Package ‘polyqtlR’

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Type Package

Title QTL Analysis in Autopolyploid Bi-Parental F1 Populations

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Description Quantitative trait loci (QTL) analysis and exploration of meiotic patterns in autopolyploid bi-parental F1 populations.
For all ploidy levels, identity-by-descent (IBD) probabilities can be estimated.
Significance thresholds, exploring QTL allele effects and visualising results are provided.
For background, see the 2018 dissertation of P.M. Bourke <doi:10.18174/444415>.

Depends R (>= 3.5.0)

Imports abind, doParallel, foreach, Hmisc, knitr, nlme, RColorBrewer,
Rcpp (>= 0.12.19), reshape2

Suggests igraph, mappoly, polymapR, rmarkdown

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LazyData true

LinkingTo Rcpp, RcppArmadillo

NeedsCompilation yes

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**bivTM**

**Description**

Bivalent transition matrix function

**Usage**

```r
bivTM(r)
```
Arguments

<table>
<thead>
<tr>
<th>r</th>
<th>recombination frequency</th>
</tr>
</thead>
</table>

BLUE

Calculate Best Linear Unbiased Estimates using linear mixed model from nlme package

Description

Calculation of BLUES from data frame of genotype names and phenotypes (assuming repeated measurements)

Usage

BLUE(data, model, random, genotype.ID)

Arguments

data | Data frame of genotype codes and corresponding phenotypes
model | The model specification of fixed terms, eg. Yield ~ Clones
random | The random component of the model (repeat structure, can be nested), eg. ~1 | Blocks if only Blocks are used
genotype.ID | The colname used to describe genotypes, e.g. "Clones"

Value

A data-frame with columns "geno" for the genotype names, and "blue" for the BLUES.

Examples

data("Phenotypes_4x")
blue <- BLUE(data = Phenotypes_4x, model = pheno~geno, random = ~1|year, genotype.ID = "geno")

BLUES.pheno

A data-frame of best linear unbiased predicted (BLUE) phenotypes (4x)

Description

A data-frame of best linear unbiased predicted (BLUE) phenotypes (4x)

Usage

BLUES.pheno

Format

An object of class data.frame with 50 rows and 2 columns.
Rcpp internal function Backward from forward-backward algorithm

Description
Rcpp internal function Backward from forward-backward algorithm

Usage
bx

Format
An object of class function of length 1.

check_cofactors

Description
The function check_cofactors initially fits all significant QTL positions as co-factors, both individually and in combination. Significance thresholds are re-estimated each time, yielding threshold-corrected LOD scores. If this leads to a change in the estimated position of QTL, or detection of subsequent peaks, a second round of co-factor inclusion is performed for all new QTL or novel QTL combinations. Finally, the multi-QTL model that maximises the individual significance of each QTL is returned as a data.frame. This can be directly passed to the function PVE to estimate the percentage variance explained by the full multi-QTL model and all possible sub-models. Note: this function estimates the most likely QTL positions by maximising the threshold-corrected LOD at QTL peaks. Non-additive interactions between QTL may be missed as a result. It is recommended to run a manual co-factor analysis as well, as described in the package vignette.

Usage
check_cofactors(
  IBD_list,
  Phenotype.df,
  genotype.ID,
  trait.ID,
  LOD_data,
  min_res = 20,
  ncores = 1,
  verbose = TRUE
)
check_cofactors

Arguments

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>IBD_list</td>
<td>List of IBD_probabilities as estimated using one of the various methods available (e.g. estimate_IBD).</td>
</tr>
<tr>
<td>Phenotype.df</td>
<td>A data.frame containing phenotypic values</td>
</tr>
<tr>
<td>genotype.ID</td>
<td>The colname of Phenotype.df that contains the population identifiers (F1 names) (must be a colname of Phenotype.df)</td>
</tr>
<tr>
<td>trait.ID</td>
<td>The colname of Phenotype.df that contains the response variable to use in the model (must be a colname of Phenotype.df)</td>
</tr>
<tr>
<td>LOD_data</td>
<td>Output of QTLscan function.</td>
</tr>
<tr>
<td>min_res</td>
<td>The minimum genetic distance (resolution) assumed possible to consider 2 linked QTL (on the same linkage group) as independent. By default a value of 20 cM is used. This is not to suggest that 20 cM is a realistic resolution in a practical mapping study, but it provides the function with a criterion to consider 2 significant QTL within this distance as one and the same. For this purpose, 20 cM seems a reasonable value to use. In practice, closely linked QTL will generally &quot;explain&quot; all the variation at nearby positions, making it unlikely to be able to disentangle their effects. QTL positions will vary slightly when co-factors are introduced, but again this variation is presumed not to exceed 20 cM either side.</td>
</tr>
<tr>
<td>ncores</td>
<td>How many CPU cores should be used in the evaluation? By default 1 core is used.</td>
</tr>
<tr>
<td>verbose</td>
<td>Logical, by default TRUE - should progress messages be printed to the console?</td>
</tr>
</tbody>
</table>

Value

Data frame with the following columns:

- LG: Linkage group identifier
- cM: CentiMorgan position
- deltaLOD: The difference between the LOD score at the peak and the significance threshold (always positive, otherwise the QTL would not be significant)
- CofactorID: A identifier giving the co-factor model used in detecting the QTL (if no co-factors were included then NA). The co-factor model is described by concatenating all co-factor positions with a '+' , so for example 1_10+4_20 would mean a co-factor model with 2 positions included as co-factors, namely 10 cM on linkage group 1 and 20 cM on linkage group 4.

Examples

data("IBD_4x","BLUEs.pheno","qtl_LODs.4x")
check_cofactors(IBD_list=IBD_4x,Phenotype.df=BLUEs.pheno,
genotype.ID="Geno",trait.ID="BLUE",LOD_data=qtl_LODs.4x)
**colour.bar**

*Add colour bar scale to heatplot*

**Description**

Function to generate a scale for heatplots

**Usage**

```r
colour.bar(
    col.data,
    min,
    max = -min,
    cex.ticks = 1.2,
    nticks = 11,
    ticks = seq(min, max, len = nticks),
    title = "",
    ylab = "",
    cex.lab = 1
)
```

**Arguments**

- `col.data`: vector of colours
- `min`: minimum colour
- `max`: maximum colour
- `cex.ticks`: size of ticks on colour bar
- `nticks`: number of ticks on colour bar
- `ticks`: vector of positions of ticks on colour bar
- `title`: optional title for colour bar
- `ylab`: optional y-axis label for colour bar
- `cex.lab`: size of labels on colour bar

---

**convert_mappoly_to_phased.maplist**

*Function to extract the phased map from a mappoly.map object*

**Description**

Convert MAPpoly.map object into a phased maplist, needed for IBD estimation

**Usage**

```r
convert_mappoly_to_phased.maplist(mappoly_object)
```
count_recombinations

**Arguments**

mappoly_object  An object of class ‘mappoly.map’, for example output of the function `mappoly::est_rf_hmm_sequential`

**Value**

A phased.maplist, with linkage group names LG1 etc. Each list item is a data.frame with columns marker, position followed by the phased map, coded in 1 and 0 for presence/absence of SNP (alternative) allele on parental homologues (h) numbered 1:ploidy for parent 1 and ploidy + 1 : 2*ploidy for parent 2.

**Examples**

data(maps.hexafake, package = "mappoly")
phased.maplist <- convert_mappoly_to_phased.maplist(maps.hexafake)

---

count_recombinations  

**Predict recombination breakpoints using IBD probabilities**

**Description**

The function `count_recombinations` returns a list of all predicted recombination breakpoints. The output can be passed using the argument `recombination_data` to the function `visualiseHaplo`, where the predicted breakpoints overlay the haplotypes. Alternatively, a genome-wide visualisation of the recombination landscape both per linkage group and per individual can be generated using the function `plotRecLS`, which can be useful in identifying problematic areas of the linkage maps, or problematic individuals in the population. Currently, recombination break-points are only estimated from bivalents in meiosis; any offspring resulting from a predicted multivalent is excluded from the analysis and will be returned with a NA value.

**Usage**

count_recombinations(IBD_list, plausible_pairing_prob = 0.4)

**Arguments**

IBD_list  List of IBD_probabilities as estimated using one of the various methods available (e.g. `estimate_IBD`).

plausible_pairing_prob  
The minimum probability of a pairing configuration needed to analyse an individual’s IBD data. The default setting of 0.4 is rather low - but accommodates scenarios where e.g. two competing plausible pairing scenarios are possible. In such situations, both pairing configurations (also termed "valencies") would be expected to have a probability close to 0.5. Both are then considered, and the output contains the probability of both situations. These can then be used to generate a probabilistic recombination landscape. If a more definite set of predictions is required, simply increase `plausible_pairing_prob` to eliminate such uncertainty. These individuals will then be returned with a NA value.
**Value**

A nested list corresponding to each linkage group. Within each LG, a list with 3 items is returned, specifying the plausible_pairing_prob, the map and the predicted recombinations in each individual in the mapping population. Per individual, all valencies with a probability greater than plausible_pairing_prob are returned, specifying both the Valent_probability and the best estimate of the cM position of the recombination_breakpoints involving pairs of homologues A, B, C etc. (in the order parent 1, parent 2). If no recombinations are predicted, a NA value is given instead.

**Examples**

```r
data("IBD_4x")
recom.ls <- count_recominations(IBD_4x)
```

---

## estimate_GIC

**Estimate the Genotypic Information Coefficient (GIC)**

**Description**

Function to estimate the GIC per homologue using IBD probabilities

**Usage**

```r
estimate_GIC(IBD_list)
```

**Arguments**

- `IBD_list` List of IBD probabilities

**Value**

A nested list; each list element (per linkage group) contains the following items:

- **GIC**: Matrix of GIC values estimated from the IBD probabilities
- **map**: Integrated linkage map positions of markers used in IBD calculation
- **parental_phase**: The parental marker phasing, coded in 1 and 0's

**Examples**

```r
data("IBD_4x")
GIC_4x <- estimate_GIC(IBD_list = IBD_4x)
```
estimate_IBD

Generate IBD probabilities from marker genotypes and a phased linkage map

Description

estimate_IBD is a function for creating identity-by-descent (IBD) probabilities. Two computational methods are offered: by default IBD probabilities are estimated using hidden Markov models, but a heuristic method based on Bourke et al. (2014) is also included. Basic input data for this function are marker genotypes (either discrete marker dosages (ie scores 0, 1, ..., ploidy representing the number of copies of the marker allele), or the probabilities of these dosages) and a phased linkage map. Details on each of the methods are included under method.

Usage

```r
estimate_IBD(
  input_type = "discrete",
  genotypes,
  phased_maplist,
  method = "hmm",
  remove_markers = NULL,
  ploidy,
  ploidy2 = NULL,
  parent1 = "P1",
  parent2 = "P2",
  individuals = "all",
  log = NULL,
  map_function = "haldane",
  bivalent_decoding = TRUE,
  error = 0.01,
  full_multivalent_hexa = FALSE,
  verbose = FALSE,
  ncores = 1,
  fix_threshold = 0.1,
  factor_dist = 1
)
```

Arguments

- **input_type**: Can be either one of 'discrete' or 'probabilistic'. For the former (default), dosage_matrix must be supplied, while for the latter probgeno_df must be supplied. Note that probabilistic genotypes can only be accepted if the method is default ('hmm').

- **genotypes**: Marker genotypes, either a 2d matrix of integer marker scores or a data.frame of dosage probabilities. Details are as follows:
• **discrete**: If `input_type` is 'discrete', genotypes is a matrix of marker dosage scores with markers in rows and individuals in columns. Both (marker) rownames and (individual or sample) colnames are needed.

• **probabilistic**: If `input_type` is 'probabilistic', genotypes is a data frame as read from the scores file produced by function `saveMarkerModels` of R package fitPoly, or alternatively, a data frame containing at least the following columns:
  - SampleName: Name of the sample (individual)
  - MarkerName: Name of the marker
  - P0: Probabilities of dosage score '0'
  - P1, P2... etc.: Probabilities of dosage score '1' etc. (up to max offspring dosage, e.g. P4 for tetraploid population)

`phased_maplist` A list of phased linkage maps, the output of `polymapR::create_phased_maplist`

`method` The method used to estimate IBD probabilities, either "hmm" or "heur". By default, the Hidden Markov Model (hmm) method is used. This uses an approach developed by Zheng et al (2016), and implemented in the 'TetraOrigin' package. However, unlike the original TetraOrigin software, it does not re-estimate parental linkage phase, as this is assumed to have been generated during map construction. Alternatively, a heuristic algorithm can be employed (`method = "heur"`), providing computational efficiency at higher ploidy levels (hexaploid, octoploid etc.), but at the cost of some accuracy. If `method = "hmm"` is specified, only diploid, triploid, autotetraploid and autohexaploid populations are currently allowed, while `method = "heur"` caters for all possible ploidy levels. Furthermore, the argument `bivalent_decoding` can only be set to `FALSE` in the case of the 'hmm' method (i.e. allowing for the possibility of multivalent formation and double reduction).

`remove_markers` Optional vector of marker names to remove from the maps. Default is NULL.

`ploidy` Integer. Ploidy of the organism.

`ploidy2` Optional integer, by default NULL. Ploidy of parent 2, if different from parent 1.

`parent1` Identifier of parent 1, by default assumed to be "P1"

`parent2` Identifier of parent 2, by default assumed to be "P2"

`individuals` By default "all" offspring are included, but otherwise a subset can be selected, using a vector of offspring indexing numbers (1,2, etc.) according to their order in `dosage_matrix`

`log` Character string specifying the log filename to which standard output should be written. If NULL log is send to stdout.

`map_function` Mapping function to use when converting map distances to recombination frequencies. Currently only "haldane" or "kosambi" are allowed.

`bivalent_decoding` Option to consider only bivalent pairing during formation of gametes (ignored for diploid populations, as only bivalents possible there), by default TRUE

`error` The (prior) probability of errors in the offspring dosages, usually assumed to be small but non-zero
full_multivalent_hexa

Option to allow multivalent pairing in both parents at the hexaploid level, by default FALSE. Note that if TRUE, a very large available RAM may be required (>= 32Gb) to process the data.

verbose

Logical, by default TRUE. Should progress messages be written?

cores

How many CPU cores should be used in the evaluation? By default 1 core is used.

fix_threshold

If method = "heur", the threshold to fix the IBD probabilities while correcting for the sum of probabilities.

factor_dist

If method = "heur", the factor by which to increase or decrease the recombination frequencies as calculated from the map distances.

Value

A list of IBD probabilities, organised by linkage group (as given in the input phased_maplist). Each list item is itself a list containing the following:

- IBDtype: The type of IBD; for this function only "genotypeIBD" are calculated.
- IBDarray: A 3d array of IBD probabilities, with dimensions marker, genotype-class and F1 individual.
- map: A 3-column data-frame specifying chromosome, marker and position (in cM)
- parental_phase: Phasing of the markers in the parents, as given in the input phased_maplist
- marginal.likelihoods: A list of marginal likelihoods of different valencies if method "hmm" was used, otherwise NULL
- valency: The predicted valency that maximised the marginal likelihood, per offspring. For method "heur", NULL
- offspring: Offspring names
- biv_dec: Logical, whether bivalent decoding was used in the estimation of the F1 IBD probabilities.
- gap: The size of the gap (in cM) used when interpolating the IBD probabilities. See function spline_IBD for details.
- genocodes: Ordered list of genotype codes used to represent different genotype classes.
- pairing: log likelihoods of each of the different pairing scenarios considered (can be used e.g. for post-mapping check of preferential pairing)
- ploidy: ploidy of parent 1
- ploidy2: ploidy of parent 2
- method: The method used, either "hmm" (default) or "heur". See argument method
- error: The error prior used, if method "hmm" was used, otherwise NULL

References

• Bourke P.M. (2014) QTL analysis in polyploids: Model testing and power calculations. Wageningen University (MSc thesis)

Examples

```r
data("phased_maplist.4x", "SNP_dosages.4x")
estimate_IBD(phased_maplist=phased_maplist.4x, genotypes=SNP_dosages.4x, ploidy=4)
```

---

exploreQTL

**Explore the possible segregation type of a QTL peak using Schwarz Information Criterion**

Description

Function to explore the possible segregation type at a QTL position using the Schwarz Information Criterion

Usage

```r
e xploreQTL(
  IBD_list,
  Phenotype.df,
  genotype.ID,
  trait.ID,
  linkage_group,
  LOD_data,
  cM = NULL,
  QTLconfig = NULL,
  plotBIC = TRUE,
  deltaBIC = 6,
  testAllele_Effects = TRUE,
  log = NULL
)
```

Arguments

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>IBD_list</td>
<td>List of IBD probabilities</td>
</tr>
<tr>
<td>Phenotype.df</td>
<td>A data.frame containing phenotypic values</td>
</tr>
<tr>
<td>genotype.ID</td>
<td>The colname of Phenotype.df that contains the population identifiers (F1 names) (must be a colname of Phenotype.df)</td>
</tr>
<tr>
<td>trait.ID</td>
<td>The colname of Phenotype.df that contains the response variable to use in the model (must be a colname of Phenotype.df)</td>
</tr>
</tbody>
</table>
linkage_group Numeric identifier of the linkage group being tested, based on the order of IBD_list. Only a single linkage group is allowed.

LOD_data Output of QTLscan function

cM By default NULL, in which case the position of maximum LOD score is taken as the position of interest. Otherwise, the cM position to be explored.

QTLconfig Nested list of homologue configurations and modes of action of QTL to be explored and compared, the output of segMaker. Note that a default List is available of all possible bi-allelic QTL if none is provided. Each list element is itself a list with components

- homs: a vector of length at least 1, describing the proposed homologues the functional allele Q is on
- mode: Vector of same length as homs with codes "a" for additive and "d" for dominant.

plotBIC Logical, with default TRUE - should the calculated BIC values be plotted?

deltaBIC Numeric, by default 6. Configurations within this distance of the minimum BIC are considered plausible.

testAllele_Effects Logical, with default TRUE - should the effects of the different alleles be tested using the most likely QTL configuration?

log Character string specifying the log filename to which standard output should be written. If NULL log is send to stdout.

Value

List with the following items:

- BIC: Vector of BIC values corresponding to elements of QTLconfig provided for testing
- Allele.effects: Summary of the means and standard errors of groups with (+) and without(-) the specified allele combinations for the most likely QTLconfig if testAllele_Effects = TRUE (NULL otherwise).

Examples

data("IBD_4x","BLUEs.pheno","qtl_LODs.4x")
exploreQTL(IBD_list = IBD_4x,
Phenotype.df = BLUEs.pheno,
genotype.ID = "Geno",
trait.ID = "BLUE",
linkage_group = 1,
LOD_data = qtl_LODs.4x)
**Description**

The method of "quick-and-dirty" IBD estimation was originally developed by Bourke (2014) for tetraploid data only, and was subsequently generalised by van Geest et al. (2017). Can be useful for a first quick analysis, particularly in large hexaploid datasets. However, the higher accuracy of IBD probabilities generated by `hmm_IBD` makes that function the preferred choice.

**Usage**

```r
fast_IBD(phased_maplist, dosage_matrix, map_function = "haldane", ploidy, ploidy2 = NULL, fix_threshold = 0.1, factor_dist = 1, ncores = 1)
```

**Arguments**

- `phased_maplist`: A list of linkage maps calculated by `polymapR::create_phased_maplist`
- `dosage_matrix`: An integer matrix with markers in rows and individuals in columns
- `map_function`: The mapping function to calculate recombination frequency based on map distance (haldane or kosambi)
- `ploidy`: Ploidy level of parents or of the first parent
- `ploidy2`: Ploidy level of the second parent. By default `NULL`, if parents have equal ploidy levels.
- `fix_threshold`: The threshold to fix the IBD probabilities while correcting for the sum of probabilities.
- `factor_dist`: Factor to increase or decrease the recombination frequencies as calculated from the map distances.
- `ncores`: Number of cores to use for multi-core processing.

**Value**

A nested list (with the same length as `phased_maplist`). Each list element contains the following items:

- `IBDtype`: Always "haplotypeIBD" for the output of this function
IBDarray An array of IBD probabilities. The dimensions of the array are: markers, homologues and individuals.
map Integrated linkage map positions of markers used in IBD calculation
parental_phase The parental marker phasing, coded in 1 and 0's
biv_dec NULL
gap The gap size used in IBD interpolation, by default NULL. See spline_IBD
genocodes NULL
pairing NULL
ploidy ploidy of parent 1
ploidy2 ploidy of parent 2
method The method used, here "heur" (heuristic)
error The error prior used, not relevant here thus NULL

References
Bourke P.M. (2014) QTL analysis in polyploids: Model testing and power calculations. Wageningen University (MSc thesis)

Examples

data("phased_maplist.4x","SNP_dosages.4x")
IBD_list.4x <- fast_IBD(phased_maplist = phased_maplist.4x,
dosage_matrix = SNP_dosages.4x,
ploidy = 4)

---

fast_permute Extension of the QTLscan function, offering an optimised permutation test when the experimental setting (i.e. phenotype structure) is simple

Description
Function to run optimised QTL permutation test using IBD probabilities

Usage

fast_permute(  
IBD_list,  
Phenotype.df,  
genotype.ID,  
trait.ID,  
ploidy,  
ploidy2 = NULL,  
N_perm = 1000,  
alpha = 0.05,  
ncores = 1,
Arguments

IBD_list List of IBD probabilities
Phenotype.df A data.frame containing phenotypic values
genotype.ID The colname of Phenotype.df that contains the offspring identifiers (F1 names)
trait.ID The colname of Phenotype.df that contains the response variable to use in the model
ploidy Integer. The ploidy of parent 1
ploidy2 The ploidy of parent 2, by default NULL i.e. assumed (unless specified) to be equal to the ploidy of parent 1.
N_perm The number of permutations to run, by default this is 1000.
alpha The P-value to be used in the selection of a threshold, by default 0.05 (i.e. the 0.95 quantile).
ncores Number of cores to use, by default 1 only. Works both for Windows and UNIX (using doParallel). Use parallel::detectCores() to find out how many cores you have available.
verbose Logical, by default TRUE. Should messages be printed during running?
log Character string specifying the log filename to which standard output should be written. If NULL log is send to stdout.

Value

A nested list; each list element (per linkage group) contains the following items:

- QTL.res: Single matrix of QTL results with columns chromosome, position, LOD
- Perm.res: List of the results of the permutation test, with (at least) list items "quantile", "threshold" and "scores". Quantile refers to which quantile of scores was used to determine the threshold. Note that scores are each of the maximal LOD scores across the entire genome scan per permutation, thus returning a genome-wide threshold rather than a chromosome-specific threshold. If the latter is preferred, restricting the IBD_list to a single chromosome and re-running the permutation test will provide the desired threshold.
- Map: Original map of genetic marker positions upon which the IBDs were based, most often used for adding rug of marker positions to QTL plots.

Examples

data("IBD_4x","Phenotypes_4x")
qtl_LODs.4x <- fast_permute(IBD_list = IBD_4x,
                          Phenotype.df = Phenotypes_4x,
genotype.ID = "geno",
trait.ID = "pheno",
ploidy = 4)
findPeak

Function to find the position of maximum LOD on a particular linkage group

Description

Given QTL output, this function returns the position of maximum LOD for a specified linkage group.

Usage

findPeak(LOD_data, linkage_group, verbose = TRUE)

Arguments

LOD_data Output of QTLscan function.
linkage_group Numeric identifier of the linkage group being tested, based on the order of IBD_list. Only a single linkage group is allowed.
verbose Should messages be written to standard output? By default TRUE.

Examples

data("qtl_LODs.4x")
findPeak(LOD_data=qtl_LODs.4x,linkage_group=1)

findQTLpeaks

Extract QTL peak positions from the results of a QTL scan

Description

Function to find QTL peaks from output of a QTL scan

Usage

findQTLpeaks(LOD_data)

Arguments

LOD_data Output of QTLscan function
findSupport

Function to find a LOD - x support interval around a QTL position

Description

Given QTL output, this function returns the LOD - x support for a specified linkage group, taking the maximum LOD position as the desired QTL peak.

Usage

findSupport(LOD_data, linkage_group, LOD_support = 2)

Arguments

LOD_data Output of QTLscan function.
linkage_group Numeric identifier of the linkage group being tested, based on the order of IBD_list. Only a single linkage group is allowed.
LOD_support The level of support around a QTL peak, by default 2 (giving a LOD - 2 support interval, the range of positions with a LOD score within 2 LOD units of the maximum LOD on that linkage group).

Examples

data("qtl_LODs.4x")
findSupport(LOD_data=qtl_LODs.4x,linkage_group=1)

fx

Rcpp internal function Forward from forward-backward algorithm

Description

Rcpp internal function Forward from forward-backward algorithm

Usage

fx

Format

An object of class function of length 1.
GIC_4x  
*A list of GIC estimates (4x)*

**Description**  
A list of GIC estimates (4x)

**Usage**  
GIC_4x

**Format**  
An object of class `list` of length 2.

hexa.list  
*A list of hexaploid bivalent pairing configurations*

**Description**  
A list of hexaploid bivalent pairing configurations

**Usage**  
hexa.list

**Format**  
An object of class `list` of length 15.

hexTM  
*Hexavalent transition matrix function*

**Description**  
Hexavalent transition matrix function

**Usage**  
hexTM(r)

**Arguments**  

- `r`  
  recombination frequency
hmm_IBD

Generate IBD probabilities from marker genotypes and a phased linkage map using HMM

Description

hmm_IBD is a function for creating identity-by-descent (IBD) probabilities using hidden Markov models, from marker genotypes (either discrete marker dosages (ie scores 0, 1, ..., ploidy representing the number of copies of the marker allele), or the probabilities of these dosages) and a phased linkage map. Unlike the original TetraOrigin software, it does not re-estimate parental linkage phase, and has been generalised for use in diploid, triploid, tetraploid and hexaploid populations.

Usage

hmm_IBD(
  input_type = "discrete",
  genotypes,
  phased_maplist,
  remove_markers = NULL,
  ploidy,
  ploidy2 = NULL,
  parent1 = "P1",
  parent2 = "P2",
  individuals = "all",
  log = NULL,
  map_function = "haldane",
  bivalent_decoding = TRUE,
  error = 0.01,
  full_multivalent_hexa = FALSE,
  verbose = FALSE,
  ncores = 1
)

Arguments

input_type Can be either one of ‘discrete’ or ‘probabilistic’. For the former (default), dosage_matrix must be supplied, while for the latter probgeno_df must be supplied

genotypes Marker genotypes, either a 2d matrix of integer marker scores or a data.frame of dosage probabilities. Details are as follows:

• discrete: If input_type is 'discrete', genotypes is a matrix of marker dosage scores with markers in rows and individuals in columns. Both (marker) rownames and (individual or sample) colnames are needed.

• probabilistic: If input_type is 'probabilistic', genotypes is a data frame as read from the scores file produced by function saveMarkerModels of R package fitPoly, or alternatively, a data frame containing at least the following columns:
- SampleName : Name of the sample (individual)
- MarkerName : Name of the marker
- P0 : Probabilities of dosage score '0'
- P1, P2... etc. : Probabilities of dosage score '1' etc. (up to max offspring dosage, e.g. P4 for tetraploid population)

**phased_maplist**
A list of phased linkage maps, the output of `polymapR::create_phased_maplist`

**remove_markers**
Optional vector of marker names to remove from the maps. Default is NULL.

**ploidy**
Integer. Ploidy of the organism.

**ploidy2**
Optional integer, by default NULL. Ploidy of parent 2, if different from parent 1.

**parent1**
Identifier of parent 1, by default assumed to be "P1"

**parent2**
Identifier of parent 2, by default assumed to be "P2"

**individuals**
By default "all" offspring are included, but otherwise a subset can be selected, using a vector of offspring indexing numbers (1,2, etc.) according to their order in `dosage_matrix`

**log**
Character string specifying the log filename to which standard output should be written. If NULL log is send to stdout.

**map_function**
Mapping function to use when converting map distances to recombination frequencies. Currently only "haldane" or "kosambi" are allowed.

**bivalent_decoding**
Option to consider only bivalent pairing during formation of gametes (ignored for diploid populations, as only bivalents possible there), by default TRUE

**error**
The (prior) probability of errors in the offspring dosages, usually assumed to be small but non-zero

**full_multivalent_hexa**
Option to allow multivalent pairing in both parents at the hexaploid level, by default FALSE. Note that if TRUE, a very large available RAM may be required (>= 32Gb) to process the data.

**verbose**
Logical, by default TRUE. Should progress messages be written?

**ncores**
How many CPU cores should be used in the evaluation? By default 1 core is used.

**Value**

A list of IBD probabilities, organised by linkage group (as given in the input `phased_maplist`). Each list item is itself a list containing the following:

- **IBDtype**: The type of IBD; for this function only "genotypeIBD" are calculated.
- **IBDarray**: A 3d array of IBD probabilities, with dimensions marker, genotype-class and F1 individual.
- **map**: A 3-column data-frame specifying chromosome, marker and position (in cM)
- **parental_phase**: Phasing of the markers in the parents, as given in the input `phased_maplist`
- **biv_dec**: Logical, whether bivalent decoding was used in the estimation of the F1 IBD probabilities.
**IBD_4x**

- gap: The size of the gap (in cM) used when interpolating the IBD probabilities, if performed.
- genocodes: Ordered list of genotype codes used to represent different genotype classes.
- pairing: log likelihoods of each of the different pairing scenarios considered (can be used e.g. for post-mapping check of preferential pairing)
- ploidy: ploidy of parent 1
- ploidy2: ploidy of parent 2
- method: The method used, here "hmm" (Hidden Markov Model)
- error: The error prior used

**References**


**Examples**

```r
data("phased_maplist.4x", "SNP_dosages.4x")
hmm_IBD(phased_maplist=phased_maplist.4x, genotypes=SNP_dosages.4x, ploidy=4)
```

<table>
<thead>
<tr>
<th>IBD_4x</th>
<th>A list of identity-by-descent probabilities (4x)</th>
</tr>
</thead>
</table>

**Description**

A list of identity-by-descent probabilities (4x)

**Usage**

```r
IBD_4x
```

**Format**

An object of class list of length 2.
import_IBD

Import IBD probabilities as estimated by TetraOrigin

Description

Imports the summarised IBD probability output of TetraOrigin (which estimates IBD probabilities at all marker positions), and interpolates these at a grid of positions at user-defined spacing.

Usage

import_IBD(
  folder = NULL,
  filename.vec,
  bivalent_decoding = TRUE,
  error = 0.01,
  log = NULL
)

Arguments

folder
  The path to the folder in which the TetraOrigin output is contained, default is NULL if files are in working directory.

filename.vec
  A vector of the character filename(s) of the .csv file(s) containing the output of TetraOrigin. Should be in order according to LG/ chromosome numbering.

bivalent_decoding
  Logical, if TRUE then only bivalent pairing was allowed in TetraOrigin, specify FALSE if multivalent pairing was also allowed.

error
  The offspring error prior used in the offspring decoding step, here assumed to be 0.01

log
  Character string specifying the log filename to which standard output should be written. If NULL log is send to stdout.

Value

Returns a list with the following items:

IBDtype : Always "genotypeIBD" for the output of TetraOrigin

IBDarray : An array of IBD probabilities. The dimensions of the array are: markers, genotype classes and individuals.

map : Integrated linkage map positions of markers used in IBD calculation

parental_phase :
  The parental marker phasing as used by TetraOrigin, recoded in 1 and 0’s

marginal.likelihoods :
  A list of marginal likelihoods of different valencies, currently NULL
**impute_dosages**

The predicted valency that maximised the marginal likelihood, per offspring. Currently NULL

**offspring**

Offspring names

**biv_dec**

Logical, the bivalent_decoding parameter specified.

**gap**

The gap size used in IBD interpolation, by default NULL. See `spline_IBD`

**genocodes**

Ordered list of genotype codes used to represent different genotype classes.

**pairing**

Log likelihoods of each of the different pairing scenarios considered (can be used e.g. for post-mapping check of preferential pairing)

**ploidy**

The ploidy of parent 1, by default assumed to be 4

**ploidy2**

The ploidy of parent 2, by default assumed to be 4

**method**

The method used, always returned as "hmm_TO" (Hidden Markov Model TetraOrigin)

**error**

The error prior used in the calculation, assumed to be 0.01

---

**impute_dosages**

Re-estimate marker dosages given IBD input estimated using a high error prior.

---

**Description**

Function to correct marker dosage scores given a list of previously estimated IBD probabilities. This may prove useful to correct genotyping errors. Running the `estimate_IBD` function with a high error prior (e.g. 0.6) will result in suppressed predictions of double recombination events, associated with genotyping errors. By forcing the HMM to penalise double recombinations heavily, a smoothed haplotype landscape is achieved in which individual genotype observations are down-weighted. This smoothed output is then used to re-estimate marker dosages, dependent on (correct) parental scores.

**Usage**

```r
impute_dosages(
  IBD_list, 
  dosage_matrix, 
  parent1 = "P1", 
  parent2 = "P2", 
  rounding_error = 0.05, 
  min_error_prior = 0.1, 
  verbose = TRUE
)
```
Arguments

IBD_list  List of IBD probabilities

dosage_matrix  An integer matrix with markers in rows and individuals in columns. Note that probabilistic genotypes are not currently catered for here.

parent1 The identifier of parent 1, by default "P1"

parent2 The identifier of parent 2, by default "P2"

rounding_error The maximum deviation from an integer value that an inputed value can have, by default 0.05. For example, an imputed score of 2.97 or 3.01 would both be rounded to a dosage of 3, while 2.87 would be deemed too far from an integer score, and would be made missing. If you find the output contains too many missing values, a possibility would be to increase the rounding_error. However this may also introduce more errors in the output!

min_error_prior Suggestion for a suitably high error prior to be used in IBD calculations to ensure IBD smoothing is achieved. If IBD probabilities were estimated with a smaller error prior, the function aborts.

verbose Should messages be written to standard output?

Examples

## Not run:
# Toy example only, as this will result in an Error: the original error prior was too low
data("IBD_4x","SNP_dosages.4x")
impute_dosages(IBD_list=IBD_4x,dosage_matrix=SNP_dosages.4x)

## End(Not run)

---

list.depth  Find depth of a list

Description

Recursive function checking list depth to determine subsequent subsetting rules

Usage

list.depth(obj, objdepth = 0)

Arguments

obj An input object, may be a (nested) list

objdepth Counter to record how deep the recursion has gone
### mapseq

*Generate a sequence of map positions for splining function*

#### Description

Function to return a sequence of map positions at steps of size gap from given input

#### Usage

```r
mapseq(cMvect, gap)
```

#### Arguments

- `cMvect`: Vector of map positions
- `gap`: Gap size in cM

---

### meiosis_report

*Generate a 'report' of predicted meiotic behaviour in an F1 population*

#### Description

Function to extract the chromosome pairing predictions as estimated by `estimate_IBD`. Apart from producing an overview of the pairing during parental meiosis (including counts of multivalents, per linkage group per parent), the function also applies a simple chi-squared test to look for evidence of non-random pairing behaviour from the bivalent counts (deviations from a polysomic model)

#### Usage

```r
meiosis_report(IBD_list, visualise = TRUE, precision = 2)
```

#### Arguments

- `IBD_list`: List of IBD probabilities as estimated by `estimate_IBD` using method 'hmm', or externally (e.g. using TetraOrigin)
- `visualise`: Logical, by default TRUE, in which case a plot of the pairing results is produced per LG. In order to flag extreme deviations from the expected numbers (associated with polysomic inheritance, considered the Null hypothesis), barplots are coloured according to the level of significance of the X2 test. Plots showing red bars indicate extreme deviations from a polysomic pattern.
- `precision`: To how many decimal places should summed probabilities per bivalent pairing be rounded? By default 2.
Value

The function returns a nested list, with one element per linkage group in the same order as the input IBD list. Per linkage group, a list is returned containing the following components:

- **P1_multivalents**: The count of multivalents in parent 1 (only relevant if `bivalent_decoding = FALSE` during IBD calculation)
- **P2_multivalents**: Similarly, the count of multivalents in parent 2
- **P1_pairing**: The counts of each bivalent pairing predicted in parent 1, with an extra column Pr(X2) which gives the p-value of the X2 test of the off-diagonal terms in the matrix. In the case of a tetraploid, pairing A with B automatically implies C with D pairing, so the count table contains a lot of redundancy. The table should be read using both row and column names, so row A and column B corresponds to the count of individuals with A and B pairing (and hence C and D pairing). In a hexaploid, A-B pairing does not imply a particular pairing configuration in the remaining homologues. In this case, row A and column B is the count of individuals where A and B were predicted to have paired, summed over all three bivalent configurations with A and B paired (AB-CD-EF, AB-CE-DF, AB-CF,DE).
- **P2_pairing**: Same as P1_pairing, except using parent 2
- **ploidy**: The ploidy of parent 1
- **ploidy2**: The ploidy of parent 2

Examples

data("IBD_4x")
mr.ls<-meiosis_report(IBD_list = IBD_4x)

---

mr.ls

A list of pairing predictions (4x)

Description

A list of pairing predictions (4x)

Usage

mr.ls

Format

An object of class list of length 2.
### NettletonDoerge

**Description**

Nettleton and Doerge 2000

**Usage**

NettletonDoerge\( (N = 100, \alpha = 0.05, \gamma = 0.05) \)

**Arguments**

<table>
<thead>
<tr>
<th>N</th>
<th>number of permutations</th>
</tr>
</thead>
<tbody>
<tr>
<td>alpha</td>
<td>The threshold level, ie. ((1 - \alpha)) quantile of sorted LOD scores defines threshold</td>
</tr>
<tr>
<td>gamma</td>
<td>The confidence interval specifier, usually 0.05</td>
</tr>
</tbody>
</table>

### Nstates.fun

**Description**

Error handling on ploidy and ploidy2, and determine the number of genotype classes for general ploidy level and pairing behaviour

**Usage**

Nstates.fun(\(biv\_dec, pl, pl2\))

**Arguments**

<table>
<thead>
<tr>
<th>biv_dec</th>
<th>Either TRUE for bivalents only or FALSE to also allow quadrivalents and double reduction</th>
</tr>
</thead>
<tbody>
<tr>
<td>pl</td>
<td>ploidy level of parent 1, assumed to be even and greater than 4</td>
</tr>
<tr>
<td>pl2</td>
<td>ploidy level of parent 2, assumed to be even and greater than 2</td>
</tr>
</tbody>
</table>
**phased_maplist.4x**  
*A list of phased maps (4x)*

### Description

A list of phased maps (4x)

### Usage

```r
phased_maplist.4x
```

### Format

An object of class `list` of length 2.

---

**Phenotypes_4x**  
*A data-frame of phenotypes (4x)*

### Description

A data-frame of phenotypes (4x)

### Usage

```r
Phenotypes_4x
```

### Format

An object of class `data.frame` with 150 rows and 3 columns.

---

**plotLinearQTL**  
*Plot the results of genome-wide QTL analysis along a single track*

### Description

QTL plotting function that plots output of `QTLscan` function along a single track, useful for overlaying plots. Only works for scan over multiple chromosomes.
plotLinearQTL

Usage

plotLinearQTL(
  LOD_data,
  inter_chm_gap = 5,
  overlay = FALSE,
  ylimits = NULL,
  sig.unit = "LOD",
  plot_type = c("lines", "points"),
  add_xaxis = TRUE,
  add_rug = TRUE,
  add_thresh = TRUE,
  override_thresh = NULL,
  thresh.lty = 3,
  thresh.lwd = 2,
  thresh.col = "darkred",
  return_plotData = FALSE,
  show_thresh_CI = TRUE,
  use_LG_names = FALSE,
  axis_label.cex = 1,
  custom_LG_names = NULL,
  LGdiv.col = "gray42",
  ...
)

Arguments

LOD_data Output of QTLscan function.
inter_chm_gap The gap size (in cM) between successive chromosomes - by default a gap of 5 cM is used.
overlay Add to an existing plot (should be produced by a comparable call to this function) or not? By default FALSE, in which case a new plot is drawn. Can be useful for displaying results of multiple analyses together. However, an alternative approach, when significance thresholds have been calculated for multiple comparable scans, that plots be rescaled so that significance thresholds overlap perfectly. For this, the plotLinearQTL_list function is advised.
ylimits Use to specify ylimits of plot region, though by default NULL in which case a suitable plot region is automatically used.
sig.unit Label to use on the y-axis for significance units, by default assumed to be LOD score.
plot_type Plots can be either in line drawings ("lines") or scatter plot format ("points").
add_xaxis Should an x-axis be drawn? If multiple QTL analyses are performed on different traits, specifying this to be FALSE and using par(mar=c(0, 4.1, 4.1, 2.1)) allows subsequent plots to be neatly stacked.
add_rug Logical, by default TRUE - should original marker points be added to plot?
add_thresh Logical, by default TRUE - should a significance threshold be added to plot?
override_thresh

By default NULL. Can be used to specify a value for the significance threshold, overriding any stored in LOD_data.

thresh.lty

Gives user control over the line type of the significance threshold to be drawn.

thresh.lwd

Gives user control over the line width of the significance threshold to be drawn.

thresh.col

Gives user control over the line colour of the significance threshold to be drawn.

return_plotData

Logical, by default FALSE. If TRUE, then the x and y coordinates of the plot data are returned, which can be useful for subsequent plot manipulations and overlays.

show_thresh_CI

Logical, by default TRUE. Should confidence interval bounds around LOD threshold be shown?

use_LG_names

Logical, by default FALSE. Should original character LG names be used as axis labels, or should numbering be used instead?

axis_label.cex

Argument to adjust the size of the axis labels, can be useful if there are many linkage groups to plot

custom_LG_names

Specify a vector that contains custom linkage group names. By default NULL

LGdiv.col

Colour of dividing lines between linkage groups, by default grey.

...

Arguments passed to plot, and lines or points as appropriate (see argument plot_type).

Value

The plot data, if return_plotData = TRUE. Otherwise NULL.

Examples

```r
## Not run:
data("qtl_LODs.4x")
plotLinearQTL(LOD_data = qtl_LODs.4x)
## End(Not run)
```

plotLinearQTL_list

Overlay the results of a number of genome-wide QTL analysis for which significance thresholds are available.

Description

Extension of the plotLinearQTL function, taking as input a list generated from combining the output of QTLscan. Its distinguishing characteristic is that overlaid plots are re-scaled so that the significance thresholds overlap. This can be useful if there are multiple results being plotted together for comparison, all of which may have different thresholds. The resulting plot can help quickly compare the power of different analyses. Warning - the y axis LOD scale is only correct for the first list element / set of results. Also as before, this function only works for QTL scan over multiple chromosomes.
plotLinearQTL_list

Usage

plotLinearQTL_list(
  LOD_data.ls,
  inter_chm_gap = 5,
  ylimits = NULL,
  sig.unit = "LOD",
  plot_type,
  add_xaxis = TRUE,
  add_rug = TRUE,
  colours = c("black", "red", "dodgerblue", "sienna4"),
  ylab.at = 2.5,
  main.size = 2,
  main.lty = 1,
  thresh.lty = 3,
  thresh.lwd = 2,
  thresh.col = "darkred",
  return_plotData = FALSE,
  highlight_positions = NULL,
  LGdiv.col = "gray42",
  ...
)

Arguments

LOD_data.ls A list, each element of which is a separate output of QTLscan, for which the setting perm_test = TRUE was used each time.
inter_chm_gap The gap size (in cM) between successive chromosomes - by default a gap of 5 cM is used.
ylimits Use to specify ylims of plot region, though by default NULL in which case a suitable plot region is automatically used.
sig.unit Label to use on the y-axis for significance units, by default assumed to be "LOD".
plot_type Plots can be either in line drawings or scatter plot format. If multiple types are required, supply as a vector of same length as LOD_data.ls
add_xaxis Should an x-axis be drawn? If multiple QTL analyses are performed on different traits, specifying this to be FALSE and using par(mar=c(0, 4.1, 4.1, 2.1)) allows subsequent plots to be neatly stacked.
add_rug Logical, by default TRUE - should original marker points be added to plot?
colours Vector of colours to be used in the plotting. A default set of 4 colours is provided.
ylab.at Distance from the y-axis to place label (by default at 2.5 points)
main.size Size of line (or point) to plot the "main" data, the first set of results in the LOD_data.ls input list, by default 2.
main.lty Line type for the "main" data, by default a normal line (lty = 1).
thresh.lty Gives user control over the line type of the significance threshold to be drawn.
thresh.lwd Gives user control over the line width of the significance threshold to be drawn.
**thresh.col**

Gives user control over the line colour of the significance threshold to be drawn, by default "darkred".

**return_plotData**

Logical, by default FALSE. If TRUE, then the x and y coordinates of the plot data are returned, which can be useful for subsequent plot manipulations and overlays.

**highlight_positions**

Option to include a list of associated positions to highlight, of the same length and in the same order as LOD_data. Each set of positions should be provided in data.frame format with 3 columns corresponding to linkage group, type and ID (same format as the cofactor.df argument of function QTLscan). This can be useful if genetic co-factor analyses are being compared. If no position is to be highlighted, add the corresponding list element as NULL.

**LGdiv.col**

Colour of dividing lines between linkage groups, by default grey.

**...**

Arguments passed to lines or points as appropriate (see argument plot_type).

**Value**

The plot data, if return_plotData = TRUE, otherwise NULL.

**Examples**

```r
## Not run:
data("qtl_LODs.4x")
## Introduce some arbitrary noise for the sake of this example:
qtl_LODs.4x_2 <- qtl_LODs.4x
qtl_LODs.4x_2$Perm.res$threshold <- 2.5
qtl_LODs.4x_2$QTL.res$LOD<-qtl_LODs.4x_2$QTL.res$LOD+rnorm(length(qtl_LODs.4x_2$QTL.res$LOD),2)
plotLinearQTL_list(list(qtl_LODs.4x,qtl_LODs.4x_2),plot_type="lines")
## End(Not run)
```

**plotQTL**

*Plot the results of a previous QTL analysis*

**Description**

Basic QTL plotting function, taking map positions and significance levels as input.

**Usage**

```r
plotQTL(
  LOD_data,
  support_interval = 0,
  ylims = NULL,
  multiplot = NULL,
  plot_type = "lines",
)```
plotQTL

overlay = FALSE,
add_xaxis = TRUE,
add_rug = TRUE,
mainTitle = FALSE,
log = NULL,
...
)

Arguments

LOD_data    Output list from QTLscan with items QTL.res and Perm.res (the latter can be NULL)
support_intervalNumeric. If 0 (by default) then there is no support interval returned. If greater than zero, then a LOD support interval is shown on output plot and the bounds are returned.
ylimits Use to specify ylims of plot region, though by default NULL in which case a suitable plot region is automatically used.
multiplot Vector of integers. By default NULL. If LOD_data contains results from multiple linkage groups, you can define the number of rows and columns in the plot layout.
plot_type How should be plots be drawn, either "lines" or "points" are possible
overlay Add to an existing plot (should be produced by a comparable call to this function) or not? By default FALSE, in which case a new plot is drawn. Can be useful for displaying results of multiple analyses together.
add_xaxis Should an x-axis be drawn? If multiple QTL analyses are performed on different traits, specifying this to be FALSE and using par(mar=c(0,4.1,4.1,2.1)) allows subsequent plots to be neatly stacked.
add_rug Logical, by default TRUE - should original marker points be added to plot?
mainTitle Vector of plot titles (single character vector also allowed and will be recycled). For no plot titles, leave as FALSE
log Character string specifying the log filename to which standard output should be written. If NULL log is send to stdout.
... Extra arguments passed to plotting functions (plot, lines / points)

Value

The cM bounds of the LOD support interval, if support_interval > 0.

Examples

data("qtl_LODs.4x")
plotQTL(LOD_data = qtl_LODs.4x,multiplot = c(1,2),ylimits = c(0,5), plot_type = "points")
plotRecLS 

Plot the recombination landscape across the genome

Description

Function which visualises the recombination landscape in two ways: per linkage group, and per individual. For the first analysis, a rudimentary spline is also fitted to estimate the recombination rate along a grid of positions defined by gap, which is also returned by the function.

Usage

plotRecLS(
  recombination_data,
  plot_per_LG = TRUE,
  plot_per_ind = TRUE,
  gap = 1,
  ...
)

Arguments

recombination_data
  Data on predicted recombination events, as returned by the function count_recombinations

plot_per_LG
  Logical argument, plot recombination events per linkage group? By default TRUE.

plot_per_ind
  Logical argument, plot recombination events per individual? By default TRUE.

gap
  The size (in cM) of the gap used to define the grid of positions to define the window in which to estimate recombination rate. By default 1 cM. Interpolated positions are taken to be the centre of an interval, so a 1 cM gap would result in predictions for positions 0.5 cM, 1.5 cM etc.

... Option to pass extra arguments to the plot function for the per_LG plots. This may lead to conflicts with arguments already declared internally (such as main for example).

Value

A list with two elements, per_LG and per_individual. The first of these is itself a list with the same length as recombination_data, giving the estimated recombination rates along the linkage group. This rate is simply estimated as the (weighted) count of recombination breakpoints divided by the population size.

Examples

data("Rec_Data_4x")
plotRecLS(Rec_Data_4x)
polyqtLR

**QTL analysis in polyploid species using identity-by-descent probabilities**

**Description**

R package to perform QTL analysis using marker data from polyploid species.

**probgeno_df_to_array**  
*Convert a progeno_df data.frame to a 3d array*

**Description**

Convert a progeno_df data.frame to a 3d array

**Usage**

```
probgeno_df_to_array(probgeno_df, ploidy)
```

**Arguments**

- **probgeno_df**  
  A data frame as read from the scores file produced by function saveMarkerModels of R package fitPoly, or alternatively, a data frame containing at least the following columns:
  - SampleName : Name of the sample (individual)
  - MarkerName : Name of the marker
  - P0 Probabilities of dosage score ’0’
  - P1, P2... etc. : Probabilities of dosage score ’1’ etc. (up to max offspring dosage, e.g. P4 for tetraploid population)

- **ploidy**  
  Ploidy of the F1 population (can be 2, 3, 4 or 6)

**PVE**  
*Function to determine the percentage variance explained (PVE) of a (maximal) QTL model, and explore sub-models.*

**Description**

This function builds a (maximal) QTL model from previously detected QTL peaks and outputs the percentage variance explained (PVE) of the full QTL model and all sub-models. It uses a similar approach to the fitting of genetic co-factors in the function `QTLscan`. The PVE is very similar to but not exactly equal to the adjusted R2 returned in `QTLscan` at each position (and note: in the former case, these R2 values are per-locus, while this function can estimate the PVE combined over multiple loci). The discrepancy has to do with how PVE is calculated using the formula 100(1 - RSS0/RSS1), where RSS0 and RSS1 are the residual sums of squares of the NULL and QTL models, respectively.
Usage

PVE(
    IBD_list,
    Phenotype.df,
    genotype.ID,
    trait.ID,
    block = NULL,
    QTL_df = NULL,
    prop_Pheno_rep = 0.5,
    log = NULL,
    verbose = FALSE
)

Arguments

  IBD_list         List of IBD probabilities
  Phenotype.df     A data.frame containing phenotypic values
  genotype.ID     The colname of Phenotype.df that contains the offspring identifiers (F1 names)
  trait.ID         The colname of Phenotype.df that contains the response variable to use in the model
  block            The blocking factor to be used, if any (must be colname of Phenotype.df). By default NULL, in which case no blocking structure (for unreplicated experiments)
  QTL_df           A 2-column data frame of previously-detected QTL; column 1 gives linkage group identifiers, column 2 specifies the cM position of the QTL. If not specified, an error results. It can be convenient to generate a compatible data.frame by first running the function check_cofactors to build a multi-QTL model.
  prop_Pheno_rep   The minimum proportion of phenotypes represented across blocks. If less than this, the individual is removed from the analysis. If there is incomplete data, the missing phenotypes are imputed using the mean values across the recorded observations.
  log              Character string specifying the log filename to which standard output should be written. If NULL log is send to stdout.
  verbose          Should messages be written to standard output?

Value

A list with percentage variance explained of maximal QTL model and all sub-models

Examples

data("IBD_4x","Phenotypes_4x")
PVE(IBD_list = IBD_4x,
    Phenotype.df = Phenotypes_4x,
    genotype.ID = "geno", trait.ID = "pheno",
    block = "year",
    QTL_df = data.frame(LG=1,cM=12.3))
**QTLscan**

*General QTL function that allows for co-factors, completely randomised block designs and the possibility to derive LOD thresholds using a permutation test*

---

**Description**

Function to run QTL analysis using IBD probabilities given (possibly replicated) phenotypes, assuming randomised experimental design

**Usage**

```r
QTLscan(
    IBD_list,
    Phenotype.df,
    genotype.ID,
    trait.ID,
    block = NULL,
    folder = NULL,
    filename.short,
    cofactor_df = NULL,
    prop_Pheno_rep = 0.5,
    perm_test = FALSE,
    N_perm.max = 1000,
    alpha = 0.05,
    gamma = 0.05,
    ncores = 1,
    log = NULL,
    verbose = TRUE,
    ...
)
```

**Arguments**

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>IBD_list</td>
<td>List of IBD probabilities</td>
</tr>
<tr>
<td>Phenotype.df</td>
<td>A data.frame containing phenotypic values</td>
</tr>
<tr>
<td>genotype.ID</td>
<td>The colname of Phenotype.df that contains the offspring identifiers (F1 names)</td>
</tr>
<tr>
<td>trait.ID</td>
<td>The colname of Phenotype.df that contains the response variable to use in the model</td>
</tr>
<tr>
<td>block</td>
<td>The blocking factor to be used, if any (must be colname of Phenotype.df). By default NULL, in which case no blocking structure (for unreplicated experiments)</td>
</tr>
<tr>
<td>folder</td>
<td>If markers are to be used as co-factors, the path to the folder in which the imported IBD probabilities is contained can be provided here. By default this is NULL, if files are in working directory.</td>
</tr>
</tbody>
</table>
filename.short If TetraOrigin was used and co-factors are being included, the shortened stem of the filename of the .csv files containing the output of TetraOrigin, i.e. without the tail "_LinkageGroupX_Summary.csv" which is added by default to all output of TetraOrigin.

cofactor_df A 2-column data frame of co-factor(s); column 1 gives linkage group identifiers, column 2 specifies the cM position of the co-factors. By default NULL, in which case no co-factors are included in the analysis.

prop_Pheno_rep The minimum proportion of phenotypes represented across blocks. If less than this, the individual is removed from the analysis. If there is incomplete data, the missing phenotypes are imputed using the mean values across the recorded observations.

perm_test Logical, by default FALSE. If TRUE, a permutation test will be performed to determine a genome-wide significance threshold.

N_perm.max The maximum number of permutations to run if perm_test is TRUE; by default this is 1000.

alpha The P-value to be used in the selection of a threshold if perm_test is TRUE, by default 0.05 (i.e. the 0.95 quantile).

gamma The width of the confidence intervals used around the permutation test threshold using the approach of Nettleton & Doerge (2000), by default 0.05.

ncores Number of cores to use if parallel computing is required. Works both for Windows and UNIX (using doParallel). Use parallel::detectCores() to find out how many cores you have available.

log Character string specifying the log filename to which standard output should be written. If NULL log is send to stdout.

verbose Logical, by default TRUE. Should messages be printed during running?

... Arguments passed to plot

Value

A nested list; each list element (per linkage group) contains the following items:

- QTL.res : Single matrix of QTL results with columns chromosome, position, LOD, adj.r.squared and PVE (percentage variance explained).
- Perm.res : If perm_test = FALSE, this will be NULL. Otherwise, Perm.res contains a list of the results of the permutation test, with list items "quantile","threshold" and "scores". Quantile refers to which quantile of scores was used to determine the threshold. Note that scores are each of the maximal LOD scores across the entire genome scan per permutation, thus returning a genome-wide threshold rather than a chromosome-specific threshold. If the latter is preferred, restricting the IBD_list to a single chromosome and re-running the permutation test will provide the desired threshold.
- Residuals : If a blocking factor or co-factors are used, this is the (named) vector of residuals used as input for the QTL scan. Otherwise, this is the set of (raw) phenotypes used in the QTL scan.
- Map : Original map of genetic marker positions upon which the IBDs were based, most often used for adding rug of marker positions to QTL plots.
- LG_names : Names of the linkage groups
Examples

```r
data("IBD_4x","Phenotypes_4x")
qtl_LODs.4x <- QTLscan(IBD_list = IBD_4x,
                        Phenotype.df = Phenotypes_4x,
                        genotype.ID = "geno",
                        trait.ID = "pheno",
                        block = "year")
```

qtl_LODs.4x  

A list of QTL results (4x)

Description

A list of QTL results (4x)

Usage

qtl_LODs.4x

Format

An object of class list of length 4.

---

quadTM  

Quadrivalent transition matrix function

Description

Quadrivalent transition matrix function

Usage

quadTM(r)

Arguments

r  recombination frequency
Rec_Data_{4x}  

A list of recombination count data (4x)

Description
A list of recombination count data (4x)

Usage
Rec_Data_{4x}

Format
An object of class \texttt{list} of length 2.

rem.hex  

Redundant genotype classes in hexavalent transition matrix (6x)

Description
Redundant genotype classes in hexavalent transition matrix (6x)

Usage
rem.hex

Format
An object of class \texttt{integer} of length 166.

rem.quad  

Redundant genotype classes in quadrivalent transition matrix (4x)

Description
Redundant genotype classes in quadrivalent transition matrix (4x)

Usage
rem.quad

Format
An object of class \texttt{integer} of length 6.
segList_2x

Description
A list of all possible bi-allelic QTL segregation types (2x)

Usage
segList_2x

Format
An object of class list of length 8.

segList_3x

Description
A list of all possible bi-allelic QTL segregation types (3x)

Usage
segList_3x

Format
An object of class list of length 27.

segList_4x

Description
A list of all possible bi-allelic QTL segregation types (4x)

Usage
segList_4x

Format
An object of class list of length 224.
segList_6x  

A list of all possible bi-allelic QTL segregation types (6x)

Description

A list of all possible bi-allelic QTL segregation types (6x)

Usage

segList_6x

Format

An object of class list of length 3735.

segMaker  

Create a list of possible QTL segregation types

Description

Function to generate list of segregation types for the exploreQTL function

Usage

segMaker(ploidy, segtypes, modes = c("a", "d"))

Arguments

ploidy  
The ploidy of the population. Currently assumed to be an even number for this function.

segtypes  
List of QTL segregation types to consider, so e.g. c(1,0) would mean all possible simplex x nullplex QTL (ie. 4 QTL, on each of homologues 1 - 4 of parent 1). Note that symmetrical QTL types that cannot be distinguished are not automatically removed and need to be manually identified. If this is an issue, use the inbuilt list for tetraploids provided with the package to search the full model space. Such an inbuilt list is currently only available for tetraploids, and is available from the exploreQTL function.

modes  
Character vector of modes of QTL action to consider, with options "a" for "additive" and "d" for dominant QTL action.
Run a single marker regression using marker dosages

Description

Function to run a single marker regression using marker dosages

Usage

singleMarkerRegression(
  dosage_matrix,  # An integer matrix with markers in rows and individuals in columns. All markers
  Phenotype.df,   # in this matrix will be tested for association with the trait.
  genotype.ID,    # The colname of Phenotype.df that contains the population identifiers (F1 names)
  trait.ID,       # (must be a colname of Phenotype.df)
  maplist = NULL, # Option to include linkage map in the format returned by MDSMap_from_list
  perm_test = FALSE, # from polymapR. If maplist is not specified (by default NULL) then no ordering of
  N_perm = 1000,   # markers from dosage-matrix is performed. Note that all markers in dosage_matrix
  alpha = 0.05,    # are tested; markers with dosages that were not on the maplist will be assigned
  ncores = 1,      # unordered to linkage group 0 with dummy cM positions 1,2,3 etc.
  return_R2 = FALSE, # Logical, by default FALSE. If TRUE, a permutation test will be performed to
t  log = NULL      # determine a genome-wide significance threshold.
)

Arguments

dosage_matrix  # An integer matrix with markers in rows and individuals in columns. All markers
Phenotype.df   # in this matrix will be tested for association with the trait.
genotype.ID    # The colname of Phenotype.df that contains the population identifiers (F1 names)
trait.ID       # (must be a colname of Phenotype.df)
maplist        # Option to include linkage map in the format returned by MDSMap_from_list
perm_test      # Logical, by default FALSE. If TRUE, a permutation test will be performed to
t               # determine a genome-wide significance threshold.
N_perm         # Integer. The number of permutations to run if perm_test is TRUE; by default
alpha          # Numeric. The P-value to be used in the selection of a threshold if perm_test is

SNP_dosages.4x

ncores  Number of cores to use if parallel processing required. Works both for Windows and UNIX (using doParallel). Use parallel::detectCores() to find out how many cores you have available.

return_R2  Should the (adjusted) R2 of the model fit also be determined?

log  Character string specifying the log filename to which standard output should be written. If NULL log is send to stdout.

Value

A list containing the following components:

- QTL.res : The -log(p) of the model fit per marker are returned as "LOD" scores, although "LOP" would have been a better description. If requested, R2 values are also returned in column "R2adj"
- Perm.res : The results of the permutation test if performed, otherwise NULL
- Map : The linkage map if provided, otherwise NULL

Examples

data("SNP_dosages.4x","BLUEs.pheno")
Trait_1.smr <- singleMarkerRegression(dosage_matrix = SNP_dosages.4x,
Phenotype.df = BLUEs.pheno,genotype.ID = "Geno",trait.ID = "BLUE")

SNP_dosages.4x  A matrix of SNP marker dosages (4x)

Description

A matrix of SNP marker dosages (4x)

Usage

SNP_dosages.4x

Format

An object of class matrix (inherits from array) with 186 rows and 52 columns.
spline_IBD  

Fit splines to IBD probabilities

Description

Fits splines to IBD probabilities at a grid of positions at user-defined spacing.

Usage

spline_IBD(IBD_list, gap, ncores = 1, log = NULL)

Arguments

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>IBD_list</td>
<td>List of IBD probabilities</td>
</tr>
<tr>
<td>gap</td>
<td>The size (in centiMorgans) of the gap between splined positions</td>
</tr>
<tr>
<td>ncores</td>
<td>Number of cores to use, by default 1 only. Works both for Windows and UNIX (using doParallel). Use parallel::detectCores() to find out how many cores you have available. Note that with large datasets, using multiple cores will use large amounts of memory (RAM). Single-core or e.g. 2-core evaluations, although slower, is less memory-intensive.</td>
</tr>
<tr>
<td>log</td>
<td>Character string specifying the log filename to which standard output should be written. If NULL log is send to stdout.</td>
</tr>
</tbody>
</table>

Value

Returns a list of similar format as IBD_list, with a splined IBD_array in place of the original IBD_array

Examples

data("IBD_4x")
IBD_4x.spl <- spline_IBD(IBD_list = IBD_4x, gap = 1)

state_fun  

Function to return the list of possible pairing states, given parental ploidies and meiotic pairing model

Description

Function to return the list of possible pairing states, given parental ploidies and meiotic pairing model

Usage

state_fun(ploidy, ploidy2, bivalent_decoding, full_multivalent_hexa = FALSE)
**Arguments**

- `ploidy` Ploidy of parent 1
- `ploidy2` Ploidy of parent 2
- `bivalent_decoding` Logical, if FALSE then multivalent pairing assumed
- `full_multivalent_hexa` Should multivalent pairing be considered in both parents simultaneously in hexaploids?

**Description**

Error and warning handling for dosage_matrix

**Usage**

test_dosage_matrix(dosage_matrix)

**Arguments**

- `dosage_matrix` An integer matrix with markers in rows and individuals in columns

**Description**

Error and warning handling for IBD_list as estimated by estimate_IBD

**Usage**

test_IBD_list(IBD_list)

**Arguments**

- `IBD_list` List of IBD probabilities
test_probgeno_df

Error and warning handling for probgeno_df data-frame of probabilistic genotypes (scores)

Description

Error and warning handling for probgeno_df data-frame of probabilistic genotypes (scores)

Usage

test_probgeno_df(probgeno_df, ploidy)

Arguments

probgeno_df A data frame as read from the scores file produced by function saveMarkerModels of R package fitPoly, or alternatively, a data frame containing at least the following columns:
• SampleName : Name of the sample (individual)
• MarkerName : Name of the marker
• P0 : Probabilities of dosage score '0'
• P1, P2... etc. : Probabilities of dosage score '1' etc. (up to max offspring dosage, e.g. P4 for tetraploid population)
ploidy Ploidy of the F1 population (can be 2, 3, 4 or 6)

thinmap Thin out map data

Description

thinmap is a function for thinning out an integrated map, in order that IBD estimation runs more quickly. Especially useful for maps with very high marker densities for which the estimate_IBD function is to be used.

Usage

thinmap(
  maplist,
  dosage_matrix,
  bin_size = 1,
  bounds = NULL,
  remove_markers = NULL,
  plot_maps = TRUE,
  parent1 = "P1",
  parent2 = "P2",
  log = NULL
)
Arguments

maplist A list of maps. In the first column marker names and in the second their position.
dosage_matrix An integer matrix with markers in rows and individuals in columns.
bin_size Numeric. Size (in cM) of the bins to include. By default, a bin size of 1 cM is used. Larger bin_size results in fewer markers being left on the resulting map.
bounds Numeric vector. If NULL (by default) then all positions are included, however if specified then output is limited to a specific region, which may be useful if fine-mapping a region of interest.
remove_markers Optional vector of marker names to remove from the maps. Default is NULL.
plot_maps Logical. Plot the marker positions of the selected markers using polymapR::plot_map.
parent1 Identifier of parent 1, by default assumed to be "P1"
parent2 Identifier of parent 2, by default assumed to be "P2"
log Character string specifying the log filename to which standard output should be written. If NULL log is send to stdout.

Value

A maplist of the same structure as the input maplist, but with fewer markers based on the bin_size.

Examples

data("phased_maplist.4x","SNP_dosages.4x")
maplist_thin<-thinmap(maplist=phased_maplist.4x,dosage_matrix=SNP_dosages.4x)

TM.biv.2 Bivalent-pairing transition matrix function, diploid

Description

Bivalent-pairing transition matrix function, diploid

Usage

TM.biv.2(m1, r_vec)

Arguments

m1 marker
r_vec Vector of adjacent recombination frequencies
**TM.biv.4**

*Bivalent-pairing transition matrix function, tetraploid*

**Description**

Bivalent-pairing transition matrix function, tetraploid

**Usage**

`TM.biv.4(m1, r_vect)`

**Arguments**

- `m1` marker
- `r_vect` Vector of adjacent recombination frequencies

**TM.biv.6**

*Bivalent-pairing transition matrix function, hexaploid*

**Description**

Bivalent-pairing transition matrix function, hexaploid

**Usage**

`TM.biv.6(m1, r_vect)`

**Arguments**

- `m1` marker
- `r_vect` Vector of adjacent recombination frequencies
Hexavalent-pairing transition matrix function, hexaploid

Usage

\[ \text{TM.hex}(m1, r\_vect, \text{rem_hex} = \text{rem.hex}) \]

Arguments

- \text{m1} marker
- \text{r_vect} Vector of adjacent recombination frequencies
- \text{rem_hex} Index of genotype classes to remove, work-around from overly-general transition matrix with redundant classes

Quadrivalent-pairing transition matrix function, tetraploid

Usage

\[ \text{TM.quad}(m1, r\_vect, \text{rem_quad} = \text{rem.quad}) \]

Arguments

- \text{m1} marker
- \text{r_vect} Vector of adjacent recombination frequencies
- \text{rem_quad} Index of genotype classes to remove, work-around from overly-general transition matrix with redundant classes
<table>
<thead>
<tr>
<th>Function</th>
<th>Description</th>
<th>Usage</th>
<th>Arguments</th>
</tr>
</thead>
</table>
| `TMfun.2x_B`            | Diploid bi-parental transition matrix                 | `TMfun.2x_B(m1, r_vect)` | `m1` marker
|                        |                                                       |                | `r_vect` Vector of adjacent recombination frequencies |
| `TMfun.3x_BB`           | Triploid bi-parental transition matrix, bivalent-bivalent pairing | `TMfun.3x_BB(...)` | `...` Arguments passed to parental TM functions |
| `TMfun.3x_QB`           | Triploid bi-parental transition matrix, quadrivalent-bivalent pairing | `TMfun.3x_QB(...)` | `...` Arguments passed to parental TM functions |
Tetraploid bi-parental transition matrix, bivalent-bivalent pairing

### Description
Tetraploid bi-parental transition matrix, bivalent-bivalent pairing

### Usage
```
TMfun.4x_BB(...)  
```

### Arguments
```
...  
Arguments passed to parental TM functions  
```

---

Tetraploid bi-parental transition matrix, bivalent-quadrivalent pairing

### Description
Tetraploid bi-parental transition matrix, bivalent-quadrivalent pairing

### Usage
```
TMfun.4x_BQ(...)  
```

### Arguments
```
...  
Arguments passed to parental TM functions  
```

---

Tetraploid bi-parental transition matrix, quadrivalent-bivalent pairing

### Description
Tetraploid bi-parental transition matrix, quadrivalent-bivalent pairing

### Usage
```
TMfun.4x_QB(...)  
```

### Arguments
```
...  
Arguments passed to parental TM functions  
```
**Description**

Tetraploid bi-parental transition matrix, quadrivalent-quadrivalent pairing

**Usage**

`TMfun.4x_QQ(...)`

**Arguments**

... `Arguments passed to parental TM functions`

---

**Description**

Hexaploid bi-parental transition matrix, bivalent-bivalent pairing

**Usage**

`TMfun.6x_BB(...)`

**Arguments**

... `Arguments passed to parental TM functions`

---

**Description**

Hexaploid bi-parental transition matrix, bivalent-hexavalent pairing

**Usage**

`TMfun.6x_BH(...)`

**Arguments**

... `Arguments passed to parental TM functions`
visualiseGIC

<table>
<thead>
<tr>
<th>Function</th>
<th>Description</th>
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<tbody>
<tr>
<td>TMfun.6x_HB</td>
<td>Hexaploid bi-parental transition matrix, hexavalent-bivalent pairing</td>
</tr>
<tr>
<td>TMfun.6x_HH</td>
<td>Hexaploid bi-parental transition matrix, hexavalent-hexavalent pairing</td>
</tr>
<tr>
<td>visualiseGIC</td>
<td>Visualise Genotypic Information Coefficient</td>
</tr>
</tbody>
</table>

**Description**

Hexaploid bi-parental transition matrix, hexavalent-bivalent pairing

**Usage**

```r
TMfun.6x_HB(...) 
```

**Arguments**

... Arguments passed to parental TM functions

Hexaploid bi-parental transition matrix, hexavalent-hexavalent pairing

**Usage**

```r
TMfun.6x_HH(...) 
```

**Arguments**

... Arguments passed to parental TM functions

Function to visualise the GIC of a certain region
visualiseHaplo

Usage

visualiseGIC(
  GIC_list,
  add_rug = TRUE,
  add_leg = FALSE,
  ylimits = NULL,
  gic.cex = 1,
  show_markers = TRUE,
  add.mainTitle = TRUE,
  plot.cols = NULL
)

Arguments

GIC_list List of GIC data, the output of estimate_GIC
add_rug Should original marker positions be added to the plot?
add_leg Should a legend be added to the plot?
ylimits Optional argument to control the plotting area, by default NULL
gic.cex Option to increase the size of the GIC
show_markers Should markers be shown?
add.mainTitle Should a main title be added to the plot?
plot.cols Optional argument to specify plot colours, otherwise suitable contrasting colours are chosen

Value

The phased map data for the specified region, recoded into 1's and 0's.

Examples

data("GIC_4x")
visualiseGIC(GIC_list = GIC_4x)

visualiseHaplo Visualise haplotypes in certain individuals in a certain region

Description

Function to visualise the haplotypes of a certain region in certain individuals
Usage

visualiseHaplo(
    IBD_list,
    display_by = c("phenotype", "name"),
    linkage_group = NULL,
    Phenotype.df = NULL,
    genotype.ID = NULL,
    trait.ID = NULL,
    pheno_range = NULL,
    cM_range = "all",
    highlight_region = NULL,
    select_offspring = NULL,
    recombinant_scan = NULL,
    allele_fish = NULL,
    presence_threshold = 0.95,
    xlabl = TRUE,
    ylabl = TRUE,
    mainTitle = NULL,
    multiplot = NULL,
    append = FALSE,
    colPal = c("white", "navyblue", "darkred"),
    hap.wd = 0.4,
    recombination_data = NULL,
    reset_par = TRUE,
    log = NULL
)

Arguments

IBD_list List of IBD probabilities

display_by Option to display a subset of the population’s haplotypes either by "phenotype" or "name". If "phenotype" is supplied, then Phenotype.df,genotype.ID,trait.ID and pheno_range must also be specified. If "name" is supplied, then select_offspring must be specified.

linkage_group Numeric identifier of the linkage group being examined, based on the order of IBD_list. Only a single linkage group is allowed. If IBD_list corresponds to a single linkage group, default value of NULL will suffice.

Phenotype.df A data.frame containing phenotypic values, which can be used to select a subset of the population to visualise (with extreme phenotypes for example). By default NULL, in which case a subset of the population may be selected using the select_offspring argument.

genotype.ID The colname of Phenotype.df that contains the population identifiers (F1 names) (must be a colname of Phenotype.df)

trait.ID The colname of Phenotype.df that contains the response variable to use in the model (must be a colname of Phenotype.df)

pheno_range Vector of numeric bounds of the phenotypic scores to include (offspring selection).
cM_range  Vector of numeric bounds of the genetic region to be explored. If none are specified, the default of "all" means all cM positions will be included.

highlight_region  Option to highlight a particular genetic region on the plot; can be a single position or a vector of 2 positions. By default NULL.

select_offspring  Vector of offspring identifiers to visualise, must be supplied if display_by = "name". Specifying "all" will result in all offspring haplotypes being visualised.

recombinant_scan  Vector of homologue numbers between which to search for recombinant offspring in the visualised region and selected individuals. By default NULL, in which case no search is preformed.

allele_fish  Vector of homologue numbers of interest, for which to search for offspring that carry these homologues (in the visualised region). By default NULL, in which case no search ("fishing") is performed.

presence_threshold  Numeric. The minimum probability used to declare presence of a homologue in an individual. This is only needed if a recombinant_scan is performed. By default a value of 0.95 is used. When searching for recombinants, this value is also used to denote the proportion of loci carrying the required number of homologues (i.e. by default 95 per cent of loci should have between 0.95 and 1.1 copies of the specified recombinant homologues).

xlabl  Logical, by default TRUE. Should an x-axis label be used?

ylabl  Logical, by default TRUE. Should a y-axis label be used?

mainTitle  Option to override default plot titles with a (vector of) captions. By default NULL.

multiplot  Vector of integers. By default NULL so haplotypes are plotted singly; otherwise a vector specifying the number of rows and columns in the plot layout.

append  Option to allow user to append new plots to spaces generated by multiplot, otherwise these are filled with blank plots. By default FALSE. If TRUE, then a large enough multiplot grid should be generated to make this option meaningful.

colPal  Colour palette to use in the visualisation (best to provide 3 colours).

hap.wd  The width of the haplotype tracks to be plotted, generally recommended to be about 0.4 (default value)

recombination_data  List object as returned by the function count_recombinations. By default NULL, in which case no overlay of predicted recombination events is performed. However, it can be useful to visualise predicted recombination events, particularly as this might help inform the choice of argument plausible_pairing_prob of that function. See count_recombinations for more details.

reset_par  By default TRUE, reset par on exit.

log  Character string specifying the log filename to which standard output should be written. If NULL log is send to stdout.
visualisePairing

Value

If recombinant_scan vector is supplied, a vector of recombinant offspring ID in the region of interest (otherwise NULL).

Examples

data("IBD_4x")
visualiseHaplo(IBD_list = IBD_4x,
    display_by = "name",
    linkage_group = 1,
    select_offspring = "all",
    multiplot = c(3,3))

visualisePairing

Visualise pairing of parental homologues

Description

Function to visualise the pairing of parental homologues across the population using graph, with nodes to denote parental homologues and edges to denote deviations from expected proportions under a polysomic model of inheritance

Usage

visualisePairing(
    meiosis_report.ls,
    pos.col = "red",
    neg.col = "blue",
    parent,
    max.lwd = 20,
    datawidemax,
    add.label = TRUE,
    return.data = FALSE,
    ...
)

Arguments

    meiosis_report.ls
      List output of function meiosis_report
    pos.col
      Colour corresponding to excess of pairing associations predicted (positive deviations), by default red
    neg.col
      Colour corresponding to lack of pairing associations predicted (negative deviations), by default blue
    parent
      The parent, either "P1" (mother) or "P2" (father)
    max.lwd
      Maximum line width, by default 20
datawidemax  This argument is currently a work-around to allow multiple plots to have the same scale (line thicknesses consistent). No default is provided. To estimate this value, simply set argument `return.data = TRUE`, and record the maximum absolute value over columns 'count', which are the deviations from random expectations. This should be done over multiple function calls if e.g. comparing both P1 and P2 values. When a global maximum (absolute) deviation is known, re-run the function with this value for `datawidemax`. The line width specified by `max.lwd` will then be used for this, and all other line widths re-scaled accordingly.

add.label  Should a label be applied, giving the maximum deviation in the plot? By default `TRUE`.

return.data  Should plot data be returned? By default `FALSE`.

...  Optional arguments passed to `plot.igraph`.

Value

If `return.data = TRUE`, the values for pairwise deviations from the expected numbers are returned, useful for determining the value `datawidemax` to provide consistent scaling across multiple plots.

Examples

data("mr.ls")
visualisePairing(meiosis_report.ls = mr.ls,
parent = "P1",
datawidemax = 3)

---

visualiseQTLeffects  Visualise QTL homologue effects around a QTL position

Description

Function to visualise the effect of parental homologues around a QTL peak across the population.

Usage

```
visualiseQTLeffects(
  IBD_list,
  Phenotype.df,
  genotype.ID,
  trait.ID,
  linkage_group,
  LOD_data,
  cM_range = NULL,
  col.pal = c("purple4", "white", "seagreen"),
  point.density = 50,
  zero.sum = FALSE,
  return_plotData = FALSE)
```
Arguments

**IBD_list** List of IBD probabilities

**Phenotype.df** A data.frame containing phenotypic values

**genotype.ID** The colname of Phenotype.df that contains the population identifiers (F1 names) (must be a colname of Phenotype.df)

**trait.ID** The colname of Phenotype.df that contains the response variable to use in the model (must be a colname of Phenotype.df)

**linkage_group** Numeric identifier of the linkage group being tested, based on the order of IBD_list. Only a single linkage group is allowed.

**LOD_data** Output of `QTLscan` function

**cM_range** If required, the plotting region can be restricted to a specified range of centiMorgan positions (provided as a vector of start and end positions).

**col.pal** Vector of colours to use in the visualisations (it is best to provide two or three colours for simplicity). By default, effects will be coloured from purple to green through white.

**point.density** Parameter to increase the smoothing of homologue effect tracks

**zero.sum** How allele substitution effect should be defined. If FALSE (by default), the effect of each homologue is computed relative to the overall phenotypic mean, otherwise contrasts (against offspring without the inherited homologue) are used.

**return_plotData** Logical, by default FALSE. If TRUE, plot data is returned, otherwise NULL.

Value

The estimated effects of the homologues, used in the visualisation

Examples

```r
data("IBD_4x", "BLUEs.pheno", "qtl_LODs.4x")
visualiseQTEffects(IBD_list = IBD_4x,
                   Phenotype.df = BLUEs.pheno,
                   genotype.ID = "Geno",
                   trait.ID = "BLUE",
                   linkage_group = 2,
                   LOD_data = qtl_LODs.4x)
```

weighted.var

*Calculate the weighted variance*

Description

Generalisation of the variance to include weights
Usage

weighted.var(x, w, na.rm = FALSE)

Arguments

x Vector of interest
w Vector of weights
na.rm Should missing values be removed? By default, FALSE

write.logheader  Write a header for the log file

Description

Functionalized writing of function name and arguments as start for log paragraph.

Usage

write.logheader(matc, log)

Arguments

matc A object of class call
log A character string specifying the log file
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