingencyTable.compare`

Examples

```
## Not run:
library("HelpersMG")
A <- rnorm(100, 10, 2)
B <- rnorm(100, 11.1, 2)
series.compare(A, B, criterion = "BIC", var.equal=TRUE)
B <- B[1:10]
series.compare(A, B, criterion = "BIC", var.equal=TRUE)
```

```
A <- rnorm(100, 10, 2)
B <- rnorm(100, 10.1, 2)
C <- rnorm(100, 10.5, 2)
series.compare(A, B, C, criterion = "BIC", var.equal=TRUE)
B <- B[1:10]
series.compare(A, B, criterion = "BIC", var.equal=TRUE)
t.test(A, B, var.equal=TRUE)
# Example with a data.frame
series.compare(t(data.frame(A=c(10, 27, 19, 20, NA), B=c(10, 20, NA, NA, NA))))
# Test in the context of big data
A <- rnorm(10000, 10, 2)
B <- rnorm(10000, 10.1, 2)
series.compare(A, B, criterion = "BIC", var.equal=TRUE)
t.test(A, B, var.equal=TRUE)
############################
w <- NULL
p <- NULL

for (i in 1:1000) {

  A <- rnorm(50000, 10, 2)
  B <- rnorm(50000, 10.01, 2)
  w <- c(w, unname(series.compare(A, B, criterion = "BIC", var.equal=TRUE)[1]))
  p <- c(p, t.test(A, B, var.equal=TRUE)$p.value)

}

layout(mat = 1:2)
par(mar=c(4, 4, 1, 1)+0.4)
hist(p, main="", xlim=c(0, 1), las=1, breaks = (0:20)/20,
     freq=FALSE, xlab = expression(italic("p")*"-value"))
hist(w, main="", xlim=c(0, 1), las=1, breaks = (0:20)/20,
    freq=FALSE, xlab = expression(italic("w")*"-value"))
############################

x <- seq(from=8, to=13, by=0.1)

pv <- NULL
aw <- NULL
A <- rnorm(100, mean=10, sd=2)
B <- A-2

for (meanB in x) {
  pv <- c(pv, t.test(A, B, var.equal = FALSE)$p.value)
  aw <- c(aw, series.compare(A, B, criterion="BIC", var.equal = FALSE)[1])
  B <- B + 0.1
}

par(mar=c(4, 4, 2, 1)+0.4)
y <- pv
plot(x=x, y=y, type="l", lwd=2,
     bty="n", las=1, xlab="Mean B value (SD = 4)", ylab="Probability", ylim=c(0,1),
     main="")
```

```
y2 <- aw
lines(x=x, y=y2, type="l", col="red", lwd=2)

l1 <- which(aw>0.05)[1]
l2 <- max(which(aw>0.05))

aw[l1]
pv[l1]

aw[l2]
pv[l2]

l1 <- which(pv>0.05)[1]
l2 <- max(which(pv>0.05))

aw[l1]
pv[l1]

aw[l2]
pv[l2]

par(xpd=TRUE)
segments(x0=10-1.96*2/10, x1=10+1.96*2/10, y0=1.1, y1=1.1, lwd=2)
segments(x0=10, x1=10, y0=1.15, y1=1.05, lwd=2)
par(xpd=TRUE)
text(x=10.5, y=1.1, labels = "Mean A = 10, SD = 2", pos=4)

v1 <- c(expression(italic("p")*"-value"), expression("based on "*italic("t")*"-test"))
v2 <- c(expression(italic("w")*"-value for A"), expression("and B identical models"))
legend("topright", legend=c(v1, v2),
       y.intersp = 1,
       col=c("black", "black", "red", "red"), bty="n", lty=c(1, 0, 1, 0))

segments(x0=min(x), x1=max(x), y0=0.05, y1=0.05, lty=2)
par(xpd = TRUE)
text(x=13.05, y=0.05, labels = "0.05", pos=4)

## End(Not run)
```

`similar` *Test if two vectors contains the same elements independently of their order*

Description

Return TRUE only if all elements of x are present and only once in y.

Usage

```
similar(x, y, test.names = FALSE)
```

Arguments

x	A vector with numeric or string elements
y	A vector with numeric or string elements
test.names	Logical. If TRUE, the names of the vector elements must be also identical and unique

Value

A logical TRUE or FALSE

Author(s)

Marc Girondot

Examples

```
## Not run:
A <- c("A", "B", "C", "D")
B <- c("A", "B", "C", "D")
similar(A, B)
similar(B, A)
A <- c(x="A", y="B", z="C", k="D")
B <- c(x="A", y="B", z="C", l="D")
similar(B, A)
similar(A, B, test.names=TRUE)
A <- c(x="A", y="B", z="C", k="D")
B <- c(x="A", z="C", k="D", y="B")
similar(B, A)
similar(A, B, test.names=TRUE)

## End(Not run)
```

---

summary.mcmcComposite *Summarize the result of a mcmcComposite object*

Description

Summary for the result of a mcmcComposite object.

Usage

```
## S3 method for class 'mcmcComposite'
summary(object, chain = NULL, ...)
```

Arguments

object	A mcmcComposite object
chain	The chain to use
...	Not used

Details

summary.mcmcComposite get info on the result of a mcmcComposite object

Value

A summary of the result

Author(s)

Marc Girondot

See Also

Other mcmcComposite functions: MHalgoGen, as.mcmc.mcmcComposite, as.parameters, merge.mcmcComposite, plot.mcmcComposite

Examples

```
## Not run:
library(HelpersMG)
require(coda)
x <- rnorm(30, 10, 2)
dnormx <- function(data, x) {
 data <- unlist(data)
 return(-sum(dnorm(data, mean=x['mean'], sd=x['sd'], log=TRUE)))
}
parameters_mcmc <- data.frame(Density=c('dnorm', 'dlnorm'),
Prior1=c(10, 0.5), Prior2=c(2, 0.5), SDProp=c(1, 1),
Min=c(-3, 0), Max=c(100, 10), Init=c(10, 2), stringsAsFactors = FALSE,
row.names=c('mean', 'sd'))
mcmc_run <- MHalgoGen(n.iter=1000, parameters=parameters_mcmc, data=x,
likelihood=dnormx, n.chains=1, n.adapt=100, thin=1, trace=1)
plot(mcmc_run, xlim=c(0, 20))
plot(mcmc_run, xlim=c(0, 10), parameters="sd")
mcmcforcoda <- as.mcmc(mcmc_run)
#' heidel.diag(mcmcforcoda)
raftery.diag(mcmcforcoda)
autocorr.diag(mcmcforcoda)
acf(mcmcforcoda[[1]][,"mean"], lag.max=20, bty="n", las=1)
acf(mcmcforcoda[[1]][,"sd"], lag.max=20, bty="n", las=1)
batchSE(mcmcforcoda, batchSize=100)
# The batch standard error procedure is usually thought to
# be not as accurate as the time series methods used in summary
summary(mcmcforcoda)$statistics[,"Time-series SE"]
summary(mcmc_run)
as.parameters(mcmc_run)
lastp <- as.parameters(mcmc_run, index="last")
parameters_mcmc[,"Init"] <- lastp
# The n.adapt set to 1 is used to not record the first set of parameters
# then it is not duplicated (as it is also the last one for
# the object mcmc_run)
mcmc_run2 <- MHalgoGen(n.iter=1000, parameters=parameters_mcmc, data=x,
```

```
likelihood=dnormx, n.chains=1, n.adapt=1, thin=1, trace=1)
mcmc_run3 <- merge(mcmc_run, mcmc_run2)
####### no adaptation, n.adapt must be 0
parameters_mcmc[,"Init"] <- c(mean(x), sd(x))
mcmc_run3 <- MHalgoGen(n.iter=1000, parameters=parameters_mcmc, data=x,
likelihood=dnormx, n.chains=1, n.adapt=0, thin=1, trace=1)

## End(Not run)
```

`sun.info` *Estimate the time of sunrise and sunset according to longitude, latitude and date*

Description

Estimate the sun fates according to latitude and date.
Can be compared with the function `sunrise.set()` of package `StreamMetabolism`.

Usage

```
sun.info(date, latitude, longitude)
```

Arguments

`date`	A vector with the time at which sun fates are needed
`latitude`	The latitude at which estimate the sun fates
`longitude`	The longitude at which estimate the sun fates

Details

sun.info estimate the time of sunrise and sunset according to longitude, latitude and date

Value

A data.frame with information about daily sun

Author(s)

Marc Girondot <marc.girondot@u-psud.fr>

References

Teets, D.A. 2003. Predicting sunrise and sunset times. The College Mathematics Journal 34(4):317-321.

See Also

Other Periodic patterns of indices: `index.periodic`, `minmax.periodic`, `moon.info`, `tide.info`

Examples

```
## Not run:
# Generate a timeserie of time
date <- seq(from=as.Date("2000-01-01"), to=as.Date("2000-12-31"), by="1 day")
plot(date, sun.info(date, latitude=23, longitude=0)$day.length, bty="n",
 las=1, type="l", xlab="Ordinal days", ylab="Day length in hours")
plot(date, sun.info(date, latitude=23, longitude=0)$sunrise, bty="n",
 las=1, type="l", xlab="Ordinal days", ylab="Sun rise in hours")

## End(Not run)
```

`symbol.Female` *Plot a female symbol in the plotting region*

Description

Plot a female symbol in the plotting region.

Usage

```
symbol.Female(centerx, centery, rayonx, lwd = 2, col = "black")
```

Arguments

`centerx`	The x position of the center of the circle
`centery`	The y position of the center of the circle
`rayonx`	The size of the rayon in the scale of the x axis
`lwd`	The width of the line of the symbol
`col`	The color of the symbol

Details

symbol.Female plot a female symbol in the plotting region

Value

Nothing

Author(s)

Marc Girondot

See Also

Other Symbol: `symbol.Male`

Examples

```
## Not run:
plot(x=1:2, y=c(10,20), type="n", bty="n", xlab="", ylab="")

rayonx <- 0.01
centerx <- 1.2
centery <- 15

symbol.Male(centerx=centerx, centery = centery, rayonx=rayonx)
symbol.Female(centerx=centerx+0.5, centery = centery, rayonx=rayonx)

rayonx <- 0.03
centerx <- 1.2
centery <- 18

symbol.Male(centerx=centerx, centery = centery, rayonx=rayonx, lwd=3)
symbol.Female(centerx=centerx+0.5, centery = centery, rayonx=rayonx, lwd=3, col="red")

rayonx <- 0.05
centerx <- 1.4
centery <- 13

symbol.Male(centerx=centerx, centery = centery, rayonx=rayonx, lwd=4, col="blue")
symbol.Female(centerx=centerx+0.5, centery = centery, rayonx=rayonx, lwd=4, col="red")

## End(Not run)
```

---

symbol.Male *Plot a male symbol in the plotting region*

Description

Plot a male symbol in the plotting region.

Usage

```
symbol.Male(centerx, centery, rayonx, lwd = 2, col = "black")
```

Arguments

centerx	The x position of the center of the circle
centery	The y position of the center of the circle
rayonx	The size of the rayon in the scale of the x axis
lwd	The width of the line of the symbol
col	The color of the symbol

Details

symbol.Male plot a male symbol in the plotting region

Value

Nothing

Author(s)

Marc Girondot

See Also

Other Symbol: symbol.Female

Examples

```
## Not run:
plot(x=1:2, y=c(10,20), type="n", bty="n", xlab="", ylab="")

rayonx <- 0.01
centerx <- 1.2
centery <- 15

symbol.Male(centerx=centerx, centery = centery, rayonx=rayonx)
symbol.Female(centerx=centerx+0.5, centery = centery, rayonx=rayonx)

rayonx <- 0.03
centerx <- 1.2
centery <- 18

symbol.Male(centerx=centerx, centery = centery, rayonx=rayonx, lwd=3)
symbol.Female(centerx=centerx+0.5, centery = centery, rayonx=rayonx, lwd=3, col="red")

rayonx <- 0.05
centerx <- 1.4
centery <- 13

symbol.Male(centerx=centerx, centery = centery, rayonx=rayonx, lwd=4, col="blue")
symbol.Female(centerx=centerx+0.5, centery = centery, rayonx=rayonx, lwd=4, col="red")

## End(Not run)
```

---

symmetricize *Make a matrix symmetric*

Description

This function was part of the package ENA. This package is no more available and it cannot be installed from archive because some dependencies are no more available.

Usage

```
symmetricize(matrix, method = c("max", "min", "avg", "ld", "ud"),
  adjacencyList = FALSE)
```

Arguments

matrix	The matrix to make symmatric
method	The method to use to make the matrix symmetric. Default is to take the maximum. • "max" For each position, $m_{i,j}$, use the maxiumum of $(m_{i,j}, m_{j,i})$ • "min" For each position, $m_{i,j}$, use the minimum of $(m_{i,j}, m_{j,i})$ • "avg" For each position, $m_{i,j}$, use the mean: $(m_{i,j} + m_{j,i})/2$ • "ld" Copy the lower triangular portion of the matrix to the upper triangular portion. • "ud" Copy the upper triangular portion of the matrix to the lower triangular portion.
adjacencyList	Logical. If false, returns the symmetric matrix (the same format as the input). If true, returns an adjacency list representing the upper triangular portion of the adjacency matrix with addressing based on the row.names of the matrix provided.

Details

Make the matrix symmetric by making all "mirrored" positions consistent. A variety of methods are provided to make the matrix symmetrical.

Value

The symmetric matrix

Author(s)

Jeffrey D. Allen <Jeffrey.Allen@UTSouthwestern.edu>

Examples

```
#Create a sample 3x3 matrix
mat <- matrix(1:9, ncol=3)

#Copy the upper diagonal portion to the lower
symmetricize(mat, "ud")

#Take the average of each symmetric location
symmetricize(mat, "avg")
```

`tide.info` *Annual tide calendar for one particular location*

Description

The script extracts tide information from
`http://tides.mobilegeographics.com/` into a data.frame.
The presence of XLM package is required for this function.

Usage

```
tide.info(file = NULL, year = as.POSIXlt(Sys.time())$year + 1900,
  location = 0, latitude = NA, longitude = NA, tz = "")
```

Arguments

`file`	An html file from the site `http://tides.mobilegeographics.com/`
`year`	Year to get the calendar
`location`	Code based on `http://tides.mobilegeographics.com/`
`latitude`	The latitude of the tide information
`longitude`	The longitude of the tide information
`tz`	Timezone

Details

tide.info gets the annual tide calendar for one particular location.

Value

Return a data.frame with tide calendar:
Level is the tide level, Tide is the High or Low Tide information and Date.Time is the date/time in POSIXlt format.

Author(s)

Marc Girondot <marc.girondot@u-psud.fr>

See Also

Other Periodic patterns of indices: `index.periodic`, `minmax.periodic`, `moon.info`, `sun.info`

Examples

```
## Not run:
library("HelpersMG")
lat <- 5.74
long <- -54
Awala2004 <- tide.info(year=2004, longitude=long, latitude=lat, tz="America/Cayenne")
with(Awala2004, plot(Date.Time, Level, bty="n", las=1, type="l",
xlab=paste("Year", as.POSIXlt(Date.Time[1])$year+1900),
ylab="Tide level in m"))

## End(Not run)
```

`tnirp` *Read an ASCII text representation of a named or not vector object*

Description

Read an ASCII text representation of a named or not vector object.
Note that paste0(rev(c("p", "r", "i", "n", "t")), collapse="") = "tnirp"

Usage

```
tnirp(x, named = TRUE)
```

Arguments

`x`	A string or a vector of strings with value and possibly names.
`named`	TRUE if names are included.

Details

tnirp reads an ASCII text representation of a named or not vector object

Value

A vector

Author(s)

Marc Girondot

See Also

Other Characters: `asc`, `chr`, `d`

Examples

```
A <- structure(runif(26), .Names=letters)
text <- capture.output(A)
tnirp(text)

tnirp("         mu   mu_season         OTN       p1.09       p1.10       p1.11
 4.63215947 10.78627511  0.36108497  0.08292101 -0.52558196 -0.76430859
       p1.12       p1.13       p1.14       p1.15       p1.16       p1.17
       -0.75186542 -0.57632291 -0.58017174 -0.57048696 -0.56234135 -0.80645122
       p1.18       p1.19       p1.20       p1.21       p1.22       p1.23
       -0.77752524 -0.80909494 -0.56920540 -0.55317302  0.45757298 -0.64155368
       p1.24       p1.25       p1.26       p1.27       p1.28       p1.29
       -0.59119637 -0.66006794 -0.66582399 -0.66772684 -0.67351412 -0.66941992
       p1.30       p1.31       p1.32       p1.33       p1.34       p1.35
       -0.67038245 -0.68938726 -0.68889078 -0.68779016 -0.68604629 -0.68361820
       p1.36       p1.37       p2.09       p2.10       p2.11       p2.12
       -0.67045238 -0.66115613  2.55403149  2.31060620  2.31348160  2.20958757
       p2.13       p2.14       p2.15       p2.16       p2.17       p2.18
       2.14304918  2.19699719  2.30705457  2.18740019  2.32305811  2.31668302
       p2.19       p2.20       p2.21       p2.22       p2.23       p2.24
       1.99424288  2.06613445  2.38092301  2.40551276  2.31987342  2.30344402
       p2.25       p2.26       p2.27       p2.28       p2.29       p2.30
       2.26869058  2.25008836  2.23385204  2.22768782  2.25341904  1.77043360
       p2.31       p2.32       p2.33       p2.34       p2.35       p2.36
       2.21606813  2.21581431  2.21153872  2.21118013  2.21375660  2.21182196
       p2.37
       1.86137833 ")
tnirp("   27.89 289.99
      90.56", named=FALSE)
```

---

wget *Download a file from internet and save it locally*

Description

Download a file from internet and save it locally. This function is a wrapper for download.files() that keep the name identical and can get several files at once.

Usage

```
wget(url = stop("At least one internet adress is required"), ...)
```

Arguments

url	The url where to download file
...	The parameters send to download.file()

Details

wget download a file from internet and save it locally

Value

Nothing

Author(s)

Marc Girondot

Examples

```
## Not run:
library(HelpersMG)
# Save locally the files send in the parameter url
wget(c("https://cran.r-project.org/web/packages/HelpersMG/HelpersMG.pdf",
         "https://cran.r-project.org/web/packages/embryogrowth/embryogrowth.pdf"))

## End(Not run)
```

Index